

Evaluating the Diversity of *Chit18-5* Gene Region across *Trichoderma* Species for Effective Biocontrol Strategies

A. Hassanzadeh

M.A. Tajick Ghanbari⊠

Department of Plant Protection, Faculty of Crop Science, Sari Agricultural Sciences and Natural Resources University, Sari, Iran.

Abstract: Trichoderma species are widely used as biological agents to control plant diseases. Chitinases are crucial in mycoparasitism and defense against other fungi or arthropods. In this study, we evaluated 41 amino acid sequences related to the Chit18-5 gene from four sections and 15 Trichoderma species. The conserved domains, motifs, and phylogenetic tree were analyzed using the InterProScan database, COBALT tool, MEME V5.5.1 software, ClustalW algorithm, and MEGA11 software. The results showed that the gene region under investigation can effectively distinguish different Trichoderma species and is an effective tool for optimizing biocontrol strategies. This study highlights the potential of exploring genetic diversity as a means of identifying new solutions for managing pests and diseases in agriculture. The putative motifs of chitinase proteins identified in this study may participate in Trichoderma antagonistic activities.

Keywords: *Trichoderma*, Chit18-5 gene region, biocontrol, phylogenetic tree, genetic diversity.

INTRODUCTION

Biological agents such as *Trichoderma* species are frequently employed to manage plant diseases (Mukherjee et al. 2022). By combining mycoparasitism, antibiosis, induced defense response (IDR), and competition, *Trichoderma* spp. aid in the disease's suppression (Sharma et al. 2017). The capacity of some species to parasitize other plantpathogenic fungi sparked the initial investigation into the potential use of *Trichoderma* in biological control (Weindling 1932). Glucanases, chitinases, and proteases are recognized as important enzymes in

Corresponding Author: E-mail: m.tajick@gmail.com

© 2023, Published by the Iranian Mycological Society http://mij.areeo.ac.ir

mycoparasitism (Cortes et al. 1998, Vazquez-Garciduenas et al. 1998, Carsolio et al. 1999), because the cell wall of true fungi is made up of glucans, chitin, and proteins (Gow et al. 2017, Ruiz-Herrera & Ortiz-Castellanos 2019, Garcia-Rubio et al. 2020). Furthermore, β-1,4-linked N-acetyl-Dglucosamine (GlcNAc) monomers form the renewable polymer known as chitin, which is the second most prevalent polysaccharide in nature. It is an important part of the exoskeleton of arthropods and the cell walls of fungi. Enzymes called chitinases (EC 3.2.1.14) hydrolyze the bonds that hold GlcNAc residues together. According to Karlsson & Stenlid (2008), they are essential for a variety of biological processes, such as autolysis, hyphal development, branching, and cell wall remodeling during spore germination and constriction. In order to supply nutrients and engage in an aggressive pattern of competition and defense against other fungi or arthropods, they also break down exogenous chitin found in the hyphal cell wall or the exoskeleton of arthropods. They accomplish this by killing their fungal prey and then feeding on the contents of the dead cells (Karlsson & Stenlid, 2008, Seidl-Seiboth et al. 2014, Seidl 2008). Because chitinases are involved in defense responses against infections, previous research on them has mostly concentrated on gene cloning and transformation, particularly with regard to the ech-42 gene (Bolar et al. 2000, De La Cruz et al. 1992, Emani et al. 2003, Garcia et al. 1994, Gentile et al. 2007). However, recent advances in genomic analyses have expanded our understanding of chitinase genes and their classification models for filamentous fungi. For instance, Seidl et al. (2005) used the T. reesei QM6a genome as a reference to describe the first genome-wide investigation of fungal chitinase genes. They identified eighteen chitinases that were phylogenetically categorized into A, B, and C, each of which was further subdivided into many subgroups. Among fungal taxa, the size of the chitinase gene family varies significantly. For instance, while *Schizosaccharomyces pombe* has only one chitinase gene, Trichoderma virens has as many

Submitted 28 Sept 2023, accepted for publication 12 Feb 2024

as 36 (Karlsson & Stenlid 2008, Kubicek et al. 2011). It is noteworthy that every fungal chitinase that has been identified solely falls under the glycoside hydrolase (GH) family 18 (Seidl et al. 2005). Each year, insect pests and plant-pathogenic fungi seriously harm crops. Fortunately, genera in the order Hypocreales, including Beauveria, Metarhizium, and Trichoderma, are effective mycoparasites that inhibit these pests (Shah & Pell 2003, Woo et al. 2014). Alternatives to fungicides that are safe, long-lasting, effective, and environmentally friendly are becoming more and more necessary in modern agriculture. In order to fulfill this requirement, these biological resources have been used as biocontrol agents in agriculture (Wang & Zhuang 2019, Wang & Zhuang 2020). Chitinases play a crucial role in the initial stage of mycoparasitic infections, which involves the lysis of the host cell wall (Herrera-Estrella & Chet 2003, Howell 2003, Druzhinina et al. 2011). Interesting concerns about the evolution of this significant gene family are raised by the expansion and contraction of the chitinase gene family as well as the molecular mechanisms underpinning these changes. Recent research has shown that mycoparasitic fungal species have higher levels of group B and group C chitinases, although their amino acid sequences have a low degree of conservation. According to these results, evolutionary paths should become more diverse (Seidl-Seiboth et al. 2014, Ihrmark et al. 2010). Through a bioinformatics analysis of the amino acid sequences of the Chit18-5 protein, this study intends to use genomic information to analyze the relationship between chitinases and the mycoparasitic lifestyle of Trichoderma species.

MATERIALS AND METHODS

In this study, were evaluated 41 amino acid sequences related to the Chit18-5 gene from four sections and 15 Trichoderma species (Table 1S). Sequences of 100-400 amino acids were received in FASTA file format from the UniProtKB protein bank (Coudert et al. 2023). The conserved domains in this gene region were checked by using the InterProScan database and COBALT tool in the NCBI database (Papadopoulos & Agarwala 2007, Jones et al. 2014). Identifying protected motifs and determining the position of these motifs in domains, using MEME V5.5.1 software with default parameters, including choosing motifs regardless of the frequency of repetition, identifying a maximum of 10 motifs and the lowest and the analysis focused on the maximum length of motifs, which were between 6 and 50 amino acids long (Bailey et al. 2015). The amino acid sequences were aligned using the ClustalW algorithm and the Neighbor-Joining tree was drawn with MEGA11 software (Tamura et al. 2021). The strength of the internal branches of the resulting trees was tested with bootstrap (BS) analysis using 1000 replications (Felsenstein 1985). In addition, the amino acid sequences were translated into nucleotide sequences using the tblastn tool in the NCBI gene bank, and the

nucleotide sequences with the highest percentage of similarity were received in FASTA file format. The nucleotide sequences were aligned using the ClustalW algorithm and the phylogeny tree was drawn as above mentioned. To root the phylogenetic tree, the amino acid and nucleotide sequence of *Trichoderma atroviride* related to the *Chit18-3* gene was used as the outgroup taxa.

RESULTS AND DISCUSSION

Assessing the gene region related to Chit18-5 by using the InterProScan database showed that there are three to five conserved domains in the amino acid sequences associated with this gene region (Table 1), and these domains were confirmed in the sequences that were evaluated (Fig. 1). Furthermore, the presence of domains was confirmed using the COBALT tool in the NCBI database (Fig. 2). The results of the assessment of sequences with MEME V5.5.1 software revealed two to nine different motifs in all the amino acid sequences that were evaluated. The differences in the positions of these motifs led to the separation of the sequences. The identified motifs and their distribution in the amino acid sequences investigated in this research are shown in Fig.s 3 and 4. The abundance of each amino acid indicates its prevalence in the related motif. Our results reveal the presence of diverse domains and motifs within the gene region associated with chitinase enzymes across Trichoderma species, both intra- and inter-species.

Two phylogenetic trees were constructed using amino acid (Fig. 5) and nucleotide (Fig. 6) sequences. Examining of both trees demonstrated a clear separation of species, with those sharing the same name clustering (Fig. 5). Furthermore, the evaluated species in this study were classified into four sections of the *Trichoderma* genus, namely *Harzianum* and *Virens, Longibrachiatum, Pachybasium,* and *Trichoderma*, based on their placement in the respective sections. The findings suggest that the gene region under investigation can effectively distinguish these sections.

Trichoderma atroviride, related to the *Chit18-3* gene, was assigned as the outgroup taxa. The results showed that the separation was done correctly due to the difference between these two gene regions, and the tree was rooted accordingly. The alignment of chitinase proteins indicated that this gene region is highly conserved, particularly in its domains, and has a relatively similar structure across different *Trichoderma* species. Upon examining both phylogenetic trees, it was observed that, in most cases, the order of species placement was consistent between the two trees.

Trichoderma species are notoriously difficult to identify morphologically due to their high diversity. As a result, molecular methods have been employed in various studies for species identification

In this research, two phylogenetic trees were compared, revealing that chitinase proteins-specifically the *Chit18-5* gene region - exhibit greater inter-species diversity than intra-species diversity. This can be attributed to the presence of conserved domains within the protein. The results suggest that the gene region studied is an effective tool for distinguishing between different *Trichoderma* species.

The results of this study highlighting the diversity in amino acid sequences of the *Chit18-5* gene region among many *Trichoderma* species may have significant practical implications for the agricultural sector. One of the most significant biopolymers found in nature is chitin, which is mostly produced by nematodes, arthropods, and fungi. It serves as a scaffold in insects, supporting the peritrophic matrices that line the intestinal epithelium as well as the cuticles of the trachea and epidermis. Growth and morphogenesis in insects are exclusively dependent on their ability to reorganize structures that include chitin. In several tissues, insects continuously manufacture chitin synthases and chitinolytic enzymes for this reason. Strict control of the involved enzymes is necessary for the coordination of chitin synthesis and degradation during development (Merzendorfer & Zimoch 2003). In true fungi such as Ascomycota, Basidiomycota, and Chytridiomycota cell walls are usually based on glucans and chitin, and also the fibers are chitin microfibrils, i.e. bundles of linear b-(1,4)-linked N-acetylglucosamine chains (Webster & Weber 2007). There is strong evidence that wall-lytic enzymes like chitinases and glucanases have an impact on the softness of the apical cell wall (Fontaine et al. 1997, Horsch et al. 1997). It has been demonstrated that the wall of resting spores contains chitin and is notably thick (Moxham & Buczacki 1983). The enzyme EC 3.2.1.14 - chitinase binds to chitin. It randomly cleaves glycosidic linkages in chitin and chitodextrins in a non-processive mode, generating chitooligosaccharides and free ends on which exo-chitinases and exo-chitodextrinases can act (Rottloff et al. 2011). Not all Trichoderma strains can exhibit strong antagonistic effects against pathogens (Liu et al. 2022).

 Table 1. The list of identified domains by using InterProScan

Domains	Accession number
Glycoside hydrolase family 18, catalytic domain	All sequences
Glycosyl hydrolases family 18 (GH18) domain profile	All sequences
Glycosyl hydrolases family 18	All sequences
Chitinase II	All sequences except E5F5N0
2g34	All sequences except E5F5N0



Fig. 1. The structure of identified domains by using InterProScan

Sequence ID		Start	Alignment																			End	Organism
			1 20	40	60	80	100	120	140	160	180	200	220	240	260	280	300	320	340	360	380 31	6	
			h.														-+					-	
Query_10001	(+)	1												-				-		3		269	
Query_10002	(+)	1																			9	266	
Query_10003	(+)	1							_													268	
Query_10004	(+)	1														_						237	
Query_10005	(+)	1						1						-		_				3		254	
Query_10006	(+)	1												-		_				_		260	
Query_10007	(+)	1					100			_				_				-				257	
Query_10008	(+)	1												_		_						253	
Query_10009	(+)	1												-		_						282	
Query_10010	(+)	1														_		-		19	1	257	
Query_10011	(+)	1																-		19	8	267	
Query_10012	(+)	1																		3		271	
Query_10013	(+)	1														_		-		4		268	
Query_10014	(+)	1														_		-		1		267	
Query_10015	(+)	1														-		1		9		269	
Query_10016	(+)	1														_	-	-				268	
Query_10017	(+)	1														-		-			8	277	
Query_10018	(+)	1																				124	
Query_10019	(+)	1														_		-				228	
Query_10020	(+)	1														_		-		8		255	
Query_10021	(+)	1												1.0		-		-		3		268	
Query_10022	(+)	1														-		-		9		269	
Query_10023	(+)	1														_	-	-		2		268	
Query 10024	(+)	1														_		-		1		270	
Query_10025	(+)	1														-		-				275	
Query 10026	(+)	1						2								-		-		3		265	
Query_10027	(+)	1														-		-		1		288	
Query 10028	(+)	1														_		-				253	
Query 10029	(+)	1														_		-		1	6	273	
Query 10030	(+)	1														_	-	-				255	
Query 10031	(+)	1														_		-		3		271	
Query 10032	(+)	1														-		-		1		271	
Query_10033	(+)	1														-		-		1		273	
Query 10034	(+)	1														_		-				228	
Query 10035	(+)	1														_		-		3	4	277	
Query 10036	(+)	1														_		-			C.	277	
Query 10037	(+)	1														_		-				255	
Query 10038	(+)	1														-		-			().	265	
Query 10039	(+)	1														-		-		1		270	
Query 10040	(+)	1						1								-		-			2	205	
Query 10041	(+)	1														_		_	-			234	
Query 10042	(+)	1	-													_		0_3				358	
Guery_10042	(+)	1							_		100											336	

NCBI Multiple Sequence Alignment Viewer, Version 1.23.1

Fig. 2. The structure of identified domains by using the COBALT tool

	Logo	E-value	Sites	Width
1.	<pre>#ENKDWGFDGIDJDWEYPADsTQAsNMJLLLLKEYRSQLDAYAAQYAPGYHF</pre>	2.1e-1771	38	50
2.	[®] ISCHDANL¥ANPSNPNATPENTDRAYKAYIKGGVPANKIVLGNPIYGRSFE	4.0e-1796	40	50
3.	·WENGYWDYKALPKAGATYQYDDYAKASYYSYDDSTKELISEDTPDWINTKY	5.7e-1661	39	50
4.	ILLSIAAPAGENNYSYLRLADLGQVLDYYNLMAYDYAGSWSD	1.9e-1384	41	41
5.	¥ lkslglggsnfweasadkkgadsligts	4.1e-660	30	29
6.	ĨŢ <u>\$Ġ</u> ļ <mark>ĠQ</mark> ₽¥ <mark>8ĠļĠ\$Ġ\$</mark>	2.9e-377	40	15
7.	ISTDANRKRFARTALT	1.7e-317	29	15
8.	······································	2.0e-101	23	15
9.	Ĩ₩Ţ <u>ġ</u> ŖŢŊ <mark>Ę</mark> ĄŞĄĄ	4.0e-055	17	11
10.	HKALGALDSTQ	4.4e-029	8	11

Fig. 3. Motif logo in amino acid sequences by MEME software; the size of each amino acid in a motif indicates its relative abundance.

Name	p-value	Motif Locations
A0A024HW81	4.73e-257	
A0A024HWC6	3.56e-257	
A0A024HWC2	2.73e-259	
A0A024HWB8	1.33e-276	
A0A024HVJ3	2.56e-216	
A0A024HWB6	3.91e-202	
A0A024HW54	5.01e-287	
A0A024HVT7	2.74e-301	
A0A024HVI2	5.44e-276	
A0A024HW52	6.76e-249	
E5F5K9	5.31e-289	
E5F5L3	2.97e-292	
E5F5L4	8.38e-287	
G9FLF9	6.82e-228	
G9FLG2	2.53e-293	
G9FLG6	1.59e-294	
G9FLG8	2.65e-254	
G9FLC5	9.71e-286	
E5F5N0	5.51e-39	
G9FLD8	1.77e-227	
G9FLE0	1.35e-264	
A0A024HVI4	1.03e-272	
A0A024HVI7	2.72e-278	
A0A024HVI9	7.94e-168	
A0A024HVU1	1.19e-220	
B8X9T5	1.54e-274	
GOZPCO	1.88e-274	
AUAUB5ABQU	2.536-277	
AUAUB5AFG4	1.388-259	
K4GDRZ	2.416-279	
K4GJCU	3.620-204	
K4GE78	1.228-270	
	1.040-277	
	1 130-277	
D2DPP3	1.04e-277	
A0A0B5AEM8	9 63e-264	
A0A0B5AGS0	1.13e-263	
A0A024HVS7	5.50e-262	
A0A024HVH8	1.21e-236	
A0A024HVT2	1.77e-262	
Q2Y0V8	5.23e-23	
Motif Symbol Mc 1. FM SG 3. WE FM 4. FM LL 5. YL TS 7. ST TS 7. ST HK 9. WT HK 10. HK HK	tif Consensus KDWGFDGIDID HDANLYANPQNI NGVWDYKALPKI SIAAPAGEVNYI KSIGIGGSMFWI CICOPYSGIGS DANRKNFAKTA ALGHLDSTQNIJ YSTNFASAA ALGALDSTQ	VEYPADSTQASNMILLLKEVRSQLDAYAAQYAPGYHF PNATPFNTDDAVKAYIKGGVPANKIVLGMPIYQGRSFE NGATVQYDDYXKASYSYPPSTKELISFDDINTKV SVLRLADLGQVLDYVNLMAYDYAGSWSN IASADKKGADSLIGTS IS IT LS

Fig. 4. Analysis of motifs in amino acid sequences and distribution of identified motifs by MEME software



Fig. 5. Phylogenetic tree of 41 amino acid sequences in different *Trichoderma* species by using MEGA11 software, Neighbor-Joining method, and Bootstrap analysis with 1000 replications



Fig. 6. Phylogenetic tree of 41 nucleotide sequences resulting from tblastn chitinase protein sequences in different *Trichoderma* spp. by using MEGA11 software, Neighbor-Joining method, and Bootstrap analysis with 1000 replications

The ability to distinguish between other Trichoderma species using this gene region could be a valuable tool for the identification of strong species optimizing biocontrol strategies and selecting the most effective Trichoderma strains for specific crop pests and diseases. Furthermore, these findings could also lead to further research exploring the evolutionary mechanisms behind the diversification of the Chit18-5 gene family and their relationship with mycoparasitism and other biological processes. The putative motifs of chitinase proteins identified in this study may participate in Trichoderma antagonistic activities. It has been shown that some proteins participate in plant defense pathways related to Trichoderma effector functions. In maize and cotton, for example, reactive oxygen species buildup and pathogen-related gene expression responses are induced by cerato-platanin proteins Sm1 from T. virens and its homolog Epl1 from T. atroviride (Seidl et al. 2006). This could shed light on new approaches for developing more effective biocontrol agents against plant pathogens and insect pests. Overall, this study highlights the potential of exploring genetic diversity as a means of identifying new solutions for managing diseases pests and in agriculture. By understanding the genetic mechanisms behind biocontrol agents like Trichoderma, we can develop targeted and sustainable strategies for enhancing crop yields and protecting the environment by reducing the usage of chemical fungicides and pesticides. As stated by Bononi et al. (2020), the biological control strategy primarily consists of the synthesis of particular and uptake chemicals the of specific micronutrients to plants in order to enhance defense and improve plant growth.

ACKNOWLEDGMENTS

The authors wish to acknowledge the support provided by Sari Agricultural Sciences and Natural Resources University in funding this study. Without their generous assistance, our research would not have been possible. We extend our deepest gratitude to the university for their commitment to advancing scientific knowledge in this field and their ongoing efforts to promote academic excellence. This support has enabled us to conduct rigorous research and gather the necessary data to contribute to the body of knowledge in this area.

- Bailey, T.L., Johnson, J., Grant, C.E. and Noble, W.S. 2015. The MEME suite. Nucleic Acids Research 43: 39-49.
- Bolar, J.P., Norelli, J.L., Wong, K.W., Hayes, C.K., Harman, G.E. and Aldwinckle, H.S. 2000. Expression of endochitinase from *Trichoderma harzianum* in transgenic apples increases resistance to apple scab and reduces vigor. Phytopathology 90: 72-77.
- Bononi, L., Chiaramonte, J.B., Pansa, C.C., Moitinho, M.A. and Melo, I.S. 2020. Phosphorussolubilizing *Trichoderma* spp. from Amazon soils improves soybean plant growth. Scientific Reports 10: 28-58.
- Carsolio, C., Benhamou, N., Haran, S., Cortes, C., Gutierrez, A., Chet, I. and Herrera-Estrella, A. 1999. Role of the *Trichoderma harzianum* endochitinase gene, ech42, in mycoparasitism. Applied and Environmental Microbiology 65: 929-935.
- Cortes, C., Gutierrez, A., Olmedo, V., Inbar, J., Chet, I. and Herrera-Estrella, A. 1998. The expression of genes involved in parasitism by *Trichoderma harzianum* is triggered by a diffusible factor. Molecular and General Genetics MGG 260: 218-225.
- Coudert, E., Gehant, S., de Castro, E., Pozzato, M., Baratin, D., Neto, T., Sigrist, C.J., Redaschi, N. and Bridge, A. 2023. Annotation of biologically relevant ligands in UniProtKB using ChEBI. Bioinformatics, 39: 1-5.
- De La Cruz, J., Hidalgo-Gallego, A., Lora, J.M., Benitez, T., Pintor-Toro, J.A. and Llobell, A. 1992. Isolation and characterization of three chitinases from *Trichoderma harzianum*. European Journal of Biochemistry 206: 859-867.
- Druzhinina, I.S., Seidl-Seiboth, V., Herrera-Estrella, A., Horwitz, B.A., Kenerley, C.M., Monte, E., Mukherjee, P.K., Zeilinger, S., Grigoriev, I.V. and Kubicek, C.P. 2011. *Trichoderma*: the genomics of opportunistic success. Nature Reviews Microbiology 9: 749-759.
- Emani, C., Garcia, J.M., Lopata-Finch, E., Pozo, M.J., Uribe, P., Kim, D.J., Sunilkumar, G., Cook, D.R., Kenerley, C.M. and Rathore, K.S. 2003.
 Enhanced fungal resistance in transgenic cotton expressing an endochitinase gene from *Trichoderma virens*. Plant Biotechnology Journal, 1: 321-336.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap evolution, 39: 783-791.
- Fontaine, T., Hartland, R.P., Beauvais, A., Diaquin, M. and Latge, J.P. 1997. Purification and Characterization of an Endo-1, 3-β-Glucanase from *Aspergillus fumigatus*. European Journal of Biochemistry 243: 315-321.

- Garcia, I., Lora, J.M., de la Cruz, J., Benitez, T., Llobell, A. and Pintor-Toro, J.A., 1994. Cloning and characterization of a chitinase (CHIT42) cDNA from the mycoparasitic fungus *Trichoderma harzianum*. Current genetics 27: 83-89.
- Garcia-Rubio, R., de Oliveira, H.C., Rivera, J. and Trevijano-Contador, N., 2020. The fungal cell wall: *Candida*, *Cryptococcus*, and *Aspergillus* species. Frontiers in microbiology 10: 2993.
- Gentile, A., Deng, Z., La Malfa, S., Distefano, G., Domina, F., Vitale, A., Polizzi, G., Lorito, M. and Tribulato, E. 2007. Enhanced resistance to *Phoma* tracheiphila and Botrytis cinerea in transgenic lemon plants expressing a Trichoderma harzianum chitinase gene. Plant Breeding 126: 146-151.
- Gow, N.A., Latge, J.P. and Munro, C.A. 2017. The fungal cell wall: structure, biosynthesis, and function. Microbiology Spectrum, 5: 1-25.
- Herrera-Estrella, A. and Chet, I. 2003. The biological control agent *Trichoderma*: from fundamentals to applications. Handbook of Fungal Biotechnology 2: 147-156.
- Horsch, M., Mayer, C., Sennhauser, U. and Rast, D.M. 1997. β-N-acetylhexosaminidase: a target for the design of antifungal agents. Pharmacology & therapeutics 76: 187-218.
- Howell, C.R. 2003. Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: the history and evolution of current concepts. Plant Disease 87: 4-10.
- Ihrmark, K., Asmail, N., Ubhayasekera, W., Melin, P., Stenlid, J. and Karlsson, M. 2010. Comparative molecular evolution of *Trichoderma* chitinases in response to mycoparasitic interactions. Evolutionary Bioinformatics, 6: 1-26.
- Jones, P., Binns, D., Chang, H.Y., Fraser, M., Li, W., McAnulla, C., McWilliam, H., Maslen, J., Mitchell, A., Nuka, G. and Pesseat, S. 2014. InterProScan 5: genome-scale protein function classification. Bioinformatics, 30: 1236-1240.
- Karlsson, M. and Stenlid, J. 2008. Comparative evolutionary histories of the fungal chitinase gene family reveal non-random size expansions and contractions due to adaptive natural selection. Evolutionary Bioinformatics 4: 47-60.
- Kubicek, C.P., Herrera-Estrella, A., Seidl-Seiboth, V., Martinez, D.A., Druzhinina, I.S., Thon, M., Zeilinger, S., Casas-Flores, S., Horwitz, B.A., Mukherjee, P.K. and Mukherjee, M. 2011. Comparative genome sequence analysis underscores mycoparasitism as the ancestral lifestyle of *Trichoderma*. Genome Biology 12: 1-15.
- Liu, Y., He, P., He, P., Munir, S., Ahmed, A., Wu, Y., Yang, Y., Lu, J., Wang, J., Yang, J. and Pan, X. 2022. Potential biocontrol efficiency of *Trichoderma* species against oomycete pathogens. Frontiers in Microbiology 13: 1-11.

- Merzendorfer, H. and Zimoch, L. 2003. Chitin metabolism in insects: structure, function, and regulation of chitin synthases and chitinases. Journal of Experimental Biology 206: 4393-4412.
- Moxham, S.E. and Buczacki, S.T. 1983. Chemical composition of the resting spore wall of *Plasmodiophora brassicae*. Transactions of the British Mycological Society 80: 297-304.
- Mukherjee, P.K., Mendoza-Mendoza, A., Zeilinger, S. and Horwitz, B.A. 2022. Mycoparasitism as a mechanism of *Trichoderma*-mediated suppression of plant diseases. Fungal Biology Reviews 39: 15-33.
- Papadopoulos, J.S. and Agarwala, R. 2007. COBALT: a constraint-based alignment tool for multiple protein sequences. Bioinformatics 23: 1073-1079.
- Rottloff, S., Stieber, R., Maischak, H., Turini, F.G., Heubl, G. and Mithöfer, A. 2011. Functional characterization of a class III acid endochitinase from the traps of the carnivorous pitcher plant genus, Nepenthes. Journal of Experimental Botany 62: 4639-4647.
- Ruiz-Herrera, J. and Ortiz-Castellanos, L. 2019. Cell wall glucans of fungi. A review. The Cell Surface, 5: 1-14.
- Seidl, V., Huemer, B., Seiboth, B. and Kubicek, C.P. 2005. A complete survey of *Trichoderma* chitinases reveals three distinct subgroups of family 18 chitinases. The FEBS journal 272: 5923-5939.
- Seidl, V., Marchetti, M., Schandl, R., Allmaier, G. and Kubicek, C.P. 2006. Epl1, the major secreted protein of *Hypocrea atroviridis* on glucose, is a member of a strongly conserved protein family comprising plant defense response elicitors. The FEBS journal 273: 4346-4359.
- Seidl, V. 2008. Chitinases of filamentous fungi: a large group of diverse proteins with multiple physiological functions. Fungal Biology Reviews 22: 36-42.
- Seidl-Seiboth, V., Ihrmark, K., Druzhinina, I. and Karlsson, M. 2014. Molecular evolution of *Trichoderma* chitinases. In Biotechnology and biology of *Trichoderma* 67-78.
- Shah, P.A. and Pell, J.K. 2003. Entomopathogenic fungi as biological control agents. Applied microbiology and biotechnology 61: 413-423.
- Sharma, V., Salwan, R. and Sharma, P.N. 2017. The comparative mechanistic aspects of *Trichoderma* and probiotics: scope for future research. Physiological and Molecular Plant Pathology 100: 84-96.
- Tamura, K., Stecher, G. and Kumar, S. 2021. MEGA11: molecular evolutionary genetics analysis version 11. Molecular Biology and Evolution 38: 3022-3027.
- Vazquez-Garciduenas, S., Leal-Morales, C.A. and Herrera-Estrella, A. 1998. Analysis of the β -1, 3glucanolytic system of the biocontrol agent

Trichoderma harzianum. Applied and environmental microbiology 64: 1442-1446.

- Wang, C. and Zhuang, W.Y. 2019. Evaluating effective *Trichoderma* isolates for biocontrol of *Rhizoctonia solani* causing root rot of *Vigna unguiculata*. Journal of Integrative Agriculture, 18: 2072-2079.
- Wang, C. and Zhuang, W.Y. 2020. Carbon metabolic profiling of *Trichoderma* strains provides insight into potential ecological niches. Mycologia 112: 213-223.
- Webster, J. and Weber, R. 2007. Introduction to fungi. Cambridge University Press 1-841.
- Weindling, R. 1932. *Trichoderma lignorum* as a parasite of other soil fungi. Phytopathology, 22: 837-845.
- Woo, S.L., Ruocco, M., Vinale, F., Nigro, M., Marra, R., Lombardi, N., Pascale, A., Lanzuise, S., Manganiello, G. and Lorito, M. 2014. *Trichoderma*-based products and their widespread use in agriculture. The Open Mycology Journal, 8: 71-126.

ارزیابی تنوع ناحیه ژن Chit18-5 در گونههای تریکودرما برای استراتژیهای کنترل زیستی موثر

آیدین حسنزاده و محمد علی تاجیک قنبری 🖾

گروه گیاهپزشکی، دانشکده علوم زراعی، دانشگاه علوم کشاورزی و منابع طبیعی ساری، ساری، ایران

چکیده: گونههای تریکودرما به عنوان عوامل زیستی برای کنترل بیماریهای گیاهی، بطور گسترده مورد استفاده قرار می گیرند. کیتینازها در میکوپارازیتیسم و دفاع در برابر سایر قارچها و بندپایان، بسیار مهم هستند. در این مطالعه، ۴۱ توالی اسیدآمینه مربوط به ناحیه ژن 5-*Chitl8* از چهار بخش و ۱۵ گونه تریکودرما، بررسی شد. شناسایی و ارزیابی دومینها، موتیفهای حفاظتشده و درخت تبارزایی با استفاده از پایگاه داده InterProScan، ابزار COBALT، نرمافزار ISS V5.51، الگوریتم MEME V5.51 و نرمافزار MEGA11، صورت گرفت. نتایج نشان داد که ناحیه ژنی مورد بررسی میتواند به طور موثر گونههای مختلف تریکودرما را از یکدیگر تفکیک نماید و ابزار موثری برای بهینهسازی استراتژیهای کنترل زیستی است. این مطالعه، پتانسیل بررسی تنوع ژنتیکی را به عنوان ابزاری برای شناسایی راه حلهای جدید برای مدیریت آفات و بیماریها در کشاورزی، برجسته مینماید. موتیفهای احتمالی پروتئینهای کیتینازی شناسایی شده در این مطالعه ممکن است در فعالیتهای آنتاگونیستی تریکودرما نقش

كلمات كليدى: تريكودرما، ناحيه ژن 5-Chit18، كنترل زيستى، درخت تبارزايي، تنوع ژنتيكى

مکاتبه کننده: محمد علی تاجیک قنبری Email: m.tajick@gmail.com تاریخ دریافت: ۱۴۰۲/۷/۶ تاریخ پذیرش: ۱۴۰۲/۱۱/۲۳