

1 **Original research**

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3 **In Vitro Protoscolicidal Efficacy of Boswellia Resin Extract and Its**  
4 **Nanoemulsion Against Echinococcus granulosus**

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7 **Nima Torabi<sup>1</sup>, Seyed Mohammad Mousavi<sup>2</sup>, Atena Mansouri<sup>3</sup>, Amir Tavakoli kareshk<sup>\*1</sup>**

8 1. Infectious Diseases Research Center, Birjand University of Medical Sciences, Birjand, Iran.

9 Atk9388@gmail.com

10 2. Research Center for Hydatid Disease in Iran, Kerman University of Medical Sciences, Kerman,

11 7616914115, Iran. mosavi.mohammad110@gmail.com

12 3. Cellular and Molecular Research Center, Birjand University of Medical Sciences, Birjand 9717853577,

13 Iran. mansouri\_atena@yahoo.com

14 **Corresponding Author: Amir Tavakoli kareshk**

15 **Email: [atk9388@gmail.com](mailto:atk9388@gmail.com)**

16 **Tel: 00985632381525**

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19 **Abstract**

20 Cystic echinococcosis, caused by *Echinococcus granulosus*, remains a significant zoonotic disease with  
21 limited treatment options, necessitating the exploration of novel therapeutic agents. This study aimed to  
22 evaluate the in vitro protoscolicidal efficacy of Boswellia resin hydroalcoholic extract and its nanoemulsion  
23 formulation against *E. granulosus* protoscoleces. Protoscoleces were obtained from liver cysts of infected  
24 sheep at Birjand slaughterhouse and treated with serial dilutions (0.1%, 0.01%, 0.001%, and 0.0001%) of  
25 both formulations over varying exposure times (5 to 30 minutes). Viability was assessed using 0.1% eosin  
26 staining, and data were analyzed with SPSS22 software using the chi-square test. The hydroalcoholic  
27 extract exhibited protoscolicidal effects only at concentrations above 0.01%, achieving 100% mortality at

28 0.1% after 30 minutes, though effects at lower concentrations were not statistically significant compared to  
29 the control ( $P > 0.05$ ). In contrast, the *Boswellia* nanoemulsion demonstrated significantly superior  
30 protoscolicidal efficacy, achieving 100% mortality at lower concentrations and shorter exposure times (e.g.,  
31 0.1% at 15 minutes and 0.01% at 20 minutes), with statistical significance confirmed at these levels ( $P <$   
32  $0.05$ ). These findings highlight the potential of *Boswellia* nanoemulsion as a promising natural agent for  
33 hydatid cyst treatment due to enhanced bioavailability and efficacy compared to the extract alone.

34  
35 **Keywords:** *Echinococcus granulosus*, *Boswellia* resin, nanoemulsion, protoscolicidal, in vitro  
36

### 37 **1.Introduction:**

38 *Echinococcus granulosus* is a parasitic cestode responsible for causing cystic echinococcosis (CE),  
39 commonly known as hydatid disease, which is a significant zoonotic infection worldwide. The parasite's  
40 life cycle involves canids, primarily domestic dogs, as definitive hosts, and ungulates like sheep as  
41 intermediate hosts. Humans become accidental intermediate hosts by ingesting *E. granulosus* eggs that are  
42 shed in the feces of infected dogs and subsequently contaminate food or water sources (1,2). The disease is  
43 globally prevalent, especially in regions where livestock farming and close contact with dogs are common,  
44 such as South America, Africa, the Middle East, and Central Asia. CE poses a substantial public health  
45 concern due to its chronic nature and potential to cause severe organ damage. Economically, it burdens the  
46 livestock industry by causing decreased productivity and condemnation of infected organs, leading to  
47 significant financial losses (3,4). Conventional treatments for CE, such as surgical excision and  
48 chemotherapy with benzimidazole drugs like albendazole and mebendazole, face significant challenges,  
49 including risks of protoscolex spillage leading to recurrence, severe immunological reactions, and toxic  
50 side effects like liver toxicity and bone marrow suppression, limiting their applicability. (5)(6). These  
51 limitations highlight the pressing necessity for alternative treatment options that are more efficient and  
52 result in fewer negative effects. Historically, medicinal plants have played a crucial role in treating parasitic  
53 infections due to their accessibility, cost-effectiveness, and lower toxicity (7). The investigation of natural  
54 products for antiparasitic treatment is, thus, a promising path to create safer and more efficient therapies.

55 The *Boswellia* species, commonly known as frankincense, are part of the Burseraceae family and originate  
56 from the arid and semi-arid regions of the Arabian Peninsula, India, and East Africa (8). The resin derived  
57 from *Boswellia* is rich in phytochemicals, notably boswellic acids, which are considered the primary active  
58 constituents responsible for its well-documented anti-inflammatory and immunomodulatory effects (9).  
59 Traditionally, *Boswellia* extracts have been employed for various medicinal purposes (10). While previous  
60 research has highlighted diverse pharmacological activities of *Boswellia*, including anticancer,  
61 antimicrobial, and antiviral properties (9,11), and some studies have suggested potential antiparasitic effects  
62 against other organisms (Reff), its specific efficacy against *E. granulosus* protoscoleces, particularly when  
63 formulated to enhance bioavailability, remains largely unexplored. This represents a significant gap this  
64 study aims to address. Nanoemulsions represent an innovative drug delivery platform, characterized by  
65 thermodynamically stable, colloidal dispersions of nanoscale droplets (typically 10-200 nm). Their unique  
66 physicochemical properties, including high surface area-to-volume ratio and enhanced stability,  
67 significantly improve the solubility and bioavailability of poorly water-soluble compounds (12). This  
68 nanotechnology offers distinct advantages over conventional formulations by facilitating better absorption,  
69 potentially enabling targeted delivery, and protecting active compounds from degradation, thereby  
70 maximizing therapeutic efficacy (13) (14). The application of nanoemulsion technology to enhance the  
71 delivery of plant-derived compounds like those found in *Boswellia* resin is particularly promising for  
72 improving treatment outcomes in parasitic diseases.

73 This study represents a pioneering effort to evaluate the protoscolicidal efficacy of *Boswellia* resin extract  
74 in a nanoemulsion formulation against *E. granulosus* protoscoleces, an approach that has not been  
75 previously explored in the context of hydatid disease treatment. To our knowledge, this is the first  
76 investigation into the potential of a *Boswellia* nanoemulsion for hydatid disease treatment. We hypothesize  
77 that the nanoemulsion formulation of *Boswellia* resin extract will exhibit superior protoscolicidal efficacy  
78 against *E. granulosus* protoscoleces compared to the hydroalcoholic extract alone, due to enhanced  
79 solubility, bioavailability, and targeted delivery of active compounds like boswellic acids.

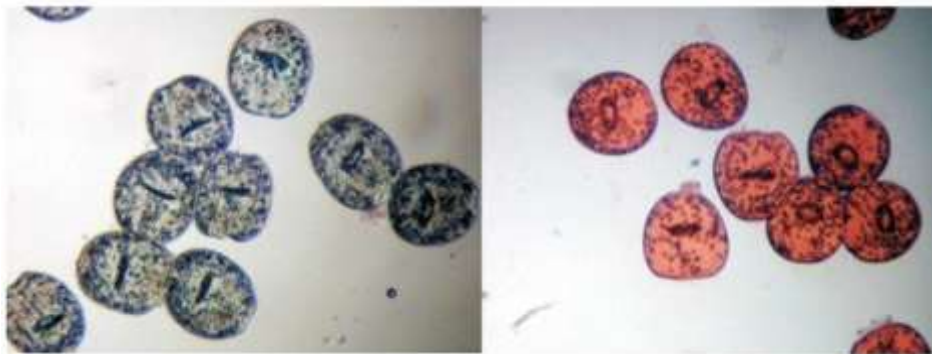
## 80 2. Materials & Method

### 81 2.1. Collection and Preparation of Protoscoleces

82 In this experimental in vitro study, protoscoleces of *E. granulosus* were obtained from liver hydatid cysts  
83 of infected sheep collected from the Birjand slaughterhouse. The hydatid cysts were identified and  
84 aseptically opened in the laboratory to aspirate protoscoleces under sterile conditions, followed by washing  
85 at least three times with sterile normal saline to remove debris. The viability of the protoscoleces was  
86 assessed using the 0.1% eosin staining method (15), counting at least 100 protoscoleces, and only samples  
87 with viability over 90% were used in the experiments (**Figure 1**).

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90

91 **Figure 1.** Microscopic evaluation of *E. granulosus* protoscoleces viability using eosin staining.

92 Left panel: Viable protoscoleces excluding eosin stain, appearing unstained with intact  
93 morphological features. Right panel: Non-viable protoscoleces after treatment with Boswellia  
94 nanoemulsion, showing eosin uptake (red staining) indicating loss of membrane integrity and death  
95 (400× magnification).

96

## 97 **2.2. Preparation of Boswellia Extract**

98 High-quality yellow resin of *Boswellia* was purchased, and its identity was confirmed by  
99 morphological comparison with authenticated specimens at the Herbarium of the School of  
100 Pharmacy, Birjand University of Medical Sciences; impurities were removed before use. For the  
101 preparation of the hydroalcoholic extract, 300 grams of powdered *Boswellia* resin were mixed with 100 mL  
102 of 70% ethanol and distilled water, gently stirred to facilitate extraction, filtered to obtain the initial extract,  
103 and then the solvent was evaporated using a rotary evaporator at 80°C to concentrate the extract.

## 104 **2.3. Preparation of Boswellia Nanoemulsion**

105 The Boswellia nanoemulsion was prepared using the spontaneous emulsification method by testing various  
106 ratios of emulsifiers and the extract to achieve a stable nanoemulsion, employing co-solvents like propylene  
107 glycol and polyethylene glycol. The selection of specific surfactants (optimized empirically through ratio  
108 testing) and co-solvents like propylene glycol and polyethylene glycol was based on their established roles  
109 in reducing interfacial tension, enhancing the solubility of hydrophobic compounds like those in Boswellia  
110 resin, their common use in pharmaceutical formulations due to favorable safety profiles, and their ability to  
111 contribute to the formation of stable nano-scale emulsions. The aqueous phase was slowly added to the oil  
112 phase under constant stirring. The spontaneous emulsification method was chosen as an initial low-energy  
113 approach to form a coarse emulsion, followed by high-speed homogenization using an ultrasonicator for 20  
114 minutes. This high-energy step is crucial for reducing droplet size to the nano-range (as confirmed by DLS),  
115 ensuring homogeneity, and enhancing the kinetic stability of the final nanoemulsion formulation, a common  
116 and effective strategy for preparing stable nanoemulsions. This process yielded a transparent, single-phase  
117 nanoemulsion. A nanoemulsion gel was formulated by hydrating carbomer polymer in distilled water,  
118 adjusting the pH to 6.8 using triethanolamine, and incorporating the nanoemulsion into the gel base until  
119 homogeneous.

## 120 **2.4. Characterization of Nanoemulsion**

121 Characterization of the nanoemulsion involved particle size analysis using Dynamic Light Scattering  
122 (DLS), which confirmed an optimal nanoscale dimension of approximately 33.7 nm, morphological  
123 examination using Transmission Electron Microscopy (TEM) to observe nano-sized particles, and Fourier  
124 Transform Infrared Spectroscopy (FTIR) analysis to identify functional groups and confirm the chemical  
125 integrity of the extract and nanoemulsion. These characterizations confirmed the successful formation and  
126 initial physicochemical properties of the nanoemulsion prior to its use in the protoscolicidal assays.

## 127 **2.5. Preparation of Test Solutions**

128 Test solutions were prepared by making a 0.1% (w/v) stock solution of the *Boswellia* extract with the  
129 addition of Tween 20 to enhance solubility, and serial dilutions were performed to obtain concentrations of  
130 0.1%, 0.01%, 0.001%, and 0.0001% for both the extract and nanoemulsion formulations.

## 131 **2.6. In Vitro Protoscolicidal Assay**

132 In the in vitro protoscolicidal assay, protoscoleces were treated with different concentrations of the  
133 hydroalcoholic extract and nanoemulsion of *Boswellia*, while control groups included a positive control  
134 using 1% silver nitrate solution and a negative control using sterile normal saline. Equal volumes of  
135 protoscolex suspension and test solutions were mixed and incubated for various time intervals (5, 10, 15,  
136 20, 25, and 30 minutes) in 48-well plates, with each concentration and time point tested in triplicate to  
137 ensure accuracy (20,21).

## 138 **2.7. Assessment of Protoscolicidal Activity**

139 Protoscolicidal activity was assessed by viability testing post-incubation, where samples were stained with  
140 0.1% eosin solution and observed under a light microscope; dead protoscoleces absorbed the stain  
141 (appearing red), while live ones excluded it, and at least 100 protoscoleces were counted per sample to  
142 determine percent mortality (22,23).

## 143 **2.8. Data Analysis**

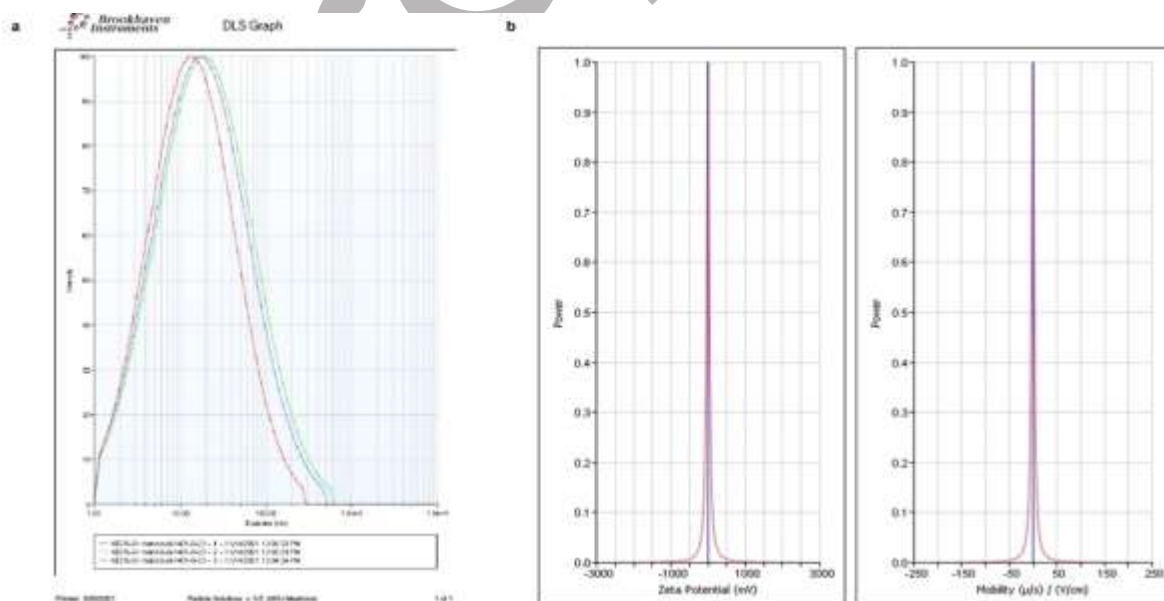
144 Data analysis was performed using SPSS software version 22. The chi-square test was employed to compare  
145 mortality rates (proportions of dead protoscolecids) between different treatment groups (extract vs.  
146 nanoemulsion vs. controls) and concentrations at each specific time point. Assumptions for the chi-square  
147 test, including categorical data type (live/dead), independence of observations, and expected cell  
148 frequencies, were verified prior to analysis. P-values less than 0.05 were considered statistically significant,  
149 indicating a significant difference in mortality proportions compared to the negative control group under  
150 the specified conditions.

151

### 152 3. Results:

153 The particle size analysis of the *Boswellia* nanoemulsion was performed using Dynamic Light Scattering  
154 (DLS), yielding an average particle size of approximately 35.07 nanometers (**Figure 2a**). Additionally, the  
155 zeta potential measurement showed that the nanoemulsion particles had a surface charge of -3.68 mV  
156 (**Figure 2b**).

157



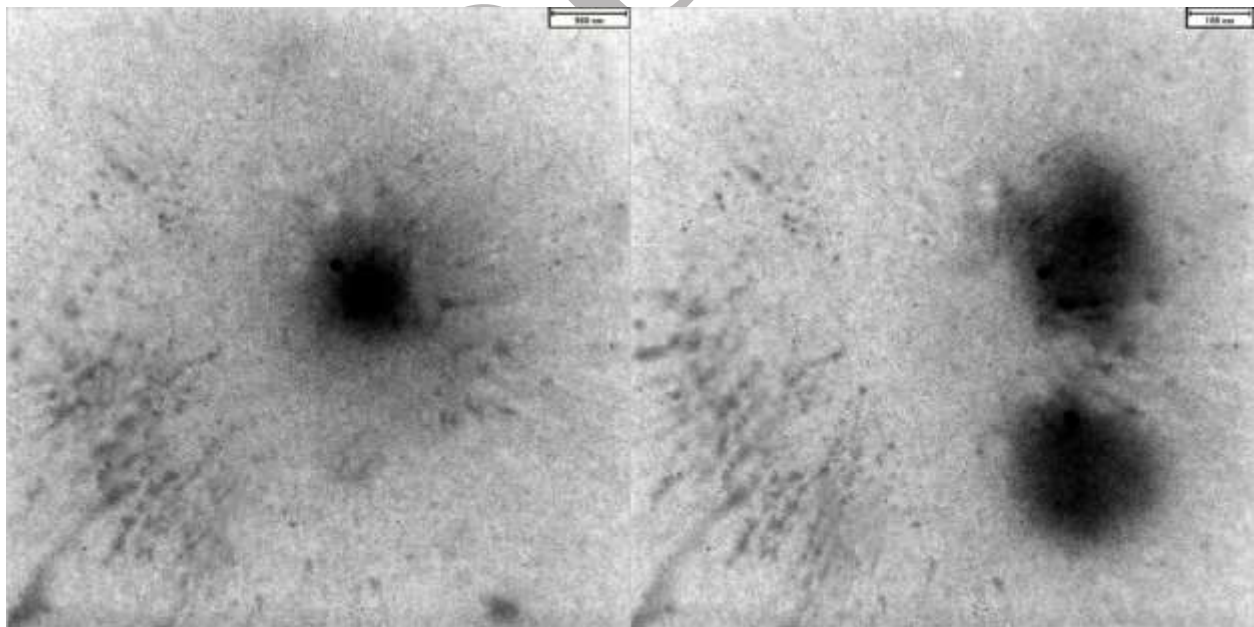
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159 **Figure 2.** Characterization of Boswellia resin nanoemulsion. (a) Dynamic Light Scattering (DLS)  
160 analysis showing particle size distribution. (b) Zeta potential measurements indicating the surface  
161 charge and mobility of the nanoemulsion particles.

162

163

164 The morphological examination of the Boswellia nanoemulsion was conducted using Transmission  
165 Electron Microscopy (TEM) to visualize the nanoparticles and confirm their morphology and size  
166 distribution. TEM imaging revealed that the nanoemulsion particles were spherical and uniformly  
167 distributed, consistent with the nanoscale size determined by Dynamic Light Scattering (DLS) analysis  
168 (approximately 35.07 nanometers). The representative TEM images showcased well-dispersed particles  
169 with consistent spherical shapes, indicating successful formulation of the nanoemulsion and homogeneity  
170 of the particle size (**Figure 3**).



171

172 **Figure 3.** Transmission electron microscopy (TEM) images of Boswellia resin nanoemulsion  
173 showing spherical nanoparticles. TEM micrograph at 900 nm and 100 nm scale bar demonstrating

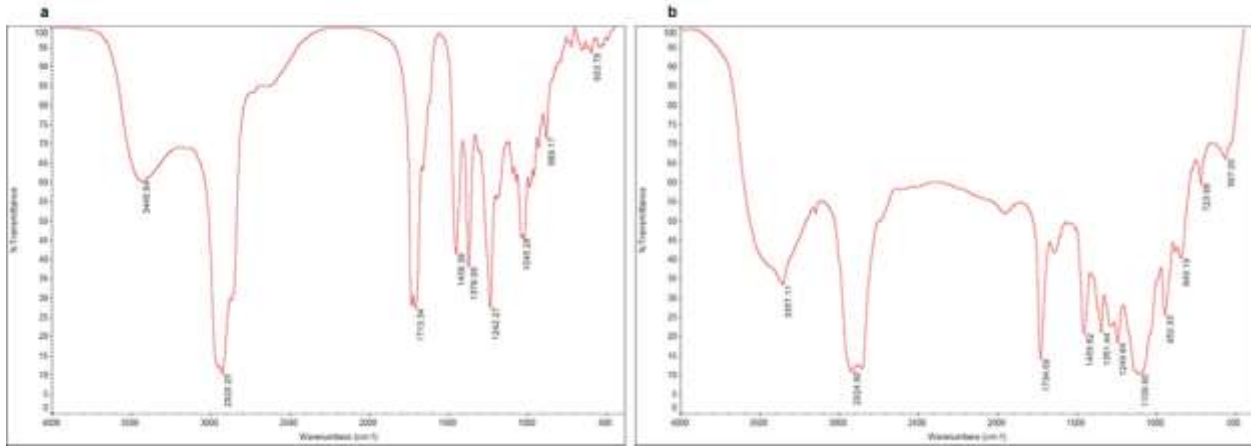


174 multiple uniform spherical particles, confirming the successful formulation of the nanoemulsion  
175 with consistent morphology.

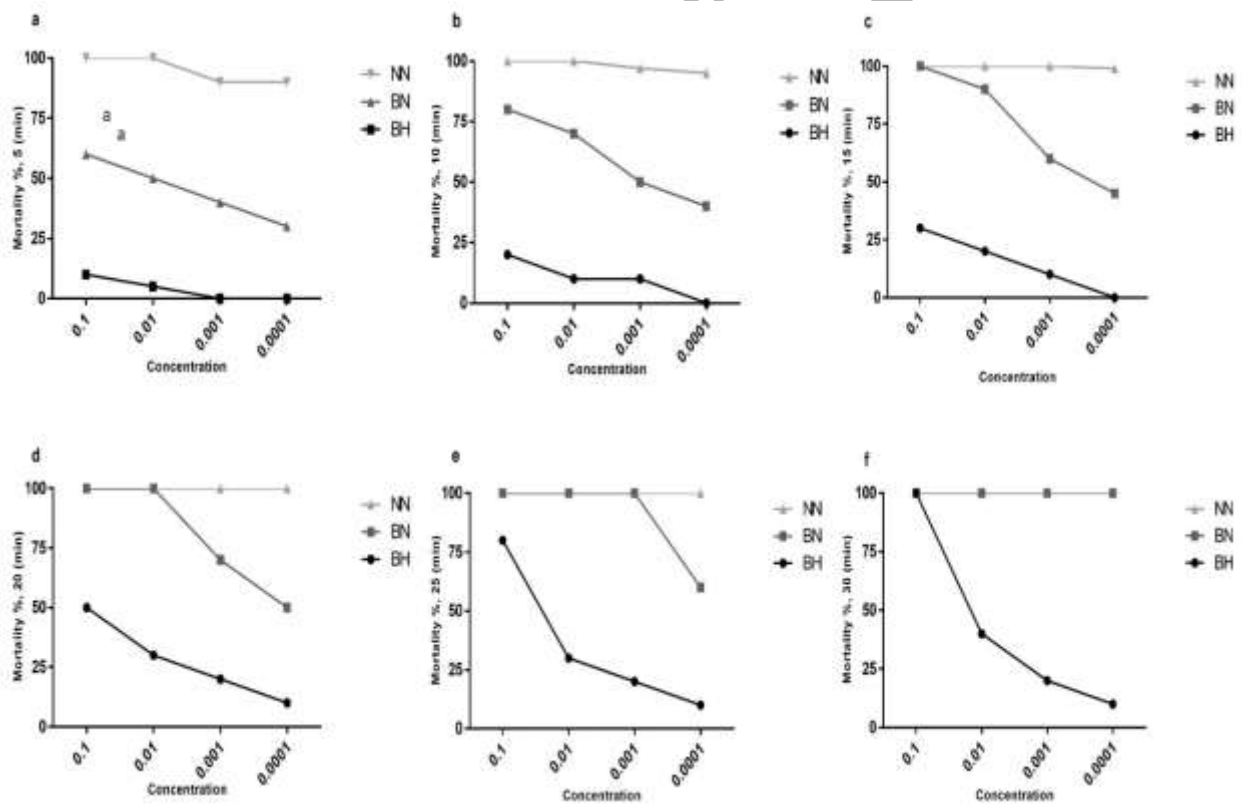
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177 FTIR spectroscopic analysis was conducted to investigate the chemical composition and molecular  
178 interactions in both the hydroalcoholic extract and the nanoemulsion formulations of *Boswellia*. The FTIR  
179 spectra of both formulations exhibited characteristic absorption bands at  $3440\text{ cm}^{-1}$ , corresponding to O–  
180 H stretching vibrations of hydroxyl groups such as phenols and alcohols;  $2929\text{ cm}^{-1}$ , attributed to C–H  
181 stretching vibrations of aliphatic hydrocarbons;  $1713\text{ cm}^{-1}$ , assigned to C=O stretching vibrations  
182 indicative of carbonyl groups like ketones, aldehydes, carboxylic acids, or esters;  $1456\text{--}1378\text{ cm}^{-1}$ , due to  
183 C–H bending vibrations confirming the presence of methyl and methylene groups; and  $1242\text{ cm}^{-1}$ ,  
184 associated with C–O stretching vibrations of ethers, esters, or carboxylic acids (**Figure 4a**). In the  
185 nanoemulsion spectra, additional peaks were observed at  $3500\text{ cm}^{-1}$ , related to amide groups, proteins,  
186 enzymes, and phenolic O–H groups;  $1380\text{ cm}^{-1}$ , representing the NO<sub>2</sub> band of nitro compounds; and  
187  $1045\text{ cm}^{-1}$ , associated with C–F bonds in aliphatic fluoro compounds (**Figure 4b**). The presence of these  
188 signature peaks in both the extract and the nanoemulsion formulations confirms the successful incorporation  
189 of the *Boswellia* extract components into the nanoemulsion while maintaining their chemical integrity. The  
190 protoscolicidal activity of both the *Boswellia* hydroalcoholic extract and its nanoemulsion formulation was  
191 evaluated against *Echinococcus granulosus* protoscoleces at varying concentrations (0.1%, 0.01%, 0.001%,  
192 0.0001%) and time intervals (5, 10, 15, 20, 25, 30 minutes) (**Figure 5**). The hydroalcoholic extract exhibited  
193 significant protoscolicidal effects only at concentrations higher than 0.01%, with mortality rates increasing  
194 over time and with higher concentrations. Specifically, at a concentration of 0.1%, the extract achieved  
195 100% mortality of protoscoleces after 30 minutes of exposure, identifying it as the minimum effective  
196 concentration for significant activity. Statistical comparisons with the negative control (sterile normal  
197 saline) showed that the extract's effects at lower concentrations (0.001% and 0.0001%) were not statistically  
198 significant (**P > 0.05**) across the tested time points.

199



200 **Figure 4.** FTIR spectra of Boswellia formulations. (a) FTIR spectrum of Boswellia hydroalcoholic  
201 (b) FTIR spectrum of Boswellia nanoemulsion gel



202

203 **Figure 5.** The mortality rates of protozoa were assessed after exposure to silver nitrate (SN),  
204 Boswellia nanoemulsion (BN), and Boswellia hydroalcoholic extract (BH) at concentrations

205 ranging from 0.1% to 0.0001% for (a) 5 minutes, (b) 10 minutes, (c) 15 minutes, (d) 20 minutes,  
206 (e) 25 minutes, and (f) 30 minutes.

207

208

209 In contrast, the *Boswellia* nanoemulsion demonstrated markedly enhanced protoscolicidal activity,  
210 achieving higher mortality rates at lower concentrations and significantly shorter exposure times compared  
211 to the extract alone. Remarkably, the nanoemulsion achieved 100% mortality at a concentration of 0.1%  
212 within just 15 minutes ( $P < 0.05$ ) and at a lower concentration of 0.01% within 20 minutes ( $P < 0.05$ ). These  
213 mortality rates were statistically significant compared to the negative control group at these time points and  
214 concentrations. Control experiments validated these findings, where the positive control (1% silver nitrate  
215 solution) demonstrated rapid and complete protoscolicidal activity as expected, confirming the assay's  
216 validity, while the negative control showed negligible mortality.

#### 217 **4. Discussion**

218 The present study investigated the protoscolicidal effects of hydroalcoholic extract of *Boswellia*  
219 (*frankincense*) and its nanoemulsion formulation against *E. granulosus* protoscoleces in vitro. The findings  
220 demonstrated that the nanoemulsion form of *Boswellia* exhibited enhanced protoscolicidal activity  
221 compared to the hydroalcoholic extract alone. Specifically, the nanoemulsion achieved 100% mortality of  
222 protoscoleces at a concentration of 0.1% within 15 minutes and at 0.01% within 20 minutes ( $P < 0.05$ ). In  
223 contrast, the hydroalcoholic extract required a higher concentration (0.1%) and longer exposure time (30  
224 minutes) to achieve similar efficacy ( $P > 0.05$ ). These results suggest that the nanoemulsion formulation  
225 significantly improves the delivery and bioavailability of the active compounds in *Boswellia*, particularly  
226 boswellic acids, which are known for their anti-inflammatory and antimicrobial properties (10). The  
227 nanoscale size of the emulsion particles (~35 nanometers) likely contributes to increased surface area and  
228 enhanced interaction with the protoscoleces, leading to more effective penetration and disruption of the

229 parasite's cellular structures (16,17). The enhanced efficacy of the *Boswellia* resin nanoemulsion can be  
230 attributed to several potential mechanisms related to its physicochemical properties. Nanoemulsions are  
231 known to increase the solubility and bioavailability of hydrophobic compounds like boswellic acids, the  
232 primary active constituents of *Boswellia* resin (12). The nanoscale droplets (~33.7 nm) provide a larger  
233 surface area for interaction with protoscolecocytes, facilitating more efficient delivery and absorption of the  
234 active compounds (18). Furthermore, the small droplet size allows for enhanced permeation through  
235 biological membranes, potentially leading to greater penetration into the parasite's tegument and  
236 intracellular spaces. The surfactants used in the nanoemulsion may also disrupt the membrane integrity of  
237 protoscolecocytes, contributing to increased mortality. Additionally, nanoemulsions can protect the active  
238 compounds from degradation, maintaining their stability and prolonging their activity during the treatment  
239 period (25). Previous studies have highlighted the antimicrobial and antiparasitic properties of *Boswellia*.  
240 For instance, Al-Harrasi and Al-Saidi (8) reported the presence of bioactive triterpenoids in *Boswellia*  
241 species, which exhibit significant antimicrobial activity. Moreover, Mohammadi et al. (25) demonstrated  
242 the antifungal effects of *Boswellia* essential oil against clinical isolates of *Candida albicans*, indicating its  
243 potential in combating fungal infections. The enhanced efficacy of the nanoemulsion observed in this study  
244 aligns with other research emphasizing the benefits of nanoparticle formulations in drug delivery.  
245 Nanoemulsions can improve the solubility of hydrophobic compounds, protect active ingredients from  
246 degradation, and facilitate targeted delivery to pathogens (25). This is particularly relevant for hydrophobic  
247 compounds like boswellic acids, where conventional formulations may have limited efficacy due to poor  
248 solubility and bioavailability. The results of the FTIR spectroscopic analysis showed characteristic peaks at  
249 ~3400  $\text{cm}^{-1}$  (O–H stretching), ~1730  $\text{cm}^{-1}$  (C=O stretching), and ~2920  $\text{cm}^{-1}$  (C–H stretching), similar  
250 to those observed in the pure extract. The absence of significant peak shifts or new peaks suggests that the  
251 chemical structure of the extract was preserved during formulation. The preservation of characteristic  
252 absorption bands corresponding to key functional groups—such as the O–H stretching vibrations at  
253 3440  $\text{cm}^{-1}$ , C–H stretching at 2929  $\text{cm}^{-1}$ , C=O stretching at 1713  $\text{cm}^{-1}$ , C–H bending at 1456–  
254 1378  $\text{cm}^{-1}$ , and C–O stretching at 1242  $\text{cm}^{-1}$ —in both the extract and nanoemulsion spectra indicates that

255 the fundamental molecular structures responsible for the extract's therapeutic properties remain intact. The  
256 additional peaks observed in the nanoemulsion spectra, including the amide groups and phenolic O–H at  
257 3500 cm<sup>-1</sup>, NO bands at 1380 cm<sup>-1</sup>, and C–F bonds at 1045 cm<sup>-1</sup>, suggest successful interaction and  
258 stabilization of the extract within the nanoemulsion matrix. This is a critical finding, as maintaining the  
259 structural integrity of the extract's functional groups is essential for preserving its biological activity and  
260 therapeutic efficacy. Therefore, the nanoemulsion formulated in this study is validated as an effective  
261 delivery system for the *Boswellia* extract, potentially enhancing its stability, bioavailability, and overall  
262 efficacy in pharmaceutical applications. Comparative studies with other plant extracts and nanoformulations  
263 further support the potential of using nanoemulsions for antiparasitic purposes. For example,  
264 Mahmoudvand et al. (20) found that plant extracts formulated in nanoparticle forms exhibited greater  
265 protoscolicidal activity against *E. granulosus* compared to their crude extracts. Similarly, research on other  
266 nanoparticles, such as zinc oxide nanoparticles synthesized with plant extracts, showed significant  
267 antiparasitic effects. The use of positive (1% silver nitrate) and negative (sterile normal saline) controls in  
268 this study validated the experimental conditions and ensured that the observed protoscolicidal effects were  
269 attributable to the *Boswellia* formulations. The negative control exhibited no significant protoscolicidal  
270 activity, confirming that the mortality rates observed were due to the treatments applied. Statistical analysis  
271 reinforced the significance of the findings, with mortality rates showing a clear dependency on both  
272 concentration and exposure time. The nanoemulsion's superior performance suggests that it could be a  
273 promising candidate for developing new protoscolicidal agents with potential applications in the treatment  
274 of hydatid cyst disease. Despite the promising in vitro results, several limitations must be acknowledged.  
275 Firstly, the study's in vitro design cannot fully replicate the complex biological environment of a host.  
276 Factors such as host immune responses, drug metabolism, and interactions with host tissues are absent in  
277 this model but would significantly influence efficacy and safety in vivo. Secondly, potential variability in  
278 protoscolex sensitivity due to genetic differences between *E. granulosus* strains or variations in  
279 developmental stages was not assessed and could impact the generalizability of these findings. Thirdly, and  
280 critically for therapeutic potential, this study did not evaluate the cytotoxicity of the *Boswellia* extract or

281 nanoemulsion against host cells. Assessing potential toxicity to relevant cells (e.g., hepatocytes, fibroblasts,  
282 immune cells) is essential before considering in vivo applications. Finally, the long-term stability of the  
283 prepared nanoemulsion under different storage conditions was not investigated, which is crucial for  
284 practical formulation development. Building upon these promising preliminary findings, future research  
285 should focus on addressing the identified limitations. Crucially, comprehensive in vitro cytotoxicity studies  
286 are required to determine the selectivity index of the *Boswellia* nanoemulsion (i.e., toxicity to parasites vs.  
287 host cells). Subsequent in vivo studies in appropriate animal models of cystic echinococcosis are essential  
288 to evaluate the nanoemulsion's efficacy, pharmacokinetics (absorption, distribution, metabolism, excretion),  
289 and safety profile, including potential organ toxicity, following relevant administration routes. Further  
290 research could also explore the precise molecular mechanisms underlying the enhanced protoscolicidal  
291 activity, potentially involving investigations into membrane disruption, metabolic interference, or apoptosis  
292 induction in the protoscoleces. Optimizing the nanoemulsion formulation for stability and potential targeted  
293 delivery could also enhance its therapeutic prospects.

294 In conclusion, the nanoemulsion formulation of *Boswellia* hydroalcoholic extract significantly enhances its  
295 protoscolicidal activity against *E. granulosus* protoscoleces in vitro. This enhancement can be attributed to  
296 the improved solubility and bioavailability of active compounds in the nanoemulsion. The results support  
297 the potential use of *Boswellia* nanoemulsion as an effective and natural protoscolicidal agent, which could  
298 contribute to safer and more efficient treatments for hydatid cyst disease. Further research, including in  
299 vivo studies and clinical trials, is warranted to fully realize its therapeutic potential.

### 300 **Abbreviations**

301 The following abbreviations are used in this manuscript:

302 **CE:** Cystic echinococcosis

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305 the parasitology laboratory in university of medical sciences, Birjand, Iran.(code: 456990)

306 **Data Availability:**

307 The dataset presented in the study is available on request from the corresponding author during  
308 submission or after publication.

309 **Ethical Approval:**

310 This study was approved under the ethical approval code [IR.BUMS.REC.1402.077](#).

311 **Conflict of interest**

312 The authors have no competing interests to declare that are relevant to the content of this article.

313 **Authors Contribution**

314 Study concept and design: **TN** and **ATK**. Analysis and interpretation of data: **SMM**. Drafting of the  
315 manuscript: **ATK**. Critical revision of the manuscript for important intellectual content: **AM**. Statistical  
316 analysis: **AM**.

317

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