

Malignant Edema in Some Sheep Flocks of Iran

Abstract

Malignant edema is a severe and swiftly fatal disease affecting domestic and wild livestock. The disease manifests following the introduction of *Clostridium* spp. into wounds or skin damage. *Clostridium septicum* is commonly linked with malignant edema. This disease, characterized by edema, doughy swelling, and skin necrosis, is underreported in Iran, leading to a lack of awareness among clinicians. Given its underreporting, addressing this issue is essential, prompting current research efforts to understand better its prognosis, bacteriological and molecular diagnosis, clinical signs, and treatment. Upon detecting suspicious signs of malignant edema in three separate flocks with imported breeds, investigations ensued, including regular clinical exams and sample collection from subcutaneous tissue. The impacted livestock consists of five Île-de-France sheep and two Romane rams, with one Île-de-France ram succumbing to the disease. The bacteriological procedure, including Gram staining and isolation of the causative agent, was meticulously carried out using the standard method. The PCR assay was conducted to validate the existence of *C. septicum* and reject the presence of *Clostridium chauvoei* by employing specific primers. The diagnosis of malignant edema in the affected sheep was confirmed through clinical, macroscopic, and bacteriological examinations, all of which corroborated the presence of *C. septicum*. The PCR assay demonstrated the presence of the *C. septicum*, verifying the bacteriological procedure. Initial signs of the infection included depression, weakness, high fever, and colic, followed by regional pain, crepitation, swelling characterized by a doughy consistency, edema, pain, and necrosis. The study highlights the potential for preventing malignant edema-related fatalities through early diagnosis and antibiotic intervention (Penicillin and Streptomycin). However, it notes a persistent challenge: the inability to repair necrotic tissue at the lesion site. Malignant edema, not being a prominently warned disease and with vaccinations available against its causative agent, has received comparatively less focus from clinicians and researchers in Iran.

Keywords: Malignant edema, *Clostridium septicum*, Sheep, Crepitation, Gas gangrene.

26 1. Introduction

27 Malignant edema (gas gangrene) is an acute and often fatal disease affecting both domestic
28 and wild animals, resulting from contamination of wounds by one or more members of the "gas
29 gangrene" group of clostridial organisms. One potential explanation for its occurrence is that the
30 causative bacteria are soil-borne organisms, gaining entry into livestock primarily through grazing
31 on low-lying, wet pastures. This disease is common in cattle and sheep but infrequently in goats,
32 with young rams most commonly affected. It is hypothesized that the nature of the fleece and the
33 relatively sparse presence of skin folds and body pleats in goats result in fewer shearing wounds,
34 thereby leading to less secondary invasion of clostridial spores (1-5).

35 *Clostridium chauvoei*, *Clostridium septicum*, *Clostridium sordelli*, and *Clostridium novyi* are
36 rod-shaped, Gram-positive, toxin-producing, spore-forming, ubiquitous, and anaerobic bacteria.
37 They are commonly found in soil and can persist for extended periods due to sporulation.
38 Additionally, these bacteria may inhabit healthy livestock's intestinal tract and liver without
39 causing disease. "False" blackleg, more accurately known as malignant edema, is primarily caused
40 by *C. septicum*, although *C. novyi*, *C. sordelli*, *C. chauvoei*, and even *Clostridium perfringens* have
41 been isolated from lesions characteristic of malignant edema (1, 6, 7).

42 Malignant edema can develop when spores enter tissues through penetrating wounds, which
43 commonly arise during routine management practices like vaccinations, blood sampling,
44 parturition, shearing, docking, castration, disbudding, and particularly shearing activities (1, 2, 5,
45 8). Fighting and head-butting among bucks can also facilitate the introduction of spores, leading
46 to a specific type of malignant edema known as "big head." The tissue damage associated with
47 wounds creates anaerobic conditions that promote bacterial proliferation. As anaerobic conditions
48 develop in body tissues, the organisms multiply and release several exotoxins that have local and
49 systemic effects. These toxins induce severe localized inflammation and generalized, fatal
50 toxemia. Local inflammation typically manifests as tissue necrosis accompanied by the
51 accumulation of edema and gas. Subcutaneous and connective tissue are often more prominently
52 affected than muscle tissue, although muscle involvement can occur (1, 2).

53 The disease typically occurs sporadically but can also manifest as outbreaks. For instance,
54 an outbreak of this disease in Brazil was likely triggered by using a shared needle to vaccinate a
55 flock of 1000 sheep. Clinical signs observed before death in this outbreak included severe

depression, swelling around the vaccination site, subcutaneous edema, lameness, and crepitation (2, 9). Morris et al. reported a case of malignant edema in a 1-year-old Friesian sheep following blood sampling from the jugular vein. Clinical signs in this Friesian sheep were similar to those observed in the previously mentioned outbreak. Additionally, during necropsy, the animal was in very good physical condition. The skin in the swollen area exhibited blue discoloration, and crepitation was evident in the subcutaneous tissue. *C. septicum* and *C. sordellii* were isolated from the lesions and were further confirmed by a direct fluorescent antibody test (8).

Sheep breeding stands as the cornerstone of Iran's livestock production sector, particularly for meat consumption, reflecting its popularity among the populace (10). However, the industry faces significant threats from diseases such as malignant edema, which poses a pervasive risk owing to its ubiquitous agents. Despite its potential danger, there remains a notable dearth of comprehensive studies on this disease in sheep, particularly within Iran and for local livestock. The limited literature available underscores the pressing need for increased research and understanding of malignant edema's epidemiology and management. Since vaccination against its causative agent is typically performed, and the disease is not considered a significant threat, clinicians and researchers tend to overlook it. Consequently, despite its lethality, fewer cases of this disease are documented. Recognizing the importance of addressing this gap, the current study aims to comprehensively explore the clinical manifestations, post-mortem observations, laboratory diagnosis techniques involving both bacteriological and molecular approaches, and treatment modalities related to malignant edema across three imported sheep industrial farms.

2. Materia and Methods

2.1. Animals and Sampling

Following the observation of concerning signs of malignant edema in three separate flocks in 2021—comprising two Île-de-France flocks and one Roman flock—a research initiative was initiated to pinpoint the causative agent and establish a definitive diagnosis for the ailment. The disease impacted two ewes within one Île-de-France flock, along with two ewes and one ram in another flock of the same breed, and two rams within a Roman breed flock. It's noteworthy that these sheep breeds were imported into Iran. Seven months prior to the occurrence of the disease, these animals had been administered the Enterotoxemia Quadrivalent Bacterin Vaccine from the

٨٦ Razi Vaccine & Serum Research Institute of Iran. This vaccine includes *C. perfringens* types D,
٨٧ C, B, and *C. septicum* Bacterins.

٨٨ Regular clinical examinations were conducted whenever signs were observed. Subsequently,
٨٩ subcutaneous swellings were aseptically aspirated using sterile syringes from the affected areas.
٩٠ The samples were promptly transported to the bacteriology laboratory within two hours under cold
٩١ chain conditions (+2/+8 °C in an icebox). Antibiotic treatment (Penicillin G
٩٢ 8,000 IU/kg + Streptomycin 10 mg/kg, administered once daily intramuscularly for seven days)
٩٣ was administered to these animals following sample collection (5). Unfortunately, one of the Île-
٩٤ de-France rams succumbed to the disease, and the animal underwent necropsy within 2 hours of
٩٥ death.

٩٦ **2.2. Bacteriological procedure**

٩٧ Samples from the affected areas were processed into smears using standard conditions and
٩٨ then subjected to Gram staining. Subsequently, a bacteriological procedure was conducted to
٩٩ isolate the disease-causing agent.

١٠٠ Samples were inoculated onto 5% sheep blood agar and Cooked Meat Medium (Oxoid, UK)
١٠١ and incubated aerobically and anaerobically at 37°C for 48 hours. Anaerobic media were supplied
١٠٢ with anaerobic gas kits (Anaerocult A, Merck). The identification process involved selecting
١٠٣ suspicious *Clostridium* colonies for further analysis, relying on their distinctive features such as
١٠٤ morphology, including shape, consistency, size, and color. The selected colonies were placed on
١٠٥ fresh agar plates, including 'Stiff' blood agar (3% agar) and normal blood agar, to obtain pure
١٠٦ cultures of *Clostridium* bacteria. The isolates were identified using conventional biochemical
١٠٧ techniques (11).

١٠٨ The biochemical tests encompassed in the analysis comprised Indole Production, Motility
١٠٩ (via stab inoculation into semi-solid media), Gas production, Starch hydrolysis, Gelatin hydrolysis,
١١٠ inoculation in Egg Yolk Agar for examination Lecithinase and Lipase Production, Casein
١١١ digestion, Nitrate reduction, and sugar fermentation, which involved Glucose, Lactose, Sucrose,
١١٢ and Maltose (11, 12).

١١٣ **2.3. Molecular assay (PCR)**

114 To validate the molecular identification of *C. septicum* after bacteriological techniques, a
115 PCR assay was conducted following the protocol used by Khiav and Paradise (13). In the current
116 study, the presence of the Alpha toxin gene, a critical virulence factor involved in the pathogenesis
117 of gas gangrene caused by *C. septicum*, was investigated using PCR. Considering that *C. chauvoei*
118 shares similar biochemical characteristics and is often isolated alongside *C. septicum*, the growth
119 colonies were also screened for the presence of *C. chauvoei* using a species-specific PCR assay.
120 The primers and PCR conditions employed in the present study are detailed in **Table 1**.

121 In summary, bacterial cells were centrifuged and diluted in TE solution containing lysozyme
122 (1 mg/mL). After adding 10% SDS, the mixture was then incubated at 37°C for a minimum of 30
123 minutes. Proteinase K (50 mg/mL) was introduced and incubated at 56°C for an hour. Extraction
124 was performed using phenol and chloroform solutions in equivalent amounts. Sodium acetate (1:10
125 v/v) and isopropanol (1 v/v) were added to the mixture, which was left to incubate overnight at -
126 20 °C. The DNA was pelleted by centrifugation at 12500 rpm for 10 minutes at 4°C. The resulting
127 sediment was washed with 70% ethanol, dried, and finally dissolved in TE buffer. A NanoDrop
128 instrument (Nanodrop Technologies, USA) was used to measure DNA quality and quantity.

129 The final reaction mixture volume was 30 µL, comprising 1.8 µL of 25 mM MgCl₂, 3 µL of
130 10X PCR buffer (SinaClon, Iran), 0.6 µL of 10 mmol dNTPs, 0.5 µL of Taq DNA polymerase (5
131 units/mL) (CinnaGen, Iran), 2.5 µL of DNA template (100 ng/µL), 1 µL of each primer (10
132 pmol/mL) (**Table 1**) (CinnaGen, Iran), and 20 µL of distilled water. Distilled water and type D *C.*
133 *perfringens* served as negative controls, while the *C. septicum* vaccine strain acted as a positive
134 control. Subsequently, the PCR product was combined with 2 µL of 6x gel loading dye,
135 electrophoresed, stained with ethidium bromide (0.5 µg/mL), and visualized using a UV
136 transilluminator.

137 **Table 1.** The primers and PCR conditions utilized in this investigation

Target gene	Sequences (5'-3')	Amplicon sizes	PCR conditions	References
<i>C. septicum</i> Alpha toxin gene*	F-ATCGGAAACATGAGTGCTGC R- AGTCTTTATGCTTCCGCTAG	270 bp	94 °C 1 min 55 °C 1 min 72 °C 1 min (30 cycles)	(13)
<i>C. chauvoei</i> flagellin gene (<i>fliC</i>)	FlaF-AGAATAAACAGAAGCTGGAGATG FlachR-TACTAGCAGCATCAAATGTACC	535 bp	94 °C 1 min 55 °C 1 min 72 °C 90 s (30 cycles)	(3, 27)

*The hemolysin gene sequence is exclusive to *C. septicum* and does not exhibit similarity to other clostridial toxins (13).

139

140 3. Results

141 3.1. Microbiological findings

142 The microscopic examination of the smears, coupled with the bacteriological procedures
143 involving isolation of the causative agent, revealed the presence of *C. septicum*, a finding
144 subsequently corroborated through PCR assay.

145 Microscopic examination following Gram staining revealed the presence of large Gram-
146 positive bacilli in the Gram-stained smears obtained from muscle and subcutaneous tissues. These
147 bacilli appeared either sporulated or non-sporulated (Spores oval and subterminal) and displayed
148 pleomorphic characteristics, suggesting variability in their cellular forms. In anaerobic conditions,
149 turbidity was observed at the bottom of the Cooked Meat Medium tube, and *C. septicum* was
150 successfully isolated upon reculture on blood agar. Conversely, no growth was observed during
151 the incubation of samples under aerobic conditions. The colonial appearance of the isolates
152 exhibited traits consistent with those of *C. septicum*, characterized by swarming and spreading,
153 along with hemolytic growth on normal agar. On 'stiff' agar, the colonies appeared irregular,
154 featuring a rhizoid edge.

155 The biochemical test outcomes indicated negative Indole production, positive motility,
156 positive gas production, positive starch hydrolysis, positive Gelatinase production, negative
157 Lecithinase and Lipase activity, positive Nitrate reduction, and fermentative ability and acid
158 production with sugars Glucose, Lactose, and Maltose. Acid production from sucrose was not
159 observed. Biochemical findings have verified the existence of *C. septicum* based on referenced

160 sources (11). Overall, these findings collectively support the diagnosis of malignant edema caused
161 by *C. septicum* in the affected sheep.

162 The amplification of the hemolysin gene (Alpha-toxin gene) of *C. septicum* bacteria using
163 specific primers yielded an amplicon with a size of 270 base pairs (bp). The 270 bp amplicon aligns
164 with the expected size for the hemolysin gene, further validating the molecular diagnosis of *C.*
165 *septicum* infection in the affected sheep. No 535 bp amplicon corresponding to the *C. chauvoei*
166 flagellin gene (*fliC*) was detected, leading to the exclusion of *C. chauvoei* presence in the lesions.

167 **3.2. Clinical observations**

168 All clinical and macroscopic findings and bacteriological observations confirmed the
169 diagnosis of malignant edema, and the presence of *C. septicum* in lesions created in infected sheep
170 was confirmed. Except for the Île-de-France ram, which was dead, the rest of the animals were
171 treated, and the signs of gas gangrene disappeared within three weeks following antibiotic therapy.
172 Still, the skin of the area where the lesions were created became necrotic and did not heal (**Figure**
173 **1**).

174 The initial signs observed during the detailed clinical examination included impaired general
175 condition, high fever (41.4 – 42.2 °C), tachypnea, tachycardia, weakness, and colic. These were
176 followed by localized or regional pain, doughy swelling, edematous and painful swelling, local
177 erythema, wetness, and gelatinous secretions (**Figure 1**). As time progressed, the swelling
178 intensified, and the skin appeared dark and taut. Eventually, the taut skin cracked and assumed a
179 yellow hue, with edematous fluid seeping from the cracks, varying from a thin serum to a
180 gelatinous deposit (**Figure 2 and Figure 3**). Typically, skin gangrene accompanied by
181 subcutaneous and intermuscular connective tissue edema surrounding the infection site became
182 apparent.

183 Signs of prostration were noted in three animals during the study. Subcutaneous crepitation,
184 a crackling sensation caused by gas within tissues, was clearly detectable in four cases. However,
185 evidence of subcutaneous gas production was less pronounced than in blackleg disease cases.
186 Clinical features observed included cellulitis at the injury site, characterized by minimal gangrene
187 and gas formation (crepitation). Additionally, tissue swelling due to edema, along with a
188 discoloration of the overlying skin and a sensation of coldness, was noted. During the necropsy of

189 the deceased Île-de-France ram, the animal was found to be in very good physical condition, but
190 clear macroscopic findings of crepitation were observed.



191
192 **Figure 1.** A sheep exhibiting signs of malignant edema disease caused by *C. septicum* infection displays
193 signs of wetness, along with gelatinous secretions and edematous, doughy swelling.

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196
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Figure 2. Malignant edema in a sheep exhibits notable discoloration and gangrene of the subcutaneous tissue, alongside the presence of gelatinous secretions.

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198
199 **Figure 3.** Tensile swelling, dark and taut skin with skin cracks, and eventually edematous fluid from
200 cracks in a sheep affected by malignant edema. Tensile swelling refers to the abnormal swelling of tissues
201 under tension. In the case of malignant edema, it's likely caused by gas and fluid buildup in the affected
202 area. The tissues become stretched, leading to visible swelling.

203 204 205 **4. Discussion**

206 The diagnosis of malignant edema in sheep relies on a combination of clinical, macroscopic,
207 and microbiological findings. This study's clinical examinations revealed a range of signs
208 indicative of the disease, including impaired general condition, weakness, respiratory and cardiac
209 abnormalities, localized pain, swelling, and high fever. These clinical manifestations are consistent
210 with previous reports of malignant edema in sheep (2, 4, 5, 14). Crepitation, observed as a
211 macroscopic feature at the lesion site of malignant edema, was evident in the current study. This
finding corroborates observations documented in other studies involving similar cases. In the

212 current investigation, malignant edema was diagnosed, and treatment was administered to six
213 sheep, resulting in their recovery. One animal succumbed to the illness, underscoring the fatal
214 consequences of this disease when not promptly diagnosed and treated (1, 2, 8, 14, 15). The current
215 research stands out as the sole investigation documenting the occurrence of malignant edema
216 disease in Île-de-France and Romane sheep breeds. It's intriguing to observe that most malignant
217 edema cases in breeding farms involve imported sheep breeds in Iran, while there are no
218 occurrences of this disease in local breeds.

219 In the present study, subcutaneous edema, doughy swelling, taut skin, and crepitation were
220 evident in sheep with malignant edema. This finding is in common with other reported cases of
221 malignant edema (3, 8, 14, 16). Alpha toxin production has resulted in subcutaneous bloating,
222 darkening of edematous skin areas, and interstitial hemorrhage in muscle tissue. Indeed, studies
223 have shown that *C. septicum* needs to produce an Alpha toxin to manifest specific clinical signs
224 (3, 6, 17, 18). In a 2005 study, Kennedy et al. reported a striking difference between alpha toxin-
225 positive and negative strains in terms of virulence (19). The Alpha toxin of *C. septicum* is
226 structurally and functionally similar to the Epsilon toxin of *C. perfringens* type B and D and the
227 Aerolysin of *Aeromonas hydrophila*. On the other hand, the Alpha toxin of *C. septicum*, unlike *C.*
228 *perfringens*, causes the infiltration of immune system cells into the infection site. In infections
229 caused by *C. septicum*, hemorrhage due to Alpha toxin-induced microvascular destruction leads to
230 decreased blood flow in the infection site. This ultimately leads to ischemia that will support the
231 survival of *C. septicum* in the absence of external trauma (3, 14, 15, 17, 20).

232 In general, *C. septicum* tends to respond well to medications like Penicillin G, Ampicillin,
233 Chloramphenicol, Clindamycin, Cephaloridine, Oleandomycin, Erythromycin, Lincomycin, and
234 Tetracyclines (5, 17, 21). In the present investigation, prompt identification of clinical signs in
235 afflicted animals, indicating the onset of the disease's early stages and appropriate treatment,
236 helped prevent animal fatalities despite leaving necrotic skin lesions untreated. This outcome
237 underscores the potential benefits of antibiotic therapy, particularly in mitigating fatalities
238 attributed to *C. septicum* infection when the disease is diagnosed early. Early intervention with
239 antibiotics not only aids in halting the progression of the infection but also contributes to
240 minimizing mortality associated with *C. septicum*. The effectiveness of antibiotic treatment in the
241 early stages of the disease highlights the importance of timely diagnosis and intervention. Early

242 detection allows for the implementation of appropriate therapeutic measures, including antibiotic
243 administration, which can arrest the spread of the infection and prevent the onset of severe
244 complications (1, 14).

245 Comparisons of malignant edema in sheep cases caused by *C. septicum* are challenging due
246 to the limited number of studies addressing this specific condition. The scarcity of comprehensive
247 research on malignant edema in livestock, particularly those focusing on the involvement of *C.*
248 *septicum*, hinders the ability to draw meaningful comparisons across different investigations.
249 Gazioglu and colleagues conducted a study in 2018, identifying nine goats with malignant edema,
250 showcasing clinical signs and necropsy findings akin to the present research—moreover, the
251 method employed for detecting and isolating *C. septicum* aligned with the present study. Among
252 the sufferers, four goats were treated following the technique outlined in the current research, and
253 they exhibited successful responses to treatment. To validate the isolation of the disease agent, they
254 conducted a PCR assay similar to the method described in the present study. Additionally, they
255 observed an amplicon 270 bp, indicative of the alpha-toxin gene of *C. septicum*. (3). Lewis (2007)
256 reported successful outcomes with early invasive antibiotic treatment in sheep afflicted with
257 malignant edema (22).

258 In a documented incident in Brazil in 2006, a flock comprising 1200 sheep and goats
259 experienced a significant loss of livestock, with 40 sheep and 20 goats succumbing to malignant
260 edema within 24 hours to 5 days following non-standard vaccination procedures. These animals
261 also had a vaccination history with *C. septicum* bacterin. The clinical manifestations observed in
262 this outbreak closely resembled those documented in the present study. Analysis of samples
263 collected from the lesion sites revealed the presence of *C. novi* and *C. septicum*, as confirmed by
264 direct FAT testing (6).

265 In a case report by Cihan et al. (2010) in Turkey, 20 sheep were diagnosed with malignant
266 edema, and the clinical signs closely resembled those observed in the present study. These animals
267 also received *C. septicum* bacterin. (23). This finding underscores the consistency of clinical
268 manifestations associated with malignant edema across different geographical regions and
269 livestock populations. A case of malignant edema linked to umbilical infection was documented
270 in a deceased Merino lamb within a flock comprising 50 ewes and 35 newborn lambs in Argentina.
271 The sampling and diagnostic approach mirrored that of the present study, utilizing Gram staining

272 and direct FAT, yielding comparable results. During the post-mortem examination, despite the
273 lamb being in good physical condition, macroscopic lesions consistent with those observed in the
274 present study were identified (24). In the abovementioned study, the affected animals had a
275 vaccination history with *C. septicum* antigen. However, akin to the present study, this disease still
276 resulted in problems and damages. The presence of this microorganism in the infection may be
277 attributed to a suboptimal vaccination method, a lack of individual immune response, or an unusual
278 challenge dose (8).

279 Microbiological analysis played a crucial role in confirming the diagnosis. Gram staining of
280 smears from affected tissues revealed the presence of large Gram-positive bacilli, characteristic of
281 *Clostridium* spp. Further confirmation was achieved through the FAT, which specifically identified
282 *C. septicum* in all cases. Culture and purification techniques supported these findings by isolating
283 *C. septicum* from the affected tissues, particularly in anaerobic conditions. *C. septicum*, commonly
284 found in soil, has also been detected in the feces of both humans and healthy animals. This
285 pathogen, acting as a post-mortem invader, can swiftly disseminate throughout the body from the
286 intestines of deceased or distressed animals, particularly ruminants. Its rapid spread raises the
287 possibility of isolating *C. septicum*, potentially leading to misdiagnosis, even if a necropsy is
288 performed immediately after the animals' demise (1, 13, 17). However, in this study, *C. septicum*
289 was isolated not only from samples taken during necropsy but also from the skin lesions of live
290 but diseased animals.

291 Microbiological and biochemical identification can be a time-consuming process,
292 compounded by the challenge of distinguishing between Clostridia, notably *C. chauvoei* and *C.*
293 *septicum*, due to their similarities (13). The current study tried to introduce a standardized protocol
294 for isolating *C. septicum*, which could serve as a valuable resource for future research endeavors
295 seeking to isolate and identify the causative agent of malignant edema. Molecular findings
296 validated the efficacy of the bacteriological method employed. In the current study, all *C. septicum*
297 strains that tested positive in biochemical and microbiological assays exhibited the expected PCR
298 product size (270 bp), confirming the presence of the bacterium in the samples. Differentiating
299 between *C. septicum* and *C. chauvoei* alpha-toxin is only possible through PCR analysis, not
300 immunological methods. The sequence of the hemolysin gene is unique to *C. septicum* and does
301 not show homology with other clostridial toxins.

302 The current study noted that even though the signs of malignant edema diminished in the
303 treated animals after antibiotic therapy, the skin in the region where the lesions had developed
304 underwent necrosis and failed to heal. This implies that although the antibiotic treatment
305 successfully managed the infection and alleviated systemic signs, it couldn't halt tissue damage
306 and necrosis at the site of the initial lesions. As time elapsed, the damage inflicted by the disease
307 remained unrepaired, indicating the severity and persistence of its effects. Additional interventions
308 or supportive care might be needed to manage the necrotic tissue and facilitate healing in these
309 affected areas (2, 14).

310 Preventing malignant edema in sheep is paramount to avoid its detrimental effects on
311 livestock health and economic losses. Several measures can be implemented to mitigate the risk
312 of disease occurrence. First and foremost, maintaining proper hygiene and sanitation in sheep
313 housing and handling areas can help minimize exposure to *Clostridium* spores in the environment.
314 Regular cleaning and disinfection of equipment used for routine management practices, such as
315 vaccinations and shearing, can also reduce the risk of wound contamination. Furthermore,
316 vaccination against *Clostridium* species, including *C. septicum*, should be considered a preventive
317 measure (14, 15, 25). The industrial flocks investigated in the present study were vaccinated
318 against *C. septicum*. Cases of malignant edema have sporadically occurred within these flocks.
319 This underscores the significance of vaccination as a preventive measure against the disease.

320 Sporadic cases of malignant edema within the vaccinated flocks highlight an important
321 aspect of disease prevention. While vaccination is an effective strategy for reducing the overall
322 incidence of the disease, its effectiveness may not be absolute in preventing every single case.
323 Factors such as variations in vaccine efficacy, environmental conditions, and individual animal
324 susceptibility can contribute to occasional breakthrough infections despite vaccination efforts.
325 Therefore, even in vaccinated populations, the "possibility" of disease occurrence remains high.
326 However, the fact that these cases are sporadic suggests that vaccination still plays a significant
327 role in disease prevention by reducing outbreaks' overall frequency and severity. It emphasizes the
328 importance of increased prognosis and continued vaccination programs to maintain herd immunity
329 and minimize the impact of the disease on livestock populations (6, 15).

330 Additionally, proper wound management practices are crucial for preventing the entry of
331 *Clostridium* spores into tissues. Prompt and thorough cleaning and disinfection of wounds, along

۳۳۲ with timely veterinary intervention, can help minimize the risk of infection and subsequent
۳۳۳ development of malignant edema. Educating sheep farmers and livestock handlers about the signs,
۳۳۴ risk factors, and preventive measures for malignant edema is essential for disease control and
۳۳۵ management. Awareness campaigns and training programs can empower farmers to implement
۳۳۶ appropriate biosecurity measures and vaccination protocols to safeguard their flocks against this
۳۳۷ potentially devastating disease (1, 13, 26).

۳۳۸ In conclusion, diagnosing malignant edema in sheep necessitates a thorough approach
۳۳۹ encompassing clinical assessment, microbiological scrutiny, and histopathological examination.
۳۴۰ Prevention measures, such as maintaining proper hygiene, administering vaccinations, and
۳۴۱ effectively managing wounds, are imperative to mitigate disease risk and safeguard the well-being
۳۴۲ and productivity of sheep herds. The escalating instances of malignant edema in small ruminants
۳۴۳ across Iran underscore the urgency of giving this disease greater attention. This study furnishes
۳۴۴ valuable insights into the prognosis of malignant edema. Considering the absence of recent
۳۴۵ research on malignant edema in Iran, heightened investigation coupled with enhanced
۳۴۶ management practices and training holds promise for reducing disease incidence and averting
۳۴۷ economic losses.

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۳۵۲ **Authors' contribution**

۳۵۳ Study concept and design: H. E.
۳۵۴ Acquisition of data: H. E. and S. M. J.
۳۵۵ Analysis and interpretation of data: H. E. and S. M. J.
۳۵۶ Drafting of the manuscript: S. M. J.
۳۵۷ Administrative, technical, and material support: H. E.
۳۵۸ Study supervision: H. E.

۳۵۹ **Ethics**

۳۶۰ The authors of this study affirm that all ethical standards were upheld in the preparation of
۳۶۱ the submitted article.

۳۶۲ **Conflict of interest**

۳۶۳ The authors declare that they have no conflict of interest.

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۳۶۶ **Data availability**

۳۶۷ The data produced and/or analyzed during the present study can be obtained from the
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