

## Research Article

# Pathological and genotoxic effects of the herbicide oxadiargyl on common carp (*Cyprinus carpio*) fingerlings

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### Abstract

Acute toxicity and effects of sublethal concentrations of oxadiargyl herbicide (3% EC) were examined on DNA damage (Comet assay) and histopathological changes in common carp (*Cyprinus carpio*) fingerlings with average weight and length of  $19.15 \pm 1.05$  g and  $10.09 \pm 0.47$  cm, respectively. The fish were exposed to 0.1, 0.3 and 0.5 mg/L of the herbicide for 30 days. Estimated 96-h LC<sub>50</sub> value for oxadiargyl in common carp was 0.6 mg/L. Histopathologically, no change occurred in different tissues of the control group, while marked lesions were induced in vital organs of fish that their severity was increased with enhancement of the herbicide concentration. Sublethal exposure to different concentrations of oxadiargyl induced: hyperplasia of lamellar epithelium, hyperemia, inflammatory cells infiltration, aneurysm and rod-like structures of secondary lamellae in gill tissues, as well as change in size and number of melanomacrophage centers in kidney and spleen tissues. Necrosis of tubular epithelium, hyperemia, and protein casts were also observed in kidney tissue. Focal necrosis, fragmentation, vaculization and shrinkage of myofibrils, and eosinophilic cytoplasm were observed in muscle tissues of exposed fish. Erythrocyte cells of fish exposed to sublethal concentrations of 0.1, 0.3 and 0.5 mg/L, showed 18.3%, 19.1%, and 31.5% tailed DNA, respectively, significantly higher than the control group ( $p < 0.05$ ). Moreover, exposure to oxadiargyl significantly decreased WBC, RBC, Hb, Hct compared with the control group ( $p < 0.05$ ). In conclusion, these results revealed that oxadiargyl is highly toxic to common carp with genotoxic and hematotoxic effects, as well as adverse effects on histopathology of vital organs.

**Keywords:** Oxadiargyl, Common carp, Histopathology, Hematology, DNA damage

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## Introduction

Herbicides are the most potentially harmful chemicals used in agriculture that can enter into aquatic environments and affect fauna, especially fish (Wany *et al.*, 1992; Arshad *et al.*, 2006).

Exposure of fish even to low environmentally-relevant concentrations of herbicides may result in abnormal behaviors (Steinberg *et al.*, 1995), retarding growth performance (Sweilum, 2006), various physiological disorders (Blahova *et al.*, 2014; Ahmadvand *et al.*, 2016), and devastating deaths (Bálint *et al.*, 1997), as well as adverse reproductive and immune effects (Ahmadvand *et al.*, 2015; Xing *et al.*, 2015).

Oxadiargyl (C<sub>15</sub>H<sub>14</sub>C<sub>12</sub>N<sub>2</sub>O<sub>3</sub>) belongs to the group of oxadiazole, and is a broad-leaf herbicide extensively applied in rice fields in North Europa and Asia, characterized by its inhibition to protoporphyrinogen IX oxidase enzyme (Hwang *et al.*, 2004). Its half-life in soil is 20-30 days and its residues can affect soil as well as water fauna (Mahmoudi *et al.*, 2011).

Oxadiazole chemical family have been found to be highly toxic to fish with adverse effects on growth and biochemical parameters, as well as chemical structure of DNA (Ajani *et al.*, 2015; Zanjani *et al.*, 2017). However, there are a limited data concerning toxic effects of oxadiargyl on aquatic animals, and most of studies focused on phytotoxic effects of the herbicide (Nethra and Jagannath, 2011; Monjezi *et al.*, 2015).

Toxic effects of herbicides on fish are investigated using many biomarkers, including histopathological studies of vital organs and haematological parameters due to association between external environment and the circulatory system, as well as changes in response to toxic substances (Wendelaar-Bonga, 1997; Ahmadvand *et al.*, 2014; Blahova *et al.*, 2014). Comet assay, another sensitive technique for detection of a wide variety of DNA damage, is widely used to determine genotoxic potential of pesticides on aquatic organisms (Jin *et al.*, 2004; Mitkovska *et al.*, 2017).

Common carp (*Cyprinus carpio*) is one of the main ichthyofauna in rivers of southern Caspian Sea, as well as commercial fish species that mostly farmed in littoral provinces of Iran (Golestan, Mazandaran, and Gilan), the area of high oxadiargyl use in rice fields (Mahmoudi *et al.*, 2011). Hence, this study was aimed to investigate acute toxicity and effects of sublethal concentration of oxadiargyl herbicide on common carp (*C. carpio*) fingerlings.

## Materials and methods

### Herbicide

Technical grade oxadiargyl herbicide (3% EC) manufactured by Saveh Herbicide Company (Arak, Iran) was used to evaluate its toxicity to carp. Stock solutions were prepared in acetone and tap water.

### Fish

Fingerlings of Caspian common carp with weight of 19.15±1.05 g and fork length of 10.09±0.47 cm were obtained

from Shahid Beheshti fish breeding center (Rasht, Iran) and were acclimated to laboratory condition in 1000 liter tanks with dechlorinated tap water for 10 days. The fish were fed using commercial FFC-extruded fish feed (Faradaneh Company; ShahreKord, Iran) twice a day, and maintained under a natural photoperiod (approximately 12h light/ 12h dark). During the experiment, physicochemical characteristics of water, including temperature, oxygen, pH, and total hardness were measured daily based on OECD guidelines (2016).

#### *Determination of 96h-LC<sub>50</sub> value*

For acute toxicity, different concentrations of herbicides (0, 0.1, 0.3, 0.5, 0.7, 1, 2, 4, 6 and 8 mg/L) were prepared and fish in duplicated groups (n=20) were exposed to them in 100 L tanks. The 96-h LC<sub>50</sub> was determined by Probit analysis. A control group with oxadiargyl free water was also maintained. Dead fish were counted and removed.

#### *Sublethal exposure to oxadiargyl*

For sublethal toxicity tests, 120 fish were selected and introduced into four duplicate 100 liter tanks (n=15) and were exposed to concentrations of 0.1, 0.3 and 0.5 mg/L of oxadiargyl to investigate DNA damage of erythrocyte cells and histopathological changes of vital organs in 30 days. Two control tanks (n=15) with oxadiargyl free water were also maintained. Water was completely renewed daily and its characteristics were monitored before and after water

exchange, as well as herbicide concentrations were adjusted.

#### *Hematology*

On day 30 of sublethal exposure, five fish from each replicate were anesthetized with clove essence (200 mg/L), and blood was collected from caudal vein puncture using a heparinized syringe and transferred to a 2 mg/L heparinized tube containing 0.01 mg/L of sodium heparin solution (5000 I.U). Hematological parameters, including white blood cells (WBCs), red blood cells (RBCs), hemoglobin (Hb), and hematocrit (Hct) were measured (Ahmadivand *et al.*, 2014).

#### *Histopathology*

For histological examinations at day 30 after sublethal exposure, three fish from each replicate were taken and a piece of kidney, gill, spleen, and muscle tissues were removed. Tissues were then fixed in 10% neutral buffered formalin (NBF), and after serial dehydration in ethanol, embedded in paraffin and sectioned at 5  $\mu$ m thickness. Tissue sections were stained with hematoxylin and eosin (Hewitson *et al.*, 2010), and examined under a light microscope (E600; Nikon).

#### *Comet assay*

After exposure periods, DNA damage of erythrocyte cells was determined by alkaline single cell gel electrophoresis (Singh *et al.*, 1988). A mixture of a blood sample (5 $\mu$ L) with 0.5% (w/v) low-melting agarose (95  $\mu$ L) was spread over degreased microscope slides, previously covered with 1% normal melting agarose,

and then allowed to solidify at 4°C for 20 min. The embedded cells were lysed in fresh cold lysing buffer (2.5M NaCl, 100mM Na<sub>2</sub>EDTA, 1% Triton X-100, 10% DMSO, 10mM Tris-HCl, and pH: 10) at 4°C overnight.

After 30 min incubation in electrophoresis buffer (300 mM NaOH, 1 mM EDTA, pH ≥ 13) electrophoresis was carried out at 20 V and 300 mA for 30 min. Unwound DNA in the slides was neutralized by three washing steps in 0.4 M Tris-HCl (pH 7.5). The slides were stained with ethidium bromide (15 µg/mg/L) to visualize DNA strand breaks, and examined using a fluorescence microscope (E600; Nikon). Two slides per specimen (25 cells per slide) were analyzed, and DNA damage was quantified as percent of tailed ones.

#### Statistical analysis

The obtained data were analyzed using

the statistical package SPSS23 software (Chicago, IL, USA) by one-way analysis of variance (ANOVA) followed by Tukey's pairwise multiple comparison test. The data were provided as mean±standard deviation and differences were considered statistically significant when  $p < 0.05$ .

#### Results

##### Determination of 96-hLC<sub>50</sub> value

Mortality rate of acute toxicity test of oxadiargyl on common carp is shown in Figure 1. Measured 96-h LC<sub>50</sub> value by probit analysis was 0.6 mg/L. During the experiment, none of unexposed control fish died and showed abnormal behavior. Abnormal behaviors observed in those exposed to oxadiargyl were erratic swimming, accelerated respiration, hanging vertically, and staying motionless on the bottom of tank.

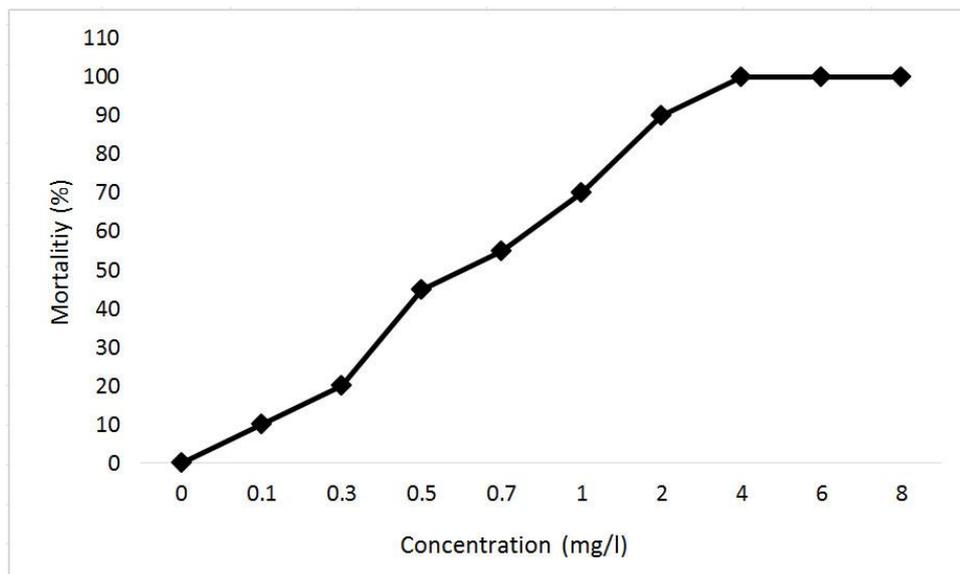


Figure 1: Mortality of common carp (*C. carpio*) at 96h after acute exposure to oxadiargyl.

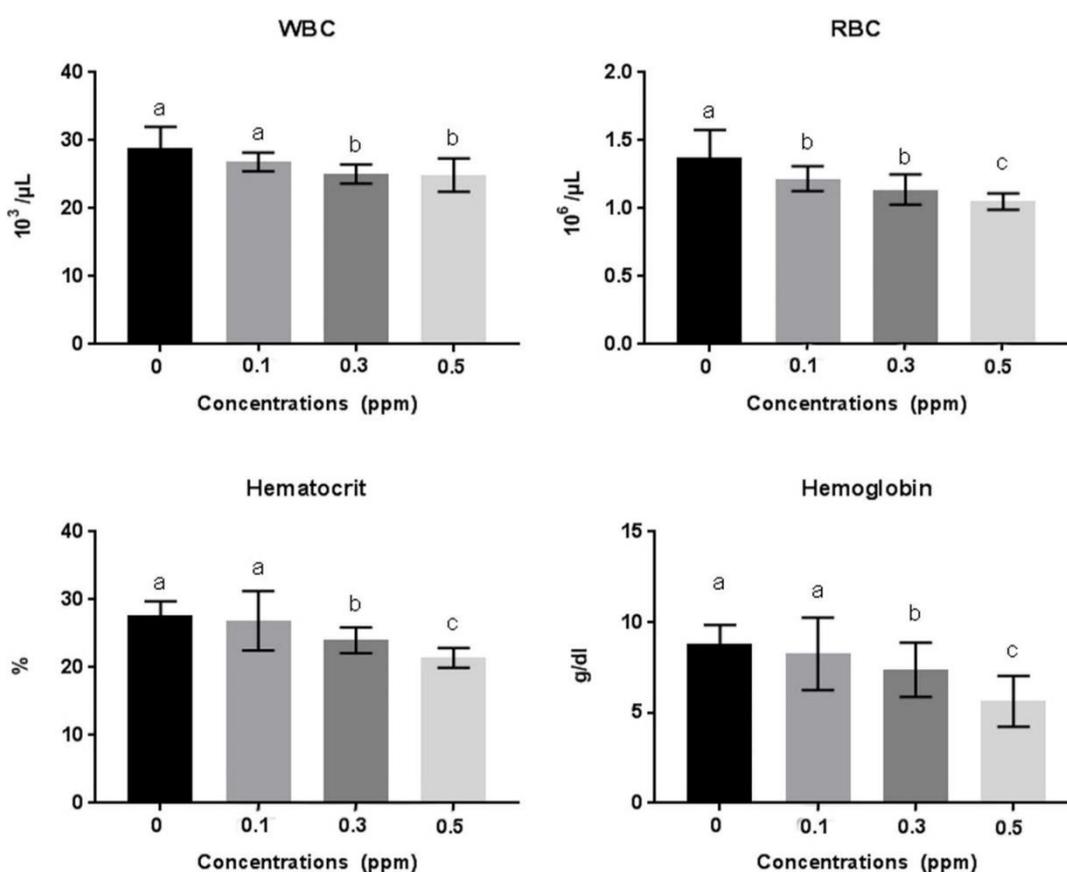
### Sublethal exposure

No mortality was observed in control and treatment groups during sublethal exposure (0, 0.1, 0.3 and 0.5 mg/L) period. However, some abnormal behavioral and swimming patterns were observed in 0.5 mg/L oxadiargyl exposed fish groups.

### Hematology

The results of measured hematological

parameters are shown in Figure 2. There was a significant increase in WBC, decrease in RBC, Hb and Hct levels of fish exposed to 0.3 and 0.5 mg/L, as well as RBC level in 0.1 mg/L trial group when compared with the control group ( $p < 0.05$ ). However, exposure to 0.1 mg/L did not show significant change in WBC, Hct and Hb levels ( $p > 0.05$ ).



**Figure 2:** Hematological parameters of common carp (*C. carpio*) after 30 days exposure to different concentrations (0, 0.1, 0.3 and 0.5 mg/L) of oxadiargyl. Different letters indicate significant differences among groups at  $p < 0.05$ . Error bars show standard deviation.

### Histopathology

No histopathological change was observed in gills, kidney, spleen, and muscle of the control group. However, marked histological alterations were

observed in examined organs of treatment groups depending on herbicide concentration (Table 1).

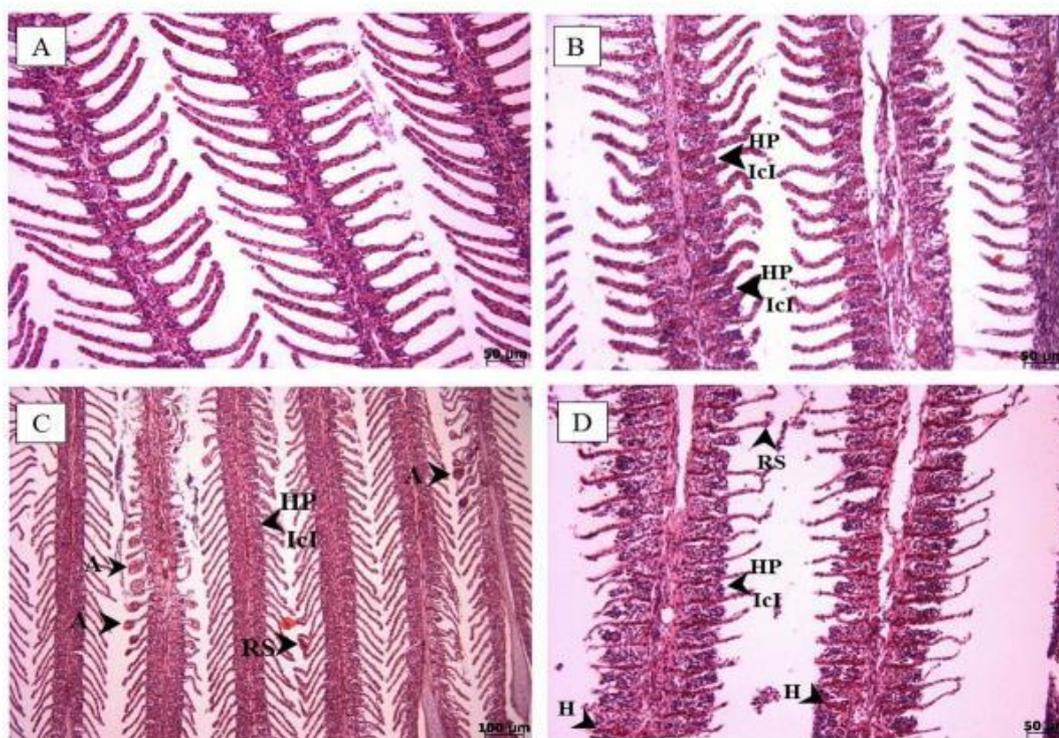
**Table 1: Histological lesions in gills, kidney, spleen, and muscle tissues of common carp exposed to oxadiargyl.**

<i>Tissue</i>	<i>Lesions</i>
Gills	Hyperemia, Hyperplasia, Inflammatory cells infiltration, Aneurysm, Necrosis, Rod-like structures
Kidney	Proteinuria, Melanomacrophage centers, Necrosis, Hyperemia
Muscle	Fragmentation, Vacuolization and shrinkage, Necrosis, Inflammatory cells infiltration
Spleen	Melanomacrophage centers

### *Gills*

Sublethal exposure to different concentrations of oxadiargyl, induced hyperplasia of lamellar epithelium, hyperemia, inflammatory cells infiltration, aneurysm and rod-like

structures of secondary gill lamellae in gills tissues. Severity of lesions increased with enhancing oxadiargyl concentrations (Fig. 3B-D).

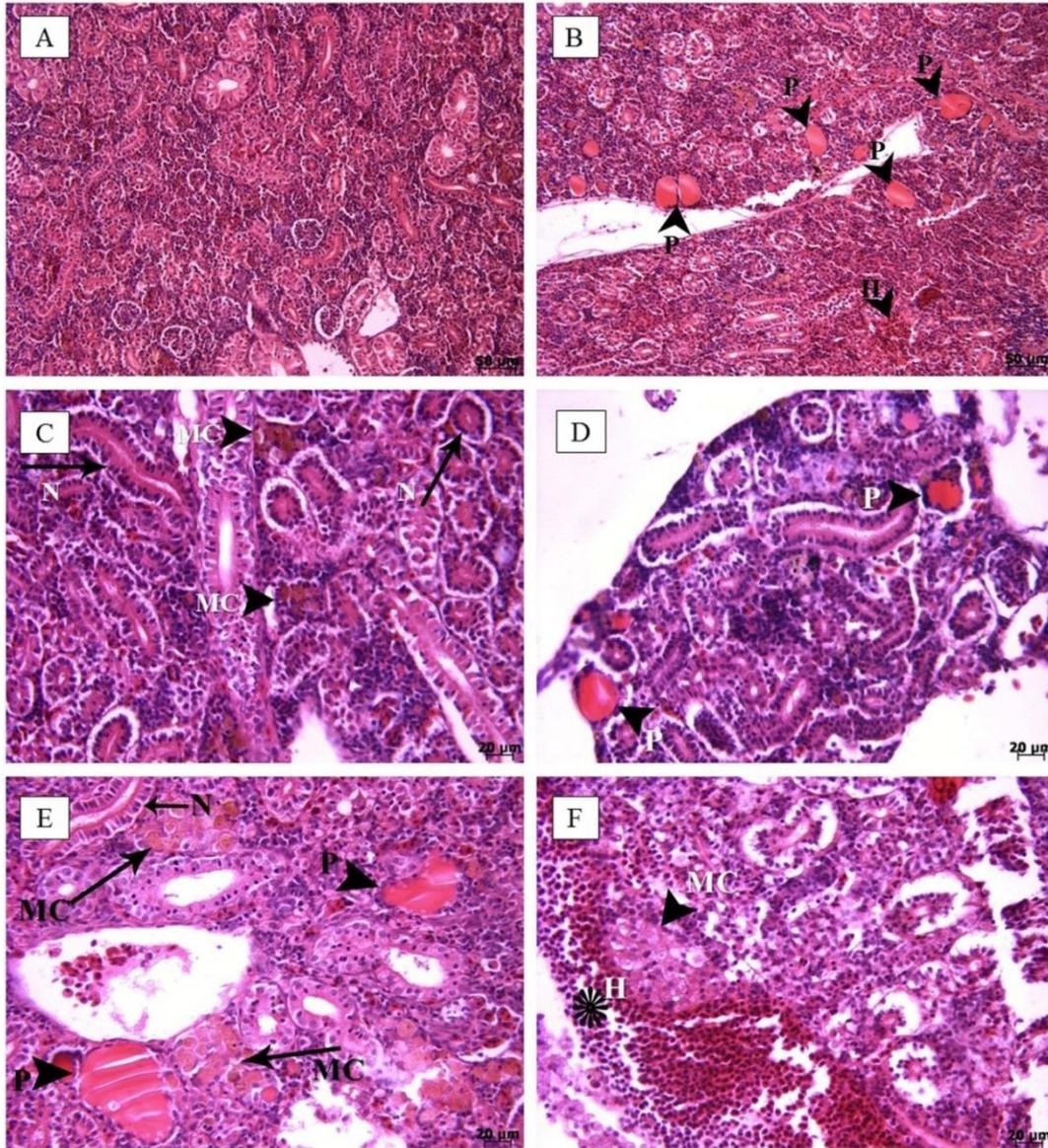


**Figure 3: Histopathological changes of gill tissue of common carp (*C. carpio*) after 30 days sublethal exposure to oxadiargyl. (A) gill tissue of control treatment, (B) fish exposed to 0.1 mg/L, (C) 0.3 mg/L, and (D) 0.5 mg/L of oxadiargyl; A: aneurysm, H: hyperemia, HP: hyperplasia, I: inflammatory cells infiltration, RS: rod-like structures.**

### *Kidney*

Kidney tissue of common carp exposed to oxadiargyl, showed histopathological lesions, including necrosis of tubular epithelium, hyperemia, and protein casts in tubules (proteinuria). Moreover,

change in size and number of melanomacrophage centers were seen. Severity of tissue alterations increased as herbicide concentration increased (Fig. 4B-F).



**Figure 4: Histopathological changes of kidney tissue of the common carp (*C. carpio*) after 30 days sublethal exposure to oxadiargyl. (A) kidney tissue of control treatment, (B) fish exposed to 0.1 mg/L, (C) 0.3 mg/L, and (D-F) 0.5 mg/L of oxadiargyl; H: hyperemia, MC: melanomacrophage centers, N: necrosis, P: proteinuria, Asterisk: cellular infiltration.**

### *Spleen*

Increase in size and number of melanomacrophage centers in spleen tissue of exposed common carp to different concentrations of oxadiargyl was frequently seen (Fig. 5B-D).

### *Muscles*

Structural details of muscle tissue of control treatment are shown in Figure 6(A). In muscle tissue of exposed fish, histopathological lesions, including focal necrosis, fragmentation, vacuolization and shrinkage of myofibrils, and eosinophilic cytoplasm were evident (Fig. 6B-D).

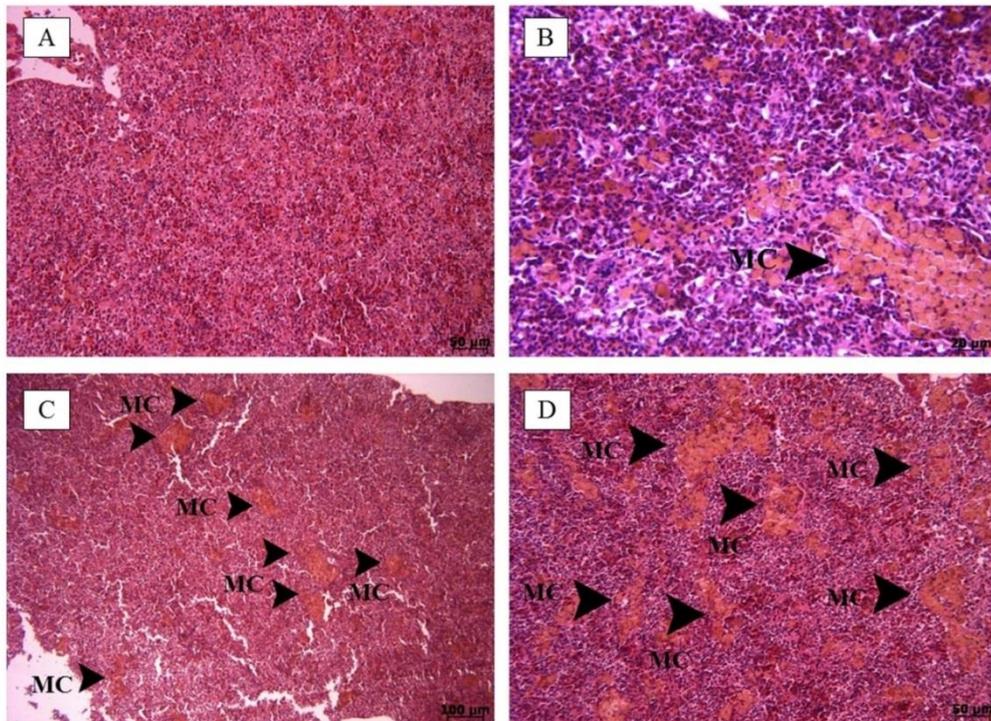


Figure 5: Histopathological changes of spleen tissue of common carp (*C. carpio*) after 30 days sublethal exposure to oxadiargyl. (A) control fish spleen, (B) spleen tissue of fish exposed to 0.1 mg/L, (C) 0.3 mg/L, and (D) 0.5 mg/L of oxadiargyl; MC: melano-macrophage centers.

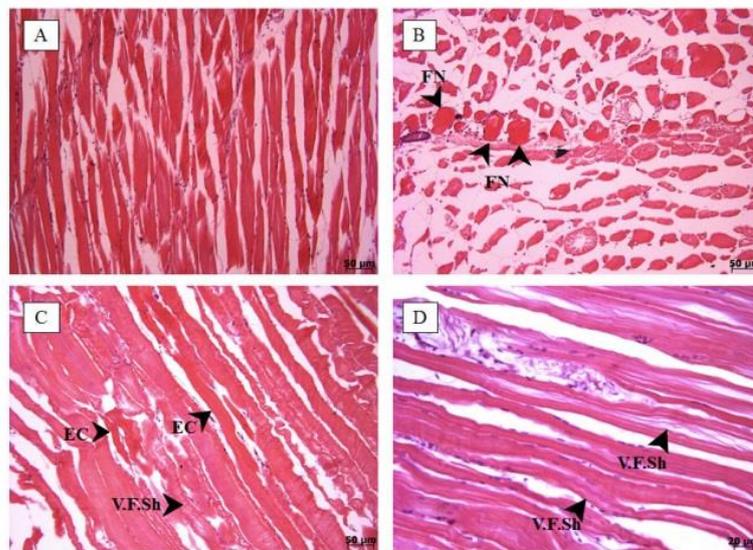
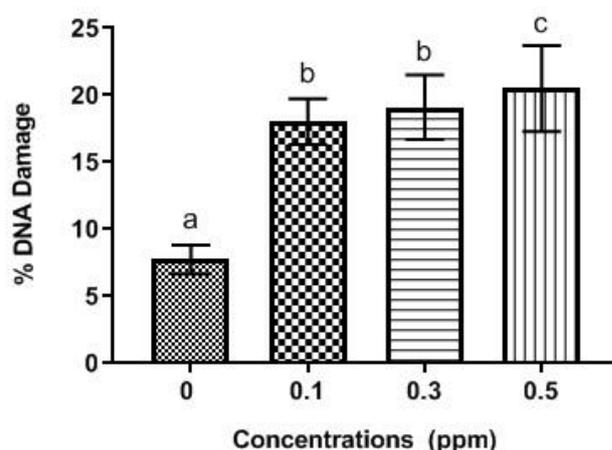


Figure 6: Histopathological changes of muscle tissue of common carp (*C. carpio*) after 30 days sublethal exposure to oxadiargyl. (A) control treatment muscle, (B) muscle tissue of fish exposed to 0.1 mg/L, (C) 0.3 mg/L, and (D) 0.5 mg/L of oxadiargyl; EC: eosinophilic cytoplasm, FN: focal necrosis, V.F.Sh: vacuolization, fragmentation, and shrinkage.

### DNA damage

Results of DNA damage (% tailed DNA) induced by oxadiargyl in blood samples of *C. carpio* are shown in Figure 7. Cell viability was above 90% in the treatments, allowing comet assay to be performed. Oxadiargyl had a genotoxic effect, and treatments with 0.1, 0.3 and 0.5 mg/L resulted in 2-3 folds increases in percent of tailed DNA compared to the control (7%). Among

the treatments, the highest damage (31.5%) was recorded in blood samples of 0.5 mg/L trial group followed by 0.3 mg/L (19.1%) and 0.1 mg/L (18.3%). However, there was no statistically significant difference in percentage of tail DNA in cells of fish exposed to 0.3 mg/L when compared with the 0.1 mg/L treatment.



**Figure 7:** DNA damage (% tail DNA) in erythrocyte cells of common carp (*C. carpio*) on day 30 of exposure to different concentrations (0, 0.1, 0.3 and 0.5 mg/L) of oxadiargyl. Different letters indicate significant difference among groups ( $p < 0.05$ ). Error bars show standard deviation.

### Discussion

Despite adverse effects of herbicide exposure to aquatic fauna, there is little information about toxicity effects of oxadiargyl, which is main herbicide used in rice farming in Gilan, Mazandaran and Golestan provinces. Residues of this herbicide and other kinds enter into rivers and may be transferred into Caspian Sea, where it may contaminate aquatic fauna and then human consumers. In this research, toxicity effects of oxadiargyl on Caspian common carp fingerlings was

studied. Obtained results for 96-h  $LC_{50}$  value for common carp was 0.6 mg/L which is lower than that reported by Sadeghi and Imanpoor (2015) for another member of oxadiazole group (oxadiazon) for platyfish (*Xiphophorus maculatus*; 7.59 mg/L) which may be due to difference in fish species, herbicide and also water quality (Gupta *et al.*, 1981; Farah *et al.*, 2004).

In response to a stressor, such as pesticide exposure, fish undergo series of hematological changes in attempt to compensate the challenge imposed on

them and thus cope with the stress (Wendelaar-Bonga, 1997). According to the results of this study, hematological parameters showed significant ( $p < 0.05$ ) reduction in levels of WBC, RBC, Hb, and Hct in the fish. These could be due to oxadiargyl herbicide effects on hematopoietic tissue, which together with observed changes in levels of erythrocyte and leukocytes can lead to impairment in process of hematopoiesis and reduce the fish innate immune system.

Saravanan *et al.* (2017), assessing the acute toxicity effects of 0.5, 5 and 50  $\mu\text{g/L}$  concentrations of oxadiazon on carp for 96 h, found that this herbicide causes a significant decrease in RBC, Hb, and Hct. Decrease in RBC, Hct, and Hb content in this study could be explained as compensatory response that reduces oxygen-carrying capacity to maintain gas transfers and indicates a change in water blood barrier for gas exchange in gill lamellae (Jee *et al.*, 2005). Obtained results indicated that exposure to sublethal concentrations of oxadiargyl, severely affected hematological parameters in common carp which was in agreement with those obtained after exposure to other herbicides (Ahmadivand *et al.*, 2014; Blahova *et al.*, 2014), indicating disruption of hematopoiesis, as well as induction of cellular apoptosis.

The same results of leucocytes, erythrocyte, hemoglobin, and hematocrit decrease were reported in *C. carpio* after sublethal exposure to oxadiazon (Zanjani *et al.*, 2017).

Decrease in RBC, Hb, and Hct also indicates hypoxic condition and/or respiration dysfunction affecting circulating system, may be due to impaired gas exchange in gill lamellae (Jee *et al.*, 2005). Moreover, change in WBC levels suggests immunotoxic potential of the herbicide (Ahmadivand *et al.*, 2015).

Histopathological studies of vital organs and serum biochemical parameters due to the association between external environment and the circulatory system, as well as their changes in response to toxic substances were widely used to determine the effects of pollutants on fishes (Wendelaar-Bonga, 1997; Ahmadivand *et al.*, 2014; Blahova *et al.*, 2014).

In this study, oxadiargyl induced mildly to severe histological lesions in gills, kidney, muscle, and spleen of common carp depending on herbicide concentration, which is in agreement with previous studies reporting different histopathological changes in common carp following exposure to herbicides (Poleksić and Karan, 1999; Blahova *et al.*, 2014; Stoyanova *et al.*, 2015). However, these lesions seem to be a result of increased cell activities and reversible in proper conditions.

The observed changes in size and number of melanomacrophage centers (MMCs) in kidney and spleen tissues, depending on the herbicide concentration, confirmed that MMCs can be considered as a biomarker of environmental stress such as pesticides (Ribeiro *et al.*, 2011).

In kidney tissue necrosis of tubular epithelium, hyperemia, and protein casts in the tubules (proteinuria) were also observed. Therefore, as kidney showed endocrine, immune and hematopoietic functions, oxadiargyl can produce toxic effects on many important physiological processes in fish. Exposure to oxadiargyl also induced histological changes in muscles tissues of the fish which may result in obstruction of circulation and digestive systems, and lower fillet quality (Wendelaar-Bonga, 1997).

Sublethal exposure to different concentrations of oxadiargyl, induced hyperplasia of lamellar epithelium, hyperemia, inflammatory cells infiltration and aneurysm of secondary gill lamellae in gill tissues. Severity of lesions increased with enhancing oxadiargyl concentration. The induced histological lesions in gill tissues can also lead to respiratory distress via reduction in oxygen up-taking by secondary lamellae, subsequently decreasing fish activity and growth performance (Caldwell, 1997). Also, toxic compounds naturally reduce respiratory function of freshwater fish, whereas oxygen consumption rate is applied to maintain cell viability during stress increase.

Significant increase of DNA damage revealed by comet assay in this study indicated genotoxic potential of oxadiargyl in common carp (*C. carpio*). Similarly, it is reported that oxadiazon, belonging to the same chemical class, could cause DNA damages in

erythrocytes of *C. carpio* (Zanjani *et al.*, 2017).

Our finding had also similarity with results of Klobučar *et al.* (2010) and Mitkovska *et al.* (2017), assessing genotoxic effects of pesticides by comet assay in common carp, confirming that species and method can be considered for assessment pesticides in aquatic environments. Also, results of genotoxic studies on pesticides are intermittently dependent on purity of active ingredient.

In this study, DNA damage in erythrocytes cells was recorded after 30 days of exposure, while Cavalcante *et al.* (2008) reported a non-persistent DNA damage in gill cells of streaked prochilod (*Prochilodus lineatus*) exposed to roundup herbicide which may be due to intrinsic differences in cell turnover and/or the repair enzyme system of erythrocytes (Moreno *et al.*, 2014).

In conclusion, this study investigated toxic effects of a widely used herbicide, oxadiargyl, in common carp fingerlings. The results showed that oxadiargyl is highly toxic to common carp and had genotoxic and hematotoxic effects, as well as adverse effects on tissue of vital organs. Moreover, our findings further confirmed that common carp can be considered as a suitable species for assessment of pesticides in aquatic environments. Further studies are needed to find out toxic effects of oxadiargyl on other species that may also naturally be exposed due to their close habitat to the area of the herbicide use.

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## References

- Ahmadivand, S., Farahmand, H., Mirvaghefi, A.R., Eagderi, S., Shokrpoor, S. and Rahmati-Holasoo, H., 2014.** Histopathological and haematological response of male rainbow trout (*Oncorhynchus mykiss*) subjected to butachlor. *Veterinarni Medicina*, 59, 433–439. <https://doi.org/10.17221/7683-vetmed>.
- Ahmadivand, S., Farahmand, H., Mirvaghefi, A., Eagderi, S. and Zargar, A., 2015.** Effects of (anti) androgenic endocrine disruptors (DEHP and butachlor) on immunoglobulin M (IgM) and leukocytes counts of male rainbow trout (*Oncorhynchus mykiss*). *Bulletin of Environmental Contamination and Toxicology*, 94(6), 695-700. <https://doi.org/10.1007/s00128-015-1503-y>.
- Ahmadivand, S., Farahmand, H., Teimoori-Toolabi, L., Mirvaghefi, A., Eagderi, S., Geerinckx, T., Shokrpoor, S. and Rahmati-Holasoo, H., 2016.** Boule gene expression underpins the meiotic arrest in spermatogenesis in male rainbow trout (*Oncorhynchus mykiss*) exposed to DEHP and butachlor. *General and Comparative Endocrinology*, 225, 235-241. <https://doi.org/10.1016/j.ygcen.2015.05.011>.
- Ajani, F., Oluyinka-Ajiboye, A. and Oluwatosin-Oyelowo, O. 2015.** Effects of oxadiazon on nutrient utilization and growth of African catfish (*Clarias gariepinus*). *American Journal of Agricultural Science*, 2(3), 121-125.
- Arshad, U., Aliakbar, A., Sadeghi, M., Jamalzad, F. and Chubian, F., 2006.** Pesticide (diazinon and butachlor) monitoring in waters of the Shahid Beheshti Sturgeon Hatchery, Rasht, Iran. *Journal of Applied Ichthyology*, 22(s1), 231-233. <https://doi.org/10.1111/j.1439-0426.2007.00957.x>.
- Bálint, T., Ferenczy, J., Kátai, F., Kiss, I., Kráczler, L., Kufcsák, O., Láng, G., Polyhos, C., Szabó, I., Szegletes, T. and Nemcsók, J., 1997.** Similarities and differences between the massive eel (*Anguilla anguilla*L.) devastations that occurred in Lake Balaton in 1991 and 1995. *Ecotoxicology and Environmental Safety*, 37(1), 17-23. <https://doi.org/10.1006/eesa.1996.1509>.
- Blahova, J., Modra, H., Sevcikova, M., Marsalek, P., Zelnickova, L., Skoric, M. and Svobodova, Z., 2014.** Evaluation of biochemical, haematological, and histopathological responses and recovery ability of common carp (*Cyprinus carpio* L.) after acute exposure to atrazine herbicide. *BioMed Research International*, 2014, 980948. <https://doi.org/10.1155/2014/980948>.

- Caldwell, C.A., 1997.** Aromatic hydrocarbon pathology in fish following a large spill into the Nemadji River, Wisconsin, USA. *Bulletin of Environmental Contamination and Toxicology*, 58(4), 574-581. <https://doi.org/10.1007/s001289900373>.
- Cavalcante, D.G.S.M., Martinez, C.B.R. and Sofia, S.H., 2008.** Genotoxic effects of Roundup® on the fish *Prochilodus lineatus*. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 655(1-2), 41-46. <https://doi.org/10.1016/j.mrgentox.2008.06.010>.
- Farah, M.A., Ateeq, B., Ali, M.N., Sabir R. and Ahmad, W., 2004.** Studies on lethal concentrations and toxicity stress of some xenobiotics on aquatic organisms. *Chemosphere*, 55(2), 257-265. <https://doi.org/10.1016/j.chemosphere.2003.10.063>.
- Gupta, P.K., Khangarot, B.S. and Durve, V.S., 1981.** The temperature dependence of the acute toxicity of copper to a freshwater pond snail, *Viviparus bengalensis* L. *Hydrobiologia*, 83(3), 461-464. <https://doi.org/10.1007/bf02187041>.
- Hewitson, T.D., Wigg, B. and Becker, G.J. 2010.** Tissue preparation for histochemistry: fixation, embedding and antigen retrieval for light microscopy. *Methods in Molecular Biology*, 611, 3-18. [https://doi.org/10.1007/978-1-60327-345-9\\_1](https://doi.org/10.1007/978-1-60327-345-9_1).
- Hwang, I.T., Hong, K.S., Choi, J.S., Kim, H.R., Jeon, D.J. and Cho, K.Y., 2004.** Protoporphyrinogen IX-oxidizing activities involved in the mode of action of a new compound N-[4-chloro-2-fluoro-5-{3-(2-fluorophenyl)-5-methyl-4,5-dihydroisoxazol-5-yl-methoxy}-phenyl]-3,4,5,6-tetrahydrophthalimide. *Pesticide Biochemistry and Physiology*, 80(2), 123-130. <https://doi.org/10.1016/j.pestbp.2004.06.006>.
- Jee, J.H., Masroor, F. and Kang, J.C., 2005.** Responses of cypermethrin-induced stress in haematological parameters of Korean rockfish, *Sebastes schlegeli* (Hilgendorf). *Aquaculture Research*, 36(9), 898-905. <https://doi.org/10.1111/j.1365-2109.2005.01299.x>.
- Jin, H.H., Lee, J.H. and Hyun, C.K., 2004.** Detection of DNA damage in carp using single-cell gel electrophoresis assay for genotoxicity monitoring. *Journal of Microbiology and Biotechnology*, 14(2), 268-275.
- Klobučar, G.I.V., Štambuk, A., Pavlica, M., Perić, M.S., Hackenberger, B.K. and Hylland, K., 2010.** Genotoxicity monitoring of freshwater environments using caged carp (*Cyprinus carpio*). *Ecotoxicology*, 19(1), 77-84. <https://doi.org/10.1007/s10646-009-0390-6>.
- Mahmoudi, M., Rahnemaie, R., Soufizadeh, S., Malakouti, M.J. and Eshaghi, A., 2011.** Residual effect of thiobencarb and oxadiargyl on spinach and lettuce in rotation with rice. *Journal of Agricultural Science and Technology*, 13, 785-794.

- Mitkovska V.I., Dimitrov, H.A. and Chassovnikarova, T.G., 2017.** In vivo genotoxicity and cytotoxicity assessment of allowable concentrations of nickel and lead: comet assay and nuclear abnormalities in acridine orange stained erythrocytes of common carp (*Cyprinus carpio L.*). *Acta Zoologica Bulgarica Supplement*, 8, 47-56.
- Monjezi, N., Razmjo J. and Karimmojeni, H., 2015.** Valerian (*Valeriana officinalis L.*) tolerance to some post-emergence herbicides. *Journal of Plant Protection Research*, 55(4), 415-420. <https://doi.org/10.1515/jppr-2015-0057>.
- Moreno, N.C., Sofia, S.H. and Martinez, C.B.R., 2014.** Genotoxic effects of the herbicide Roundup Transorb® and its active ingredient glyphosate on the fish *Prochilodus lineatus*. *Environmental Toxicology and Pharmacology*, 37(1), 448-454. <https://doi.org/10.1016/j.etap.2013.12.012>.
- Nethra, N.S. and Jagannath, S., 2011.** Phytotoxic effect of oxadiargyl on germination and early growth of sunflower (*Helianthus annuus L.*) and maize (*Zea mays L.*). *Archives of Phytopathology and Plant Protection*, 44(19), 1901-1907. <https://doi.org/10.1080/03235408.2010.507946>.
- OECD, 2016.** Guideline for the testing of chemicals, section 1: Physical-chemical properties, 18p. Organization for economic cooperation and development (OECD), Paris, France. <https://doi.org/10.1787/20745753>.
- Poleksić, V. and Karan, V., 1999.** Effects of trifluralin on carp: biochemical and histological evaluation. *Ecotoxicology and Environmental Safety*, 43(2), 213-221. <https://doi.org/10.1006/eesa.1999.1790>.
- Ribeiro, H.J., Procópio, M.S., Gomes, J.M.M. Vieira, F.O., Russo, R.C., Balzuweit, K., Chiarini-Garcia H., Castro, A.C.S., Rizzo, E. and Corrêa, J.D., 2011.** Functional dissimilarity of melanomacrophage centres in the liver and spleen from females of the teleost fish *Prochilodus argenteus*. *Cell and Tissue Research*, 346(3), 417-425. <https://doi.org/10.1007/s00441-011-1286-3>.
- Sadeghi, A. and Imanpoor, M.R., 2015.** Investigation of LC50, NOEC, and LOEC of oxadiazon, deltamethrin, and malathion on platy fish (*Xiphophorus maculatus*). *Iranian Journal of Toxicology*, 9(28), 1271-1276. <https://doi.org/10.1006/eesa.1999.1790>.
- Saravanan, M., Kim, J.Y., Hur, K.J., Ramesh, M. and Hur, J.H., 2017.** Responses of the freshwater fish *Cyprinus carpio* exposed to different concentrations of butachlor and oxidiazon. *Biocatalysis and Agriculture Biotechnology*, 11, 275-281. <https://doi.org/10.1016/j.bcab.06.011>.
- Singh, N.P., McCoy, M.T., Tice, R.R. and Schneider, E.L., 1988.** A simple technique for quantitation of low levels of DNA damage in individual cells. *Experimental Cell*

- Research*, 175(1), 184-191.  
[https://doi.org/10.1016/0014-4827\(88\)90265-0](https://doi.org/10.1016/0014-4827(88)90265-0).
- Steinberg, C.E.W., Lorenz, R. and Spieser, O.H., 1995.** Effects of atrazine on swimming behavior of zebrafish, *Brachydanio rerio*. *Water Research*, 29(3), 981-985.  
[https://doi.org/10.1016/0034-1354\(94\)00217-u](https://doi.org/10.1016/0034-1354(94)00217-u).
- Stoyanova, S., Yancheva, V., Iliev, I., Vasileva, T., Bivolarski, V., Velcheva I. and Georgieva, E., 2015.** Glyphosate induces morphological and enzymatic changes in common carp (*Cyprinus carpio* L.) liver. *Bulgarian Journal of Agricultural Science*, 21(2), 409–412.
- Sweilum, M.A., 2006.** Effect of sublethal toxicity of some pesticides on growth parameters, haematological properties and total production of Nile tilapia (*Oreochromis niloticus* L.) and water quality of ponds. *Aquaqulture Research*, 37(11), 1079–1089.  
<https://doi.org/10.1111/j.1365-2109.2006.01531.x>.
- Wany, Y.S., Jaw C.G., Tang H.C., Lin, T.S. and Chen Y.L, 1992.** Accumulation and release of herbicides butachlor, thiobencarb, and chlomethoxyfen by fish, clam, and shrimp. *Bulletin of Environmental Contamination and Toxicology*, 48, 474–480.  
<https://doi.org/10.1007/bf00195650>.
- Wendelaar-Bonga, SE., 1997.** The stress response in fish. *Physiological Reviews*, 77(3), 591–625.  
<https://doi.org/10.1152/physrev.1997.77.3.591>.
- Xing, H., Liu, T., Zhang, Z., Wang, X. and Xu, S., 2015.** Acute and subchronic toxic effects of atrazine and chlorpyrifos on common carp (*Cyprinus carpio* L.): Immunotoxicity assessments. *Fish and Shellfish Immunology*, 45(2), 327-333.  
<https://doi.org/10.1016/j.fsi.2015.04.016>.
- Zanjani, S.A., Emadi, H., Jamili, S., and Mashinchian, A., 2017.** DNA damage and hematological changes in Common carp (*Cyprinus carpio*) exposed to oxadiazon. *International Journal of Aquatic Biology*, 5(6), 387-392.  
<https://doi.org/10.22034/ijab.v5i6.417>.