

The current status of Saprolegniales in Iran: calling mycologists for better taxonomic and ecological resolutions

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Abstract: Saprolegniales have been studying for nearly 150 years and have been recognized mainly as freshwater animal pathogens. Similar to the global trend, studies on Saprolegniales in Iran have mainly focused on their pathological aspects. Therefore, this review discusses the state of the art of Saprolegniales studies in Iran and pinpoints present deficiencies. More than 80% of Iranian studies have examined the impact of various plant extracts and other plant compounds on Saprolegnia parasitica, responsible for causing deadly diseases on fish species. On the other hand, recent taxonomic and ecological reports on Saprolegniales have addressed this topic in a more abstract manner. Finally, we give recommendations to how to more systematically study Saprolegniales in Iran. This review calls mycologists in Iran and elsewhere to study Saprolegniales more seriously and in a more coordinated manner.

Key words: Disease management, Diversity, Ecology, *Oomycota*, Systematics, Pathology

INTRODUCTION

The order Saprolegniales is one of the major lineages of oomycetes. Although members of Saprolegniales have been traditionally known as water molds, they adopt various lifestyles in both aquatic and terrestrial environments. From at least 13 established genera divided into three families, most are freshwater saprophytes frequently found in various freshwater ecosystems populating organic matter, mainly in the littoral zone. Among all freshwater taxa, the order also holds some of the most notorious aquatic animal pathogens, such as Saprolegnia parasitica Coker and Aphanomyces astaci Schikora which threaten the stability of freshwater ecosystems by causing highly

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destructive diseases (Hussein et al. 2013, Van Den Berg 2013, Svoboda et al. 2017). The fact that genera of Saprolegniales are rarely found in the terrestrial environment does not necessarily mean that nonaquatic members are unimportant. Aphanomyces euteiches Drechsler, for example, is one of the significant threats to a variety of legumes (Levenfors et al. 2003, Gaulin et al. 2007). More recently, the contribution of Saprolegniales in freshwater food webs has also attracted attention. It has been shown that Saprolegniales interact with different trophic levels of food webs through parasitic and/or saprophytic lifestyles, thus, probably altering its energy flow.

Although Saprolegniales, as stated above, constitute an oomycetes order of high commercial and ecological relevance, its diversity, pathogenicity, and ecological involvements are still unknown in many regions of the world. Therefore, in this study, we aim to evaluate the current status of Saprolegniales in Iran as one of the least studied countries in the world. Research areas which are most in the focus in Iran will be discussed and the latest discoveries will be reviewed accordingly. Then, we will touch upon some of the most enduring challenges faced by scholars who study Saprolegniales. Finally, some recommendations for future investigations will be presented. This review is important for Iranian mycologists as it aims to stimulate more systematic and ecologically-driven lines of research for Saprolegniales in Iran.

Saprolegniales are successful Saprophytes

Although Saprolegniales are mainly considered as "bad microorganisms" inducing various endemic diseases in aquatic animals (discussed in the following sections), most members impact their interconnected habitats positively. Freshwater ecosystems receive a large quantity of organic matter, mainly originating from the terrestrial environment. For example, a constant wave of plant and animal debris is being discharged into the freshwater environment, which peaks during autumn. Immediately, plant debris is being attacked mainly by freshwater fungi and oomycetes, with Saprolegniales in a leading rule. Recently, a few studies have shown that Saprolegniales colonize and then get engaged in the process of transformation and degradation of plant and animal debris deposited on the sediment surface of the

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littoral zone. Although studies suggest that *Saprolegniales* and fungi both engage in the degradation of cellulose-, hemicellulose-, and chitinbased compounds, interestingly, they behave differently toward lignin- and lignin-like molecules. *Saprolegniales* lack the ligninolytic capability, which points to an ecological partitioning between them and true fungi, as robust lignin degraders (Sigoillot et al. 2012, Armand et al. 2020) (Fig. 1).

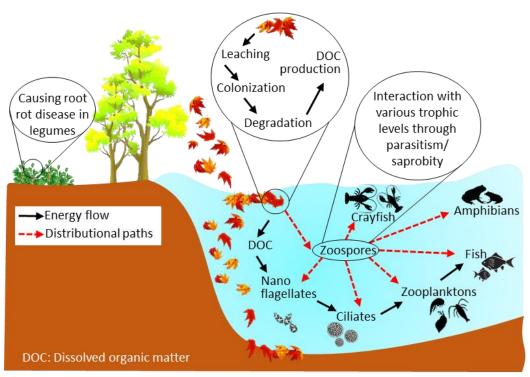


Fig. 1. Depicting contributions of Saprolegniales in both aquatic and terrestrial environments.

Pathogenicity of Saprolegniales

The order Saprolegniales is an exceptional lineage among other oomycete lineages and other eukaryotic life forms as it encompasses both plant and animal obligate parasites. Such a feature naturally implies a very complicated evolutionary history of the Saprolegniales, resulting in the acquisition of two completely contrasting lifestyles, i.e., plant and animal parasitism, among taxa (Jiang et al. 2013). Crayfish plague, caused by A. astaci, is a severe disease widely distributed across European countries (Holdich et al. 2003). In fact, A. astaci is suggested as the most dangerous threat to local European crayfish communities. Another syndrome caused by the Aphanomyces species complex is called epizootic granulomatous aphanomycosis (EGA; previously known as aphnomycosis) causes life-threatening lesions in many fish species (Kamilya et al. 2014). The next is Saprolegnia parasitica which is probably a poster child for Saprolegniales. It causes saprolegniosis against many fish and amphibian species. The symptom of saprolegniosis includes cotton-like growths on the skin and gills, depigmented

skin, and sunken eyes. However, as stated above, *Saprolegniales*, or the genus *Aphanomyces* to be more specific, contains a severe plant pathogen too. *Aphanomyces euteiches* causes aphanomyces root rot or common root rot of legumes such as pea (*Pisum sativum* L.) and alfalfa (*Medicago sativa* L.), as two of the main commercial crops mainly in North America, China, India, Australia, and some European countries (Wu et al. 2018).

The symptoms include chlorosis of leaflets in infected seedlings, gray and water-soaked roots and stems which turn light to dark brown, and stunted seedlings (Fig. 2). *Saprolegniales* are also thought to be potential parasites of protists such as cilliatesin freshwater ecosystems. In a few isolated cases, *Aphanomyces* spp. also cause substantial selective pressures in natural *Daphnia* spp. populations (Wolinska et al. 2008). Such a parasitic interaction is important because Daphnia is a keystone species in the freshwater microbial loop that plays an essential role in the energy flow being circulated among prokaryotic and eukaryotic communities (Cuenca et al. 2018) (Fig. 1).



Fig. 2. Symptoms caused by pathogenic *Saprolegniales*. (A) Aphanomyces root rot (*Aphanomyces cochlioides*) external symptoms on mature beet (Photograph by Oliver T. Neher, The Amalgamated Sugar Company, Bugwood.org), (B) and (C) Strong immune reaction, i.e., melanin formation, against infection by *Aphanomyces astaci* visualized as large melanized patches (arrows) on the joint of a chela, and also on a walking leg of *Austropotamobius pallipes* and *Pacifastacus leniusculus* (from Martin-Torrijo et al. 2017; 2018, with permission).

The current status of Saprolegniales in Iran

Saprolegniales are infrequently studied in Iran concerning their taxonomy and ecology, similar to other countries worldwide. We found nearly 60 scientific publications published in local and international peer-reviewed journals which dealt with a few aspects of *Saprolegniales'* biology. Most papers have studied the effects of plant extracts on *Saprolegnia parasitica*. Also, the identification and taxonomic position of isolated strains have been studied in some regions. Although ecological contributions of *Saprolegniales* in freshwater ecosystems have been recently received more attention, it is far from being satisfactory. The most important results from these studies will be highlighted in the following paragraphs.

Taxonomy of Saprolegniales in Iran

In general, providing a clear taxonomic description of *Saprolegniales* has rarely been conducted in Iran.

Nearly all strains have been isolated from fish species (Table 1). Mousavi et al. (2007; 2009) reported S. parasitica, S. mixta de Bary, S. monilifera de Bary, Achlya oblongata de Bary, and Brevilegnia sp. from affected eggs of the rainbow trout (Oncorhynchus mykiss) in Mazandaran Province. Saprolegnia parasitica, S. lapponica Gäum., S. ferax (Gruith.) Kütz., S. hypogyna (Pringsh.) Pringsh. and S. diclina Humphrey were also isolated as contaminants in rainbow trout eggs in Kermanshah Province (Shahbazian et al. 2010). In two other studies, Saprolegnia strains were isolated from rainbow trout eggs and broodstocks in three hatcheries in western Iran (Fadaeifard et al. 2011). Chiasi et al. (2012) and Azizi et al. (2014) also isolated Saprolegnia sp. from Caspian kutum (Rutilus frisii kutum) eggs and Caspian trout (Salmo trutta caspius) skinin Mazandaran Province, respectively. Based on physiological and molecular data, Ghiasi et al. (2013; 2014) also

characterized *Saprolegnia* sp. isolates from Persian sturgeon (*Acipenser persicus*), Caspian trout (*Salmo trutta caspius*), and rainbow trout (*O. mykiss*) and showed that some features such as repeated zoospore emergence in *Saprolegnia* could be correlated to the pathogenicity of strains. Additionally, *Saprolegnia* sp. strains were reported from Minnows and Carps in Golestan province (Sharifpour et al. 2014). Nekuie Fard et al. (2011), Khodadadi et al. (2014), and Ghorbani & Azadikhah 2019 considered *Saprolegnia* sp. and *Achlya* sp. strains as one of the reasons responsible for a massive decline in the population of the Danube crayfish (*Astacus leptodactylus*) as well as larval and adult Rainbow trout (*O. mykiss*) in West Azerbaijan Province. Oscar fish (*Astronotus ocellatus*) is another host with typical symptoms of saprolegniosis caused by *Achlya* sp. strains in Khuzestan Province (Peyghan et al. 2019).

 Table 1. List of Saprolegniales reported from Iran provinces.

 Taxon
 Substrates*

Taxon	Substrates*	Location	Reference
Achlya americana Humphrey			
	Decayed twigs, leaves, woods and infected spawns	Fars and Kermanshah	Bolboli & Mostowfizadeh- Ghalamfarsa 2019
A. bisexualis Coker and Couch	<i>Typha</i> spp.	Gilan (Anzali lagoon)	Masigol et al. 2020
<i>Achlya oblongata</i> de Bary	Rainbow trout eggs	Mazandaran	Mousavi et al. 2007,
A able and			2009
Achlya sp.	Galician crayfish	West Azerbaijan	Ghorbani & Azadikhah 2019
	Oscar fish	Khuzestan	Peyghan et al. 2019
	Rainbow trout eggs	Mazandaran	Mousavi et al. 2007, 2009
	<i>Typha</i> spp.	Gilan (Anzali lagoon)	Masigol et al. 2020
<i>Brevilegnia</i> sp.	Rainbow trout eggs	Mazandaran	Mousavi et al. 2007, 2009
Dictyuchus sp.			2009
Saprolegnia aff. australis R.F. E	<i>Typha</i> spp. Elliott	Gilan (Anzali lagoon)	Masigol et al. 2018
	Decayed twigs, leaves, woods and infected spawns	Fars and Kermanshah	Bolboli & Mostowfizadeh- Ghalamfarsa 2019
S. anisospore de Bary	-		
Construction Constitution & Konstruction	<i>Typha</i> spp.	Gilan (Anzali lagoon)	Masigol et al. 2020
S. anomala Gandhe & Kurne	Decayed twigs, leaves, woods and infected spawns	Fars and Kermanshah	Bolboli & Mostowfizadeh- Ghalamfarsa 2019
S. diclina Humphrey	D 1 1		CI 11 1 0 040
	Rainbow trout eggs <i>Typha</i> spp.	Kermanshah Gilan (Anzali lagoon)	Shahbazian et al. 2010 Masigol et al. 2020
S. ferax (Gruith.) Kütz.	i ypna spp.	Onan (Anzan tagoon)	Wasigoi et al. 2020
	Decayed twigs, leaves, woods and infected	Fars and Kermanshah	Bolboli & Mostowfizadeh- Ghalamfarsa 2019
	spawns Rainbow trout eggs <i>Typha</i> spp.	Kermanshah Gilan (Anzali logoon)	Shahbazian et al. 2010 Masigol et al. 2020
S. hypogyna (Pringsh.) Pringsh			
S. lapponica Gäum	Rainbow trout eggs	Kermanshah	Shahbazian et al. 2010
	Rainbow trout eggs	Kermanshah	Shahbazian et al. 2010

Taxon	Substrates*	Location	Reference
S. mixta de Bary			
	Decayed twigs, leaves,	Fars and Kermanshah	Bolboli &
	woods and infected		Mostowfizadeh-
	spawns		Ghalamfarsa 2019
	Rainbow trout eggs	Mazandaran Province	Mousavi et al. 2007
			2009
<i>S. monilifera</i> de Bary			
	Rainbow trout eggs	Mazandaran Province	Mousavi et al. 2007
			2009
S. parasitica Coker			
	Rainbow trout eggs	Kermanshah	Shahbazian et al. 2010
	<i>Typha</i> spp.	Gilan (Anzali lagoon)	Masigol et al. 2020
<i>Saprolegnia</i> sp.			
	Bighead carp	Golestan	Sharifpour et al. 2014
	Caspian kutum eggs	Mazandaran	Chiasi et al. 2012
	Caspian trout	Mazandaran	Chiasi et al. 2013, 2014
	Caspian trout skin	Mazandaran	Azizi et al. 2014
	Danube crayfish	West Azerbaijan	Nekuie Fard et al. 2014
	Eurasian carp	Golestan	Sharifpour et al. 2014
	Galician crayfish	West Azerbaijan	Ghorbani & Azadikhah 2019
	Grass carp	Golestan	Sharifpour et al. 2014
	Persian sturgeon	Mazandaran	Chiasi et al. 2013, 2014
	Rainbow trout brood	Farms in western Iran	Fadaeifard et al. 2011
	stock and eggs		
	Rainbow trout eggs	Mazandaran	Mousavi et al. 2007,
			2009; Chiasi et al.
			2013, 2014
	Rainbow trout	West Azerbaijan	Khodadadi et al. (2014)
		(Hasalno dam)	· · · · · · · · · · · · · · · · · · ·
	Silver carp	Golestan	Sharifpour et al. 2014

	a
Table	Continued.

Bighead carp (H. nobilis); Caspian kutum (Rutilus frisii kutum); Caspian trout (Salmo trutta caspius); Danube crayfish (Astacus leptodactylus); Eurasian carp (Cyprinus carpio); Galician crayfish (Astacus leptodactylus); Grass carp (Ctenopharyngodon idella); Oscar fish (Astronotus ocellatus); Persian sturgeon (Acipencer persicus); Rainbow trout (Oncorhynchus mykiss); Silver carp (Hypophthalmichthys molitrix)

As the studies mentioned above mainly focused on the pathogenic features of Saprolegniales, they rarely gave detailed morphological descriptions to identify strains in the species level accurately. Also, phylogenetic studies, i.e., using sequences to determine the relationship between strains, were almost lacking in all cases (Jeronimo et al. 2015). However, in three cases, a fair improvement has been obtained by giving precise morphometric features and running phylogenetic analysis using ITS and cox1 sequences. Several Dictyuchus, Saprolegnia, and Achlya spp. strains were isolated from decaying leaves of the dominant local vegetation (Typha spp.) collected from the Anzali lagoon in Northern Iran and then morphologically and phylogenetically characterized. Although Dictyuchus sp. strains were believed to be a new combination, it remained unclear due to the unavailability of suited reference sequences (Masigol et al. 2018; 2020). In another similar study, Bolboli and Mostowfizadeh-Ghalamfarsa (2019)isolated. described and identified Saprolegniales strains from brown decayed twigs, leaves, woods and infected spawns in Fars and Kermanshah Provinces (Southern and Western Iran). In addition to morphologically and phylogenetically identification of six Saprolegnia and one Achlya species, several strains were not related to any of the previously reported taxa, promising two new Saprolegnia species. In their study S. anomala Gandhe & Kurne, S. mixta, and A. americana Humphrey were found new to Iran's mycobiota.

The effect of plant extracts on Saprolegnia spp.

In parallel with getting more awareness toward diseases caused by Saprolegnia species, controlling strategies were studied to minimize damages to fishes and other aquatic animals. Previously, Malachite green was one of the most common chemicals for controlling saprolegniosis caused by Saprolegnia species. However, the application of Malachite green is currently prohibited in many countries due to its carcinogenic nature (Culp et al. 2006). Therefore,

scientists have been searching for less dangerous chemicals to control saprolegniosis. Examining safer chemicals, natural compounds such as plant extracts, and biological agents to control the disease have so far been the main research topics in Iran.

Most studies regarding disease management of S. parasitica started after Jalilpoor et al. (2006), who reported 7% to 22% mortality of Acipenser persicus (Persian sturgeon) eggs during mass incubation at a hatchery in Iran. Regarding plant extracts, several candidates have already been introduced. Mousavi et al. (2006) tested the inhibitory effect of Eucaliptus camaldolensis extract on S. parasitica leading to better protection of trout's (O. mykiss) eggs. In another study, S. parasitica showed a high sensitivity to methanolic extracts from the plant Citrullus colocynthis (Azizi 2012). Also, Sharifi et al. (2012) reported that a hydroalcoholic extract of the oak placenta could help to prevent Saprolegnia's growth compared to Malachite green. Zataria multiflora and Е. camaldolensis essential oils effectively treated S. parasitica-infected rainbow trout (O. mykiss) eggs in the aquaculture environment (Khosravi et al. 2012). Multiple other extracts have been proposed as inhibitors of S. parasitica: feverfew (Tanacetum parthenium (L.) Sch. Bip.) and horse mint (Mentha longifolia (L.) Huds.) (Ghasemi Pirbalouti et al. 2009), cashew (Anacardium occidentale L.) (Akhlaghi and Bahaedini 2012), common rue (Ruta graveolens L.) (Hashemi Karouei et al. 2012), Artemisia sp. (Firouzbakhsh et al. 2014), common nettle (Urtica dioica L.) (Firouzbakhsh et al. 2015, Alishah et al. 2019, Mehrabi et al. 2020), myrtle (Myrtus communis L.) (Salimian et al. 2015), peppermint (Mentha balsamea L.) (Hooshangi et al. 2016, Adel et al. 2017), cumin (Cuminum cyminum L.), watling street thistle (Eryngium campestre L.) (Adel et al. 2020), garden thyme (Thymus vulgaris), oregano (Origanum vulgare L.) (Golchin et al. 2017, Pazira 2017, Shahrani et al. 2018), aloe vera (Aloe vera (L.) Burm f.) (Mehrabi et al. 2017, Mehrabi et al. 2019), clove (Syzygium aromaticum (L.) Merr. & L.M.Perry), lavender (Lavandula spica L.) (Jookar et al. 2019), chamomile (Matricaria chamomilla L.) (Amiri and Meshkini 2019), ajwain (Trachyspermum ammi (L.) Sprague ex Turrill) (SavadKouhi et al. 2021), and lemon balm (Melissa officinalis L.) (Ggonani and Taghavizad 2021).

To a lesser extent, the application of other chemicals and biological agents such as bacteria have also been studied in several cases. For example, Abtahi et al. (2006) suggested that potassium permanganate is safer than Malachite green and formalin at a given relative concentration to control saprolegniosis in *Acipenser persicus* egg incubation. Other chemicals such as sodium chloride (Khodabandeh and Abtahi 2006), powdered silver zeolite (Johari et al. 2014), fructooligosaccharide (Firouzbakhsh et al. 2014a), colloidal silver nanoparticles (AgNPs) (Johari et al.

2015, Shokouh Saljoghi et al. 2018), zinc oxide nanoparticles (Sedighi et al. 2015), copper nanoparticles (Kalatehjari et al. 2015), and stabilized hydrogen peroxide (Salah et al. 2021) have shown to be effective as well. Additionally, biological agents such as Pseudomonas huorescence (Akhlaghi and Bahaedini 2012) and Pseudomonas aeruginosa (Moghaddam et al. 2012) have an inhibitory effect against S. parasitica. Nevertheless, we need to address the question of the practicality of the abovementioned compounds because studies conducted in Iran have not yet entered the application phase, making the largescale implementation of these compounds impossible. In contrast, supported by a better understanding of molecular mechanisms, other studies have successfully proven the industrial applicability owing to their immunostimulatory and anti-Saprolegnia activities (Caruana et al. 2012, Mostafa et al. 2020, Tandel et al. 2021).

Ecology of Saprolegniales in Iran

Ecology of Saprolegniales is the least studied aspect of this order in Iran and other countries. There is a long history of isolating Saprolegniales taxa from plant and animal debris, mainly from freshwater ecosystems across the world (Grossart et al. 2020). However, very little effort has been made to understand how Saprolegniales taxa interact with organic matter. Masigol et al. (2019) postulated that both fungi and Saprolegniales are involved in the degradation and transformation of organic matter as they both occupy organic matter populated habitats and share similar physiological features. They isolated Saprolegniales from Anazali lagoon and applied plate assay methods and liquid chromatography-organic carbon-organic nitrogen detection (LC-OCD-OND) experiments to determine whether/how/to what extent Saprolegniales are involved in carbon cycling in freshwater ecosystems. The authors conclude that Saprolegniales functions differently than true fungi due to their different enzymatic capabilities. However, the studies were limited to Dictyuchus and Achlya species only and hence this topic awaits further exploration. Also, the study revealed that Saprolegniales and fungi utilize low molecular weight compounds at different rates pointing to different ecological niches.

Challenges and recommendations for future investigations

As discussed in this review, the studies of *Saprolegniales* suffers from several shortcomings which can be also observed in studies of other oomycetes and fungi. Therefore, we recommended the following lines of research to compensate for such deficiencies and highlight the contribution of *Saprolegniales* to the diversity and functionality of various ecosystems (Fig. 3).

Taxonomic research

Firstly, extensive classic and modern taxonomic studies require a tremendous amount of work as identifying newly reported strains is largely hampered

by: I) biased sampling mainly from animals such as fish, crayfish, and amphibian species in Europe and North America, II) lack of precise morphometric features, III) too much emphasis on ITS (internal transcribed spacers of rDNA) sequences, IV) many non-studied regions, V) presence of too many missassigned taxa, and VI) handling Saprolegniales strains mainly by non-mycologists. Therefore, we recommend intensifying sampling frequency from unexplored regions in the first step. Iran, for example, will be a promising sampling region due to a large variety of internationally and ecologically important freshwater ecosystems such as Anzali lagoon, Urmia lake, Gavkhuni Wetland, Karun River, and other numerous freshwater-related sites. It is expected to find many more previously unknown strains and explore the diversity of Saprolegniales (see Rezakhani et al. 2019 for fungal diversity). It will be vital to provide in-depth morphological descriptions and analyses to render them comparable to other studies and delineate taxa boundaries more accurately. Also, multigene phylogenetic studies should be given priority as the sole application of ITS sequences has not been as effective as expected to separate Saprolegniales taxa. Other regions such as cox1, cox2, and LSU have been promising a better resolution to phylogenetic relationships among Saprolegniales (Rocha et al. 2016; 2018). By following the above-mentioned activities, not only will currently unknown taxa be revised/removed, but a new combination will appear, leading to a better appreciation of Saprolegniales diversity in various Iranian habitats. We are confident of such an improvement in Saprolegniales taxonomy as adopting in-depth morphological and multigene phylogenetic studies in its sister orders, Peronosporales and Pythiales, has completely changed our understanding of their hidden diversity (Kageyama et al. 2014, Scanu et al. 2021).

Diversity research

Studies on *Saprolegniales* have always been limited to culture-dependent methods, making large-scale diversity-based studies very hard to accomplish. Regardless of tremendous efforts to compensate culture-dependent methods' constraints in oomycetes (including *Saprolegniales*) and fungi (Põlme et al. 2020), high throughput sequencing (HTS) techniques can more efficiently and systematically determine the diversity of *Saprolegniales* in any given environment based upon millions of sequences. Unfortunately, not even one single study has ever studied the diversity of freshwater *Saprolegniales* using HTS, metagenomics, and long read sequencing techniques. That is how other important questions can be addressed (Singer et

al. 2016). For instance, it will be much more feasible to check whether *Saprolegniales* communities start to shift as biotic and abiotic parameters of the environment are changed. In other words, finding any correlation between *Saprolegniales* communities and various ecosystem features will be possible. However, high-throughput sequencing techniques are expensive, time-consuming, and mostly unavailable in Iran. Indeed, more international collaborations make largescale comparative studies of various Iranian freshwater ecosystems possible.

Disease management research

Although it has been shown that most plant extracts examined in Iranian studies are effective against Saprolegnia parasitica, no studies have gone tested the practicality of using such extracts in an industrial scale to control saprolegniasis. Therefore, the application of these active compounds against saprolegniasis must be given priority to avoid economic damages of S. parasitica in natural and farmed populations. Additionally, other diseases caused by Saprolegniales should be studied as well. For instance, we have an absolute lack of knowledge concerning the distribution and pathogenicity of A. astaci strains in Iran. This should be determined promptly, as endemic Iranian crayfish populations are distributed across the country and they are likely to get infected by virulent strains of A. astaci, with severe consequences for interconnected ecosystems. The same is valid with epizootic granulomatous aphanomycosis (EGA) in which the Aphanomyces species complex could attach fish species and eradicate local populations.

Ecological research

It has already been established that eukaryotes (fungi in particular) and their interaction with other life forms significantly contribute to global carbon cycling in both terrestrial and aquatic environments. It is predicted that Saprolegniales contributions are mainly facilitated through two separate paths. Some taxa have the potential to alter the energy flow at different trophic levels of food webs (De Souza et al. 2014). In addition to their impact on fish, crayfish, and amphibian populations, Saprolegniales are believed to interact with keystone planktonic organisms, resulting in altering the established equilibrium between keystone species in food webs (Buaya et al. 2019). Therefore, it is suggested to conduct pathosystembased studies to determine how significantly Saprolegniales alter the genetic composition of different planktonic populations. Meanwhile, new species and genera of Saprolegniales will likely to be explored as they are rarely isolated from planktons.

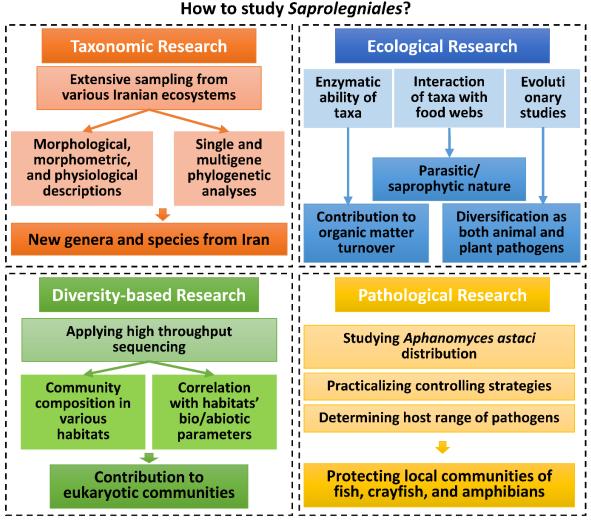


Fig. 3. Proposed avenues of Saprolegniales research in Iran.

An outlook

In this article, the current status of Saprolegniales in Iran was reviewed. Firstly, we showed that although taxonomical studies have reported several Saprolegniales taxa mainly from freshwater ecosystems, many Iranian water-dependent ecosystems have remained unexplored in terms of the diversity of Saprolegniales. Additionally, it was revealed that studying saprolegniosis management using plant extracts and other compounds has been the most focused research area in Iran for the last two decades with not much effort to make them practical for large-scale use in the fisheries industry. Finally, understanding the ecological contributions of Saprolegniales was spotted as the least studied research area, in spite of a few recent discoveries. Accordingly, we suggested five Saprolegnialesoriented research lines that Iranian mycologists are called to focus on, including I) highlighting practical disease management strategies (for both saprolegniosis and crayfish plague) with respect to ecological features

of *Saprolegniales* pathogens, II) conducting extensive sampling from unexplored sites, III) providing indepth morphological and phylogenetic characterization, and IV) evaluating the impact of *Saprolegniales* on the degradation of organic matter and its interactions with different trophic levels of any given food web, especially in freshwater ecosystems.

REFERENCES

- Abtahi B, Shafiezade P, Nazari RM, Rassouli A. 2006. Comparison of antifungal therapeutic indices of formalin, malachite green, and potassium permanganate in treating Persian Sturgeon eggs. Journal of Applied Ichthyology 22(1): 291–293.
- Adel M, Dadar M, Zorriehzahra MJ, Elahi R, Stadtlander T. 2020. Antifungal activity and chemical composition of Iranian medicinal herbs against fish pathogenic fungus, *Saprolegnia parasitica*. Iranian Journal of Fisheries Sciences 19(6): 3239–3254.

- Armand A, Khodaparast SA, Masigol H, Grossart HP. 2020. Screening of fungal strains isolated from dead insects for production of plant litter-degrading enzymes. Mycologia Iranica 7(1): 155–61.
- Moghaddam A, Hajimoradloo A, Ghiasi M, Ghorbani R. 2012. In vitro inhibition of growth in Saprolegnia sp. isolated from the eggs of Persian sturgeon Acipenser persicus (Pisces: Acipenseriformes) by *Pseudomonas aeroginosa* (PTCC: 1430). Caspian journal of environmental sciences 11(2): 233–240.
- Akhlaghi A, Bahaoodini AA. 2012. Comparison of several treatment methods against saprolegniosis in rainbow trout fertilized eggs. Veterinary Researches & Biological Products 25(1): 18–24.
- Alishah N, Firouzbakhsh F, Mehrabi Z. 2019. Bathing effects of nettle (*Urtica dioica*) hydroalcoholic extract on immunological and hematological indices in rainbow trout (*Oncorhynchus mykiss*) infected with *Saprolegnia* fungal. Iranian Scientific Fisheries Journal 28(2): 55–67.
- Amiri H, Meshkini S. 2019. Antifungal effects of Achillea officinolis and Matricaria chamomilla plant extracts on control infection of rainbow trout eggs by Saprolegnia parasitica. Journal of Veterinary Microbiology 15(1): 97–107.
- Azizi I, Hosseinifard SM, Rohi S, Moghtader H. 2014. Investigation and identification of fungal infection of Caspian salmon (Salmo trutta caspius) in fish farms of Mazandaran province. Animal biology 6(3): 45–53.
- Azizi IG, Fard MH, Tahmasbipour S. 2012. The Effect of aquatic and alcoholic extracts of *Citrullus colocynthis* on growth of the *Saprolegnia parasitica*. World Journal of Fish and Marine Sciences 4: 258–62.
- Bolboli Z, Mostowfizadeh-Ghalamfarsa R. 2019. Isolation and identification of aquatic saprolegnious stramenopiles in Fars and Kermanshah provinces (west of Iran). Rostaniha 20(1): 14–28.
- Buaya A, Kraberg A, Thines M. 2019. Dual culture of the oomycete *Lagenisma coscinodisci* Drebes and Coscinodiscus diatoms as a model for plankton/parasite interactions. Helgoland Marine Research 73(1): 1–6.
- Caruana S, Yoon GH, Freeman MA, Mackie JA, Shinn AP. 2012. The efficacy of selected plant extracts and bioflavonoids in controlling infections of *Saprolegnia australis* (Saprolegniales; Oomycetes). Aquaculture 358: 146–154.
- Culp SJ, Mellick PW, Trotter RW, Greenlees KJ, Kodell RL, Beland FA. 2006. Carcinogenicity of malachite green chloride and leucomalachite green in B6C3F1 mice and F344 rats. Food and Chemical Toxicology 44(8): 1204–1212.
- Cuenca Cambronero M, Marshall H, De Meester L, Davidson TA, Beckerman AP, Orsini L. 2018. Predictability of the impact of multiple stressors on

the keystone species Daphnia. Scientific Reports 8(1): 1–11.

- De Souza JI, Gleason FH, Ansari MA, Lastra CC, Garcia JJ, Pires-Zottarelli CL, Marano AV. 2014. Fungal and oomycete parasites of Chironomidae, Ceratopogonidae and Simuliidae (Culicomorpha, Diptera). Fungal Biology Reviews 28(1): 13–23.
- Fadaeifard F, Bahrami H, Rahimi E, Najafipoor A. 2011. Freshwater fungi isolated from eggs and broodstocks with an emphasis on Saprolegnia in rainbow trout farms in west Iran. African Journal of Microbiology Research 5(22): 3647–3651.
- Firouzbakhsh F, Zolfaghari A, Mehrabi Z, Khalesi MK. 2015. In vitro antifungal activity of Nettle (*Urtica dioica*) and Basil (*Ocimum basilicum*) extracts on *Saprolegnia parasitica*. Journal of Animal Environment 7(3): 211–216.
- Firouzbakhsh F, Afsarian MH, Hooshangi S, Badali H. 2014. Evaluation of in vitro antifungal activity of *Foeniculum, Achillea, Satureja, Cinnamomum* and *Artemisia* against *Saprolegnia parasitica*. Arak Medical University Journal 17(5): 60–69.
- Firouzbakhsh F, Mehrabi Z, Heydari M, Khalesi MK, Tajick MA. 2014a. Protective effects of a synbiotic against experimental *Saprolegnia parasitica* infection in rainbow trout (*Oncorhynchus mykiss*). Aquaculture Research 45(4): 609–618.
- Gaulin E, Jacquet C, Bottin A, Dumas B. 2007. Root rot disease of legumes caused by *Aphanomyces euteiches*. Molecular Plant Pathology 8(5): 539– 548.
- Ghasemi Pirbalouti A, Taheri M, Raiesi M, Bahrami H, Abdizadeh R. 2009. In vitro antifungal activity of plant extracts on *Saprolegnia parasitica* from cutaneous lesions of rainbow trout (Oncorhynchus mykiss) eggs. Journal of Food, Agriculture & Environment 7(2): 94–96.
- Ghiasi M, Khosravi AL, Soltani M, Sharifpour I, Binaii M, Ebrahimzadeh Mosavi H, Bahonar A. 2014. Evaluation of physiological aspects and molecular identification of *Saprolegnia* isolates from rainbow trout (*Oncorhynchus mykiss*) and Caspian trout (*Salmo trutta caspius*) eggs based on RAPD–PCR. Iranian fisheries science research institute 22(4): 82–93.
- Ghiasi M, Shokri H, Binai M, Farabi SMV, Saeidi AA. 2012. Identification and comparison of the frequency of fungal agents isolated from the eggs of whitefish (Rutilus frisii kutum) in Mazandaran province. Journal of aquaculture development 6(1): 79–90.
- Ghiasi M, Khosravi AR, Soltani M, Binaii M, Shokri H, Tootian Z, Rostamibashman M, Ebrahimzademousavi H. 2010. Characterization of Saprolegnia isolates from Persian sturgeon (Acipencer persicus) eggs based on physiological and molecular data. Journal de mycologie médicale 20(1): 1–7.

- Ghorbani E, Azadikhah D. 2019. Identification and determination of prevalence of saprophytic fungi in the larval stage of the rainbow trout (*Oncorhynchus mykiss*) in hatcheries of west Azarbaijan province. Veterinary clinical pathology the quarterly scientific journal 13(49): 91-99.
- Ggonani S, Taghavizad R. 2021. The effect of methanolic, aqueous and hydroalcoholic extracts of peacock and lemongrass on fish (Saprolegnia) in comparison with the effect of antibiotics. Aquaculture development (biological sciences) 14(4): 83–91.
- Golchin MA, Jarkani Bayat S, Tarhami M. 2017. Experimental evaluation of antifungal effect of thyme extract (*Thymus vulgaris*) and marjoram (*Origanum vulgare*) on *Saprolegnia fungi* isolated from rainbow trout in Shahid Motahari breeding complex in Yasou. New technologies in aquaculture development (fisheries) 10(4): 11–18.
- Grossart HP, Hassan EA, Masigol H, Arias-Andres M, Rojas-Jimenez K. 2020. Inland water fungi in the Anthropocene: Current and future perspectives. In: The Encyclopedia of Inland Waters, (Kendra Cheruvelil eds.): Elsevier Inc, Netherlands. https://doi.org/10.1016/B978-0-12-819166-8.00025-6.
- Hashemi Karouei SM, Sadeghpour Haji M, Gholampour Azizi I. 2012. Isolation of Saprolegnia and the influence of root ethanolic extract of *Ruta graveolens* on Saprolegnia spp growth. International Journal of Bioscience, Biochemistry and Bioinformatics 2(1): 64–67.
- Holdich DM, Sibley PJ. 2003. Crayfish in European overview of taxonomy, legislation, distribution, and crayfish plague outbreaks. Management & conservation of crayfish. 8(5): 15–34.
- Hooshangi S, Firouzbakhsh F, Badalii H. 2016. Evaluation of Invitro Antifungal Activity of Mentha Piperita and Carum Capticum against Saprolegnia Parasitica. Journal of Veterinary Microbiology 12(2): 47–55.
- Hussein MA, Hassan WH, Mahmoud MA. 2013. Pathogenicity of *Achlya proliferoides* and *Saprolegnia diclina* (Saprolegniaceae) associated with Saprolegniosis outbreaks in cultured Nile tilapia (Oreochromis niloticus). World Journal of Fish and Marine Sciences 2: 188–193.
- Jeronimo H, Rocha SC, Goncalves DR, Boro MC. 2015. Achlya catenulata sp. nov., a new Saprolegniales (Oomycetes, Straminipila) from Brazilian mangrove swamp. Phytotaxa 212(3): 221–228.
- Jalilpoor J, Shenavar Masouleh A, Masoumzadeh M. 2006. Fungal flora in *Acipenser persicus* eggs with particular emphasis on *Saprolegnia* sp. (Oomycetes) and mortality during mass incubation at the Shahid Beheshti hatchery. Journal of Applied Ichthyology 22: 265–8.

- Jiang RH, de Bruijn I, Haas BJ, Belmonte R, Löbach L, Christie J, van den Ackerveken G, Bottin A, Bulone V, Díaz-Moreno SM, Dumas B. 2013. Distinctive expansion of potential virulence genes in the genome of the oomycete fish pathogen *Saprolegnia parasitica*. PLoS genetics 9(6): e1003272.
- Johari SA, Kalbassi MR, Soltani M, Yu IJ. 2015. Study of fungicidal properties of colloidal silver nanoparticles (AgNPs) on trout egg pathogen, *Saprolegnia* sp. International Journal of Aquatic Biology (3): 191–198.
- Johari SA, Kalbassi MR, Yu IJ. 2014. Inhibitory effects of silver zeolite on in vitro growth of fish egg pathogen, Saprolegnia sp. Journal of Coastal Life Medicine 2(5): 357–361.
- Jookar Z, Gholipour Kanani H, Jafarian H, Taheri Mirghaed A. 2019. A comparative study of the in vitro antimicrobial and antifungal effect of *Trachyspermum ammi*, *Lavandula angustifolia* and *Eugenia aromatica* essence on rainbow trout Some pathogen. Journal of applied ichthyological research 6(4): 109–126.
- Kageyama K. 2014. Molecular taxonomy and its application to ecological studies of *Pythium* species. Journal of general plant pathology 80(4): 314–326.
- Kalatehjari P, Yousefian M, Khalilzadeh MA. Assessment of antifungal effects of copper nanoparticles on the growth of the fungus *Saprolegnia* sp. on white fish (*Rutilus frisii kutum*) eggs. 2015. The Egyptian Journal of Aquatic Research 41(4): 303–306.
- Kamilya D, Baruah A. 2014. Epizootic ulcerative syndrome (EUS) in fish: history and current status of understanding. Reviews in fish biology and fisheries (1): 369–380.
- Khodabandeh S, Abtahi B. 2006. Effects of sodium chloride, formalin and iodine on the hatching success of common carp, *Cyprinus carpio*, eggs. Journal of Applied Ichthyology 22(1): 54–56.
- Khodadadi A, Arabzadeh P, Rasouli S, Moradpour A, Abedian Amiri A. 2014. Investigation of the causes of rainbow trout losses in Cage Culture farms (Hasanloo Dam) Hasanlu Dam, West Azerbaijan Province. Pathology of veterinary clinic (Tabriz Veterinary) 8(2): 461–472.
- Khosravi AR, Shokri H, Sharifrohani M, Mousavi HE, Moosavi Z. 2012. Evaluation of the antifungal activity of Zataria multiflora, Geranium herbarium, and Eucalyptus camaldolensis essential oils on Saprolegnia parasitica–infected rainbow trout (Oncorhynchus mykiss) eggs. Foodborne Pathogens and Disease (7): 674–679.
- Levenfors JP, Wikström M, Persson L, Gerhardson B. 2003. Pathogenicity of *Aphanomyces* spp. from different leguminous crops in Sweden. European Journal of Plant Pathology 109(6): 535–543.

- Malherbe W, Christison KW, Wepener V, Smit NJ. 2019. Epizootic ulcerative syndrome–First report of evidence from South Africa's largest and premier conservation area, the Kruger National Park. International Journal for Parasitology: Parasites and Wildlife 10: 207–210.
- Martín-Torrijos L, Kawai T, Makkonen J, Jussila J, Kokko H, Diéguez-Uribeondo J. 2018. Crayfish plague in Japan: A real threat to the endemic *Cambaroides japonicus*. PLoS One 13(4): e0195353.
- Martín-Torrijos L, Campos Llach M, Pou-Rovira Q, Diéguez-Uribeondo J. 2017. Resistance to the crayfish plague, *Aphanomyces astaci* (*Oomycota*) in the endangered freshwater crayfish species, *Austropotamobius pallipes*. PLoS One. 2017 Jul 27;12(7): e0181226.
- Masigol H, Woodhouse JN, van West P, Mostowfizadeh-Ghalamfarsa R, Rojas-Jimenez K, Goldhammer T, Khodaparast SA, Grossart HP. 2021. Phylogenetic and Functional Diversity of Saprolegniales and Fungi Isolated from Temperate Lakes in Northeast Germany. Journal of Fungi 7(11): 968.
- Masigol H, Khodaparast SA, Mostowfizadeh-Ghalamfarsa R, Rojas-Jimenez K, Woodhouse JN, Neubauer D, Grossart HP. 2020. Taxonomical and functional diversity of Saprolegniales in Anzali lagoon, Iran. Aquatic Ecology 54(1): 323–336.
- Masigol H, Khodaparast SA, Woodhouse JN, Rojas-Jimenez K, Fonvielle J, Rezakhani F, Mostowfizadeh-Ghalamfarsa R, Neubauer D, Goldhammer T, Grossart HP. 2019. The contrasting roles of aquatic fungi and oomycetes in the degradation and transformation of polymeric organic matter. Limnology and Oceanography 64(6): 2662–2678.
- Masigol H, Khodaparast SA, Mostowfizadeh-Ghalamfarsa R, Mousanejad S, Rojas-Jimenez K, Grossart HP. 2018. Notes on *Dictyuchus* species (Stramenopila, Oomycetes) from Anzali lagoon, Iran. Mycologia Iranica 5(2): 79–89.
- Mehrabi Z, Firouzbakhsh F, Rahimi-Mianji G, Paknejad H. 2020. Immunity and growth improvement of rainbow trout (Oncorhynchus mykiss) fed dietary nettle (Urtica dioica) against experimental challenge with Saprolegnia parasitica. Fish & Shellfish Immunology 104: 74– 82.
- Mehrabi Z, Firouzbakhsh F, Rahimi-Mianji G, Paknejad H. 2019. Immunostimulatory effect of *Aloe vera (Aloe barbadensis)* on non-specific immune response, immune gene expression, and experimental challenge with *Saprolegnia parasitica* in rainbow trout (*Oncorhynchus mykiss*). Aquaculture (2019): 330–338.
- Mehrabi Z, Firouzbakhsh F, Rahimi G, Kolangi H. 2017. The Effect of Adding Aloe Vera (*Aloe*

barbadensis) in the Diet on Growth Performance and Some Blood Parameters and Serum Biochemical Indices in Rainbow Trout (*Oncorhynchus mykiss*) Challenged with *Saprolegnia parasitica*. Journal of Fisheries 70(1): 60–69.

- Mousavi HA, Soltani M, Khosravi A, Mood SM, Hosseinifard M. 2009. Isolation and characterization of Saprolegniaceae from rainbow trout (*Oncorhynchus mykiss*) eggs in Iran. Journal of Fisheries and Aquatic Science 4(6): 330–333.
- Mousavi HA. 2007. Isolation and identification of parasite and saprophyte fungi from fungal affected eggs of the rainbow trout [*Oncorhynchus mykiss*] in Mazandaran province. Iranian Journal of Veterinary Research 62(3): 163–168.
- Mousavi H, Sharif Rouhani M, Khosravi AR, Mehrabi Yad E, Akhoundzadeh Basti. 2006. The Evaluation of *Eucaliptus Camaldolensis* Dehnh esscence application in control of fungal pollution of trout eggs. Journal of Medical Plants 5(20): 42–47.
- Mostafa AA, Al-Askar AA, Yassin MT. 2020. Antisaprolegnia potency of some plant extracts against *Saprolegnia diclina*, the causative agent of saprolengiasis. Saudi Journal of Biological Sciences 27(6): 1482–1487.
- Nekuie Fard A, Motalebi AA, Jalali Jafari B, Aghazadeh Meshgi M, Azadikhah D, Afsharnasab M. 2011. Survey on fungal, parasites and epibionts infestation on the *Astacus leptodactylus* (Eschscholtz, 1823), in Aras Reservoir West Azarbaijan, Iran. Iranian Journal of Fisheries Sciences 10(2): 266–275.
- Neitzel DA, Elston RA, Abernethy CS. 2004. Prevention of prespawning mortality: cause of salmon headburns and cranial lesions. Pacific Northwest National Lab. (PNNL), Richland, WA (United States).
- Pazira AR. 2017. Antifungal activity of Thymus essential oil (*Thymus vulgaris*) on fungi isolated from the skin of Koi (*Cyprinus carpio* var. *Koi*). Journal of Aquaculture Development 11(2): 13–22.
- Peyghan R, Rahnama R, Dezfuly ZT, Shokoohmand M. 2019. Achlya infection in an Oscar (Astronotus ocellatus) with typical symptoms of saprolegniosis. Veterinary Research Forum 10(1): 89–92.
- Põlme S, Abarenkov K, Henrik Nilsson R, Lindahl BD, Clemmensen KE, Kauserud H, Nguyen N, Kjøller R, Bates ST, Baldrian P, Frøslev TG. 2020. FungalTraits: a user-friendly traits database of fungi and fungus-like stramenopiles. Fungal Diversity 105(1): 1–16.
- Rezakhani F, Khodaparast SA, Masigol H, Roja-Jimenez K, Grossart HP, Bakhshi M. 2019. A preliminary report of aquatic hyphomycetes isolated from Anzali lagoon (Gilan province, North of Iran). Rostaniha 20(2): 123–143.
- Rocha SC, Lopez-Lastra CC, Marano AV, de Souza JI, Rueda-Páramo ME, Pires-Zottarelli CL. 2018.

New phylogenetic insights into Saprolegniales (Oomycota, Straminipila) based upon studies of specimens isolated from Brazil and Argentina. Mycological Progress 17(6): 691–700.

- Rocha SC, Sandoval-Sierra JV, Dieguez-Uribeondo J, Goncalves DR, Jeronimo GH, De Jesus AL, Marano AV, Pires-Zottarelli CL. 2016. Saprolegnia milanezii sp. nov., a new species of Saprolegniales (Oomycota, Straminipila) from Brazil. Phytotaxa 270(4): 286–294.
- Salah M, Zorriezahra S, Sepahdari A, Hosseni A, Falahatnaserabad I, Rastiannasab A. 2021. Comparison of Huwa-San TR-50 solution efficiency with malachite green and Formalin on fungal contamination control of rainbow trout (*Oncorhynchus mykiss*) eggs in incubation stage. Journal of aquaculture development 14(1): 53–61.
- Salimian F, Fattollahi M, Nematollahi A, Nikookhah F, Kharazian N. 2015. Antifungal activity in vitro of aqueous and total flavnoids extracts of plant *Myrtus communis* L. against two pathogenically important fungi, Saprolegnia and Fusarrium isolated from rainbow trout eggs. Journal of marine sciences and technology 16(1): 16–28.
- Scanu B, Jung T, Masigol H, Linaldeddu BT, Jung MH, Brandano A, Mostowfizadeh-Ghalamfarsa R, Janoušek J, Riolo M, Cacciola SO. 2021. *Phytophthora heterospora* sp. nov., a New Pseudoconidia-Producing Sister Species of *P. palmivora*. Journal of Fungi 7(10): 870.
- Sedighi A, Sodagar M, Hashemi-Koroni M, Hosseini SS. 2015. Comparing the effect of antifungal Zinc oxide nanoparticles and Malachite green on *Saprolegnia* sp. The Journal of Aquatic Exploitation and Breeding 4(1): 29–37.
- Shahbazian NA, Ebrahimzadeh Mousavi HA, Soltani MA, Khosravi AR, Mirzargar S, Sharifpour I. 2010. Fungal contamination in rainbow trout eggs in Kermanshah province propagations with emphasis on Saprolegniaceae. Iranian Journal of Fisheries Sciences 9(1): 151–160.
- Shahrani M, Azari Takami G, Sharif Rohani M, Motallebi A, Yazdani Sadati M. 2021. Antifungal effects of alcoholic extract of *Thymus vulgaris* on Siberian sturgeon (*Acipenser baerii*) eggs compared with malachite green effects. Iranian Journal of Fisheries Sciences 20(1): 218–229.
- Sharifi A, Gorjipour R, Gorjipour AA, Mohammadi R, Jabarnejad A. 2012. Antifungal effect of quercus infectoria gall (oak) on *saprolegnia* fungi. Armaghane danesh 17(1): 78–84.
- Sharifpour I, Mazandarani M, Khoshbavar Rostami HA. 2014. Investigation of farmed Cyprinidae contaminants from Golestan province. The Journal of Aquatic Exploitation and Breeding 3(3): 15–25.
- Shokouh Saljoghi Z, Farhadian O, Ramezanian N, Mehraban Sangatash M. 2020. Synthesis and characterization of novel compounds and

determining their antifungal properties against rainbow trout pathogen, *Saprolegnia* sp. in vitro. Iranian Journal of Fisheries Sciences 19(3): 1396– 1414.

- Sigoillot JC, Berrin JG, Bey M, Lesage-Meessen L, Levasseur A, Lomascolo A, Record E, Uzan-Boukhris E. 2012. Fungal strategies for lignin degradation. Advances in botanical research 61: 263–308.
- Singer D, Lara E, Steciow MM, Seppey CV, Paredes N, Pillonel A, Oszako T, Belbahri L. 2016. Highthroughput sequencing reveals diverse oomycete communities in oligotrophic peat bog microhabitat. Fungal Ecology 1(23): 42–47.
- Svoboda J, Mrugała A, Kozubíková-Balcarová E, Petrusek A. 2017. Hosts and transmission of the crayfish plague pathogen *Aphanomyces astaci*: a review. Journal of fish diseases 40(1): 127–140.
- Tandel RS, Dash P, Bhat RA, Thakuria D, Sawant PB, Pandey N, Chandra S, Chadha NK. 2021. Antioomycetes and immunostimulatory activity of natural plant extract compounds against *Saprolegnia* spp.: Molecular docking and in-vitro studies. Fish & Shellfish Immunology 114: 65–81.
- SavadKouhi P, Ahari H, Anvar AA, Jafari B. 2021. Effect of *Carum copticum* nano-essence against *Saprolegnia* and *Fusarium*, and the use of multiplex PCR assay for the detection of these organisms in rainbow trout *Oncorhynchus mykiss*. Archives of Razi Institute 76(2): 231–241.
- Van Den Berg AH, McLaggan D, Diéguez-Uribeondo J, Van West P. 2013. The impact of the water moulds Saprolegnia diclina and Saprolegnia parasitica on natural ecosystems and the aquaculture industry. Fungal Biology Reviews 27(2): 33–42.
- Wolinska J, King KC, Vigneux F, Lively CM. 2008. Virulence, cultivating conditions, and phylogenetic analyses of oomycete parasites in Daphnia. Parasitology 135(14): 1667–1678.
- Wu L, Chang KF, Conner RL, Strelkov S, Fredua-Agyeman R, Hwang SF, Feindel D. 2018. *Aphanomyces euteiches*: a threat to Canadian field pea production. Engineering 4(4): 542–551.

وضعیت کنونی راسته ساپرولگنیالز در ایران: فراخوان قارچشناسان برای دستیابی به وضوح بیشتر در آرایهبندی و بومشناسی

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چکیده : قدمت مطالعه راسته ساپرولگنیالز به ۱۵۰ سال میرسد. طی این دوران، اعضای این راسته غالباً به عنوان عوامل بیماریزای موجود در آب شیرین شناخته شدهاند. در ایران نیز جنبههای بیماریشناختی اعضای این راسته، همسو با مطالعات جهانی، محور اصلی بیشتر پژوهشها بوده است. از این رو، نقد حاضر تازهترین پژوهشهای صورت گرفته در ایران را بررسی کرده، به نارساییهای موجود می پردازد. ارزیابی ما نشان داد که تمرکز بیش از ۸۰ درصد از مقالههای منتشر شده در ایران روی اثر عصارههای گیاهی مورت گرفته در ایران را بررسی کرده، به نارساییهای موجود می پردازد. ارزیابی ما نشان داد که تمرکز بیش از ۸۰ درصد از مقالههای منتشر شده در ایران روی اثر عصارههای گیاهی مختلف و دیگر ترکیبات گیاهی روی گونه یه مروز بیش از ۸۰ درصد از مقالههای منتشر شده در ایران روی اثر عصارههای گیاهی مختلف و دیگر ترکیبات گیاهی روی گونه یه تمرکز بیش از ۸۰ درصد از مقالههای منتشر شده در ایران روی اثر عصارههای گیاهی مختلف و دیگر ترکیبات گیاهی روی گونه یه تمرکز بیش از ۲۰ درصد از مقالههای منتشر شده در ایران روی اثر عصاره های گیاهی مختلف و دیگر ترکیبات گیاهی روی گونه یه موجود می در می رولی مهیان بوده است. از سوی دیگر، مشخص شد که پژوهشهای ایرانی مربوط به آرایهبندی و بومشناسی اعضای راسته ساپرولگنیالز به شکلی محدود انجام گرفته است. این نوشتار در نهایت برای مطالعه سازمانیافته این راسته در ایران پیشنهادهایی مطرح کرده است. در این مقاله از تمامی قارچشناسان، به ویژه در نهایت برای مقاله ایران، دعوت شده است تا مطالعه راسته ساپرولگنیالز را جدی تر و با برنامه دیزی منسجم تر در دستور کار مود.

كلمات كليدى: أأميكوتا، بومشناسى، بيمارىشناسى، تنوع، سيستماتيك، مديريت بيمارى