Research Article

Biochemical composition and investigation on the economic feasibility of sodium alginate production of brown seaweed Sargassum illicifolium (Turner) C. Agardh, 1820 from Chabahar Bay (Gulf of Oman, Iran)

Hafezieh M.1*; Abkenar A.M.2; Jadgal S.3; Ajdari A.3

Received: August 2020 Accepted: October 2020

Abstract

More than 3000 MT of Sargassum illicifolium, annually washed-up from the Oman Sea (Sistan and Baluchestan province)., According to the estimates of the Iranian Fisheries Sciences Researchinstitiute. The brown seaweed biomass has been considered as one of the best free sources for production of sodium alginate. A key objective of this study was to determine the biochemical composition of Sargassum illicifolium collected from Chabahar Bay in November 2018 and to understand the economic potential and cost drivers of sodium alginate on basis of the present macroalgae. Alginates were purified by re-precipitation with ethanol and characterized by infrared spectroscopy. The results showed that the Chabahar Sargassum was characterized by total protein (TP), total lipid (TL) and carbohydrate as 9.8±0.8%, 4.4±0.2% and 33.2±4.1% dry weight, respectively. The ash content contained 41.6±2.3% DW. Moreover, the n-6/n-3 ratio was 2.62 and total essential amino acids and total minerals were 29.1±0.2 mg g⁻¹ DW and 102. 2±0.6 mg g⁻¹ DW, respectively. sodium alginate of Sargassum illicifolium was found to be high as 28.2% purification with molecular weight of 8.06×105 g mol⁻¹. Its total production price was evaluated 7.66 \$ per kg sodium alginate, which is much cheaper than existing ones on the Iranian market.

Keywords: Alginic acid yield, Macro algae, Purification, Proximate composition, Chabahar coast, Sea of Oman

¹⁻Iranian Fisheries Science Research Institute (IFSRI), Agricultural Research Education and Extension Organization (AREEO), P.O. Box: 15745/133. Tehran, Iran.

² Department of Fisheries, Chabahar Branch, Islamic Azad University, Chabahar, Iran

³⁻ Offshore Fisheries Research Center, Iranian Fisheries Science Research Institute (IFSRI), Agricultural Research Education and Extension Organization (AREEO), Chabahar, Iran

^{*}Corresponding author's Email: jhafezieh@yahoo.com

Introduction

potential renewable Seaweeds are resource in the marine environment where about 6,000 species have been identified and grouped in various shades of green (Chlorophyceae), (Phaeophyceae) brown and red (Rhodophyceae); among them, 221 species have commercial value and ten species are intensively cultivated (FAO, 2020FAO, 2020). These seaweeds, mainly the brown one, contribute greatly to the nutritional status of due communities to their composition of micro (I, Fe, Zn, Co, Se, Mo, Fl, Mg, Bo, Ni and Co) and macronutrients such as minerals (Na. Ca, Mn, K, Cl, S and in P), vitamins (B12, A and K), essential amino acids, fatty acids and pigments (FAO, 2020) destributed in the Persian Gulf and the Oman Sea (Karkhaneh Yousefi, 2020). One of the valuable compounds in seaweeds is hydrocolloids. There are agar and carrageenan in red seaweed and alginic acid in brown one.

Sodium alginate is the sodium salt of alginic acid, a natural polysaccharide presents in brown seaweed cell walls (Kloareg and Quatrano, 1988). In principle, the isolating process of alginates includes stages of preextraction with acid, washing, filtration and neutralization with alkali. Lastly, sodium alginate is precipitated from the solution by alcohol and re-precipitated in the same way. Sodium alginate is widely used as a stabilizer, thickener in agri-food industry and viscosifying, rheological and gelling

properties it is applied in various industries like textile cosmetic. biomedical and pharmaceutical (Draget et al., 2009). The properties of the alginate vary between species, so the choice of which seaweeds to harvest is based on both the availability of particular species and the properties of the alginate that they contain. Total global aquatic plant aquaculture in 2016 was more than 15 billion USD, approximately 30 million tones, most comprises human food products, some carrageen, agar and alginate extraction (FAO, 2020).

The length of the Iran coastline, based on fractal scientists is less than 2440 kilometers along the Persian Gulf and Gulf of Oman (Wikipedia) which is propel for natural cultivated brown seaweed Sargassum spp. According to seaweed stock assessment projects were carried out by the Iranian Fisheries Sciences Research Institute, more than 3000 Mt of Sargassum illicifolium (Turner) C. Agardh is washed-up from the Oman Sea annually (Ajdari et al., 2003; Gharanjic and Rohani, 2010; Hafezieh et al., 2014). It is one of the best free resources for collecting/ harvesting and extracting hydrocolloids and sodium alginate based on previous laboratory analysis (Abkenar et al., 2014).

In order to determine total price production of sodium alginate extracted from *S. illisifolium*, this project was done in pilot commercial scale (100 kg wet seaweed *S. illisifolium*) to extraction, purification and price

production determination of sodium alginate, in Offshore Fisheries Research Center, Chabahar, Sistan and Baluchestan province, Iran, 2018.

Materials and methods

Sampling and preparing seaweed In order to avoid alginate destruction, brown seaweed must be collected, transported and dried as quickly as possible (Truus *et al.*, 2001). So, fresh samples of brown seaweed *S. ilicifolium* were collected from coastal line of Chabahar Bay, Oman Sea, Sistan and Baluchestan province, Iran (25°21'40"N 60°36'27"E), in November 2018 (Fig.

1). Then, they were cleaned and washed with sea water to remove impurities, transported to the Off-Shore Fisheries Research Center, Chabahar, rinsed with freshwater several times, dried under sun light, chopped and cut into the small pieces. Some parts of the samples were grinded to obtain powder with a particle size lower than 0.8 mm, using a mincer and passed through a 0.8 mm mesh sieve and then stored under vacuum in plastics bags at -20°C until of micro analyzing and macro nutritional compositions.

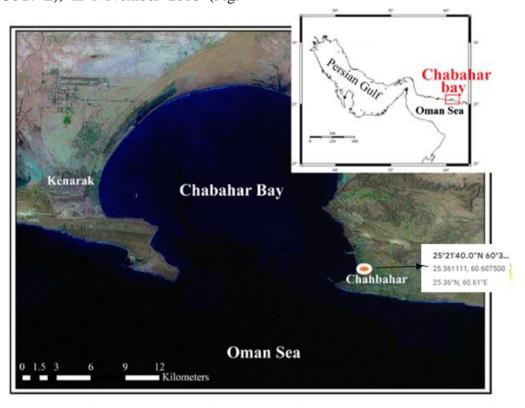


Figure 1: Sampling station in Chabahar Bay, Sistan and Baluchestan Province.

Determination of biochemical composition

Proximate composition of *Sargassum* meal including dry matter, crude protein and fiber, ash, neutral detergent

fiber (NDF) and acid detergent fiber (ADF) was determined according to the procedures of the Association of Official Analytical Chemists (AOAC, 2009). Moisture, protein, and ash

contents were determined following the ISO recommendations (ISO 936:1998). Dry matter (DM) was determined within 3 g weight change calculation before and after 105 °C drying, crude protein (CP) was determined by Kjeldahl total nitrogen method (total nitrogen content was multiplied by×6.25). For this, 500 mg of seaweed with reacted catalyst H₂SO₄ (CuSO₄·5H₂O) in a digester, organic nitrogen was transformed into (NH₄)₂SO₄ and distilled in alkali condition. Amino acids were extracted following the method provided by determined by ethyl ether extraction (Soxhlet technique) and 50 mg of lipid extraction was used for fatty acids profile determination. Trans esterification (Domínguez et al., 2015), equipment with a FAMEs expressed in g/100 g of FAME. Ash was measured after drying in muffle oven at 500°C. Minerals (Ca, Fe, K, Mg, Mn, Na, P, Zn and Cu) were measured using flame photometry.

Extraction of sodium alginate

Alginic acid is present in brown seaweeds mainly in the form of sodium and calcium salts. The purpose of the extraction step is to convert the alginate to the soluble form of sodium alginate and remove it from the algae (Hernández-Carmona *et al.*, 2002). Sodium alginate was extracted from chopped and cut seaweed, chemically at 40°, using 0.5% formalin for 2 hours, rinsed with freshwater then placed in

0.2 N sulfuric acid for 5 hours, rinsed again to obtain pH 7, and using 3% sodium carbonate for 6 hours, then it was filtered. After adding ethylic alcohol. the viscous mixture separated from its residue bv centrifugation at 14,000×g. A paste form sediment which has been dried to produce clod form, was powdered by grounder to obtain sodium alginate extraction (Larsen et al., 2003; Torres et al., 2007). The yield of alginate was extracted as percentage/ dry weight.

Purification monitoring by fluorescence spectroscopy

To follow the purification procedure, fluorescence spectroscopy was used. Alginates are strongly fluorescent due to small amounts of polyphenolic residues. This is a routine technique to measure these contaminants in a wide range of alginates. The spectra were obtained with USB2000-FLG spectrofluorometer following the method described by Klock *et al.* (1997).

Results

Biochemical Compositions of Sargassum illicifolium

The proximate chemical compositions of seaweed meal were determined through laboratory analysis as shown in Table 1. The minerals, essential amino acids and fatty acids profiles (mean± standard deviation values) (n=5, five replicates) are given in Table 2.

Table 1: Biochemical compositions of *S. ilicifolium* collected from Chabahar Bay in November 2018 (Mean ± SD. % dry matter, DM).

2010 (1120m = 52) / 0 mi j matter)							
Parameter	Dry matter* (moisture)	Crude Protein	Ash	NDF	ADF	Crude lipid	Crude fiber
Sargassum meal	91.0±1/1 (9.0±0/1)	9.8±0.8	41.6±2.27	26.4±1.80	12.1±0.89	4.4±0.18	38.5±0.4

Note: * Values are in % DM; NDF, neutral detergent fiber; ADF, acid detergent fiber.

Table 2: Minerals (mg/g), essential amino acids and fatty acids profiles (mg/kg) of *S. illicifolium* collected from Chabahar Bay in November 2018 (Mean \pm SD, n = 5, five replicate specimens).

Minerals	Ca	Fe	K	Mg	Mn	Na	P	Zn	Cu
	98.7 ±	13.3 ±	378.3 ±	86.8 ±	1.9 ±	457.7 + 50.0	n.q.	n.q.	n.q.
	47.2	0.9	13.4	12.0	0.7	457.7 ± 50.0			
Essential Amino	Threonine	Valine	Methionine	Isoleucine	Leucine	Phenylalanine	Lysine	Histidine	Arginine
Acids	(% total AA)								
$(2912.42 \pm 204.93$	363.2 ±	353.8 ±	147.5 ±	295.2 ±	537.3 ±	340.1+ 17.74	431.7 ±	126.4 ±	316.7 ±
mg/100 g DW)	17.1	32.9	18.7	25.7	38.8	340.1± 17.74	38.4	10.6	14.0
Fatty Acids	Saturate acid		Mono- unsaturated fatty acids	Poly-uns		Omega 3 fatty acid	_	a 6 fatty cids	n-6/n-3
	25.1 ±	0.5	31.1 ± 0.2	43.5 -	± 0.5	12.0 ± 0.1	31.4	1 ± 0.4	2.6 ± 0.0

Impurity monitoring by fluorescence spectroscopy

One of the most important characters of alginate extracted from seaweed is purification rate which is calculated after impurities determination. purification is crucial for alginate applications in the biomedical field, since this natural polymer is known to be largely impure alginates that can lead to the development of fibrotic cell over growth around alginate microcapsules and be consequently responsible for side effects on humans. The principal alginate contaminants are polyphenols, endotoxins, and proteins.

S. illicifolium alginate impurities was 71.80%, so its purification was 28.20%.

Sodium alginate contents of S. illicifolium and production cost

Ninety percent of the wet seaweed disappeared after drying which was measured as moisture. From 100 g rest powdered seaweed, it can be extracted 28.2 g sodium alginate as bleached powder. Thus, from 100 kg wet seaweed, it can be obtained 10 kg DW and 2.82 kg sodium alginate. Total production cost for production one kg sodium alginate is detailed in Table 3.

Table 3: The final production price for one kg feed grade sodium alginate extracted from Sargassum ilicifolium collected from Chabahar Bay (Sistan and Baluchestan Province).

	Surgussum uniquum conected from Chabanar Bay (Sistan and Baidchestan Fronnice).					
	Procedure	Amount	The cost \$			
1	Collecting, rinsing and drying	one kg DW seaweed	1.14 \$			
2	Chemical including formalin, sulfuric acid and sodium carbonate, ethylic alcohol and bleaching	2.5 lit.	3 \$			
6	Drying, powdering and packing	Produced one kg sodium alginate	1.2 \$			
7	Electricity	Produced one kg sodium alginate	0.66 \$			
8	Water supplying	Produced one kg sodium alginate	0.33 \$			
9	Workers	Produced one kg sodium alginate	1.33 \$			
10	Total production cost	Produced one kg sodium alginate	7.66 \$			
11	Price in market (Chinese brand)	One kg	11.6\$			
12	Benefit of local production		4\$			

Discussion

Nine percent of the content of this seaweed was moisture and 91.0±1/1 DM, which is in close agreement with the data reported by Rodrigues et al. (2015), who also noticed that the moisture content of different edible seaweeds species ranged from 8 to 10%. Gómez-Ordoñez et al. (2010) also reported similar moisture contents (between 8.64% and 9.86%) in seaweeds from the northwestern Spanish coast. However, the differences in the result between this project with Chan and Matanjun (2017) in freezedried Gracilaria changii seaweed (lower moisture content, 5.32%) is due to difference in seaweed species.

S. illicifolium species presented total protein content of 9.8±0.8% DW, which is completely in agreement with the data reported by Fleurence (1999) (<15% DW in F. vesiculosus, A. nodosum, Laminaria digitata and Himanthalia elongata). Similar values were found by Gómez-

Ordoñez et al. (2010) and Alves et al. (2016) in B. bifurcata (10.92% DW and 8.57% DW, respectively) and Chan and Matanjun (2017) in G. changii (12.57% DW), but it was lower than those obtained by Rodrigues et al. (2015) for brown (14.4-16.9% DW), red (20.2-23.8% DW) and green (18.8% DW) seaweeds and the observations by Fleurence (1999) in other seaweed species such as Porphyra tenera (47% and Palmaria DW) palmata (35% DW). On the contrary, Sánchez-Machado et al. (2004) obtained lower protein content (5.46% DW) in H. elongata dried seaweed. The protein level varied among different algal species, geographic areas, seasons, or environmental conditions (Denis et al., 2010).

Ash contents of *S. illicifolium* was 41.6±2.27 % DW which is in agreement with the data reported as 40.58±2.10% DM by Alves *et al.* (2016) and 42.01±1.78% DM by Gómez-Ordoñez *et al.* (2010), but higher than that

reported by Peinado *et al.* (2014) in *F. vesiculosus* (21–19% DW). It is known that high amounts of ash are linked with high levels of minerals. Mineral salts could be found on surface and in thallus. Conditions of hydrology and hydrochemistry on the habitat also influence the ash content.

NDF (26.4±1.80% DM) and ADF (12.1±0.89% DM) were obtained in *S.illicifolium* fiber analysis. Similarly, Peinado *et al.* (2014) recorded NDF and ADF in *Sargassum tenerrium* as 27.90±1.09% DM and 11.0±1.10% DM and also Alves *et al.* (2016) in *B. bifurcata* (25.78±1.26% DM and 12.98±0.27% DM, respectively)

Seaweeds exhibit low fat content (bellow 4%) (Herbreteau et al., 1997). and varies significantly through the year (Manivannan et al., 2008). Our value $(4.4\pm0.18 \% DW)$ was similar to those reported by Peinado et al. (2014) from 3.95 to 4.64% DW in F. vesiculosus at different seasons and by Gómez-Ordoñez et al. (2010) and Alves et al. (2016), who observed fat levels of 5.67% DW and 5.81% DW in B. bifurcata, respectively. The World Health Organization (WHO) (2007) recommended a n-6/n-3 ratio below 10. In our study, we observed n-6/n-3 ratio of 2.62 placing this studied brown seaweed according to WHO recommendations. This outcome is in agreement with those reported by other authors (; Alves et al., 2016; Chan. and Matanjun 2017) who found n-6/n-3 ratios between 4.1 and 0.02.

The carbohydrate content or crude fiber of dried brown seaweeds ranged from 21.93% to 56.75%. In S. illicifolium collected from Chabahar Bay it was 38.5±0.4% DW. The maximum carbohydrate content was recorded in Colpomenia implexa and Lobophora variegata found the minimum content. Similarly, Peinado etal.(2014)carbohydrate recorded high in Sargassum tenerrium as 67.90% DW.

Seaweeds, especially brown ones are usually eaten whole plants as a good source of minerals. In this research, Ca (98.7 ± 47.2) , Fe (13.3 ± 0.9) , K (378.3 ± 13.4) , Mg (86.8 ± 12.0) , (1.9 ± 0.7) and Na (457.7 ± 50.0) were measured in S.illicifolium based on mg/g but P, Zn and Cu were not detected. Kasimala and Coworkers (2015) revealed that Hypneu pannosa had Na content (127.65 mg/g), and Padina tenuis had Ca (48.00 mg/g), Mg (44.13 mg/g), and Fe (6.64 mg/g). The differences between minerals contents of seaweeds mainly referred to species, the habitatswhere they grow and the water content of minerals (Mæhre et al., 2014). The total EAAs content of 2912.42±204.93 mg/100 g DW for S. illicifolium, accords with other finding $(3000.81\pm194.67 \text{ mg}/100 \text{ g} \text{ DW})$ by Chan and Matanjun (2017).

In fatty acids profile of the seaweed studied, polyunsaturated fatty acids (PUFAs) were the most abundant (43.47% for the *S. illicifolium*), which is in line with the data reported by the other authors (Cofrades *et al.*, 2010; Alves *et al.*, 2016; Chan and Matanjun,

2017) who found that PUFAs were the main fatty acids (more than 40%) in seaweeds. However, Peng *et al.* (2013) and Maehre *et al.* (2014) observed higher saturated fatty acids (SFA) content in different seaweed species.

Sodium alginate content S. illicifolium collected from Chabahar Bay has 28.2% purity; regularly this active compound has purities ranging from 20 to 35% in different brown seaweed due to seasonal harvesting, water temperature and other physicochemical parameters of surrounded water (Alves et al., 2016). According to Viswanathan and Nallamuthu (2014) the purification of sodium alginate in P. gymnospora and Colpomenia implexa was 23.01% and 21.53%, respectively.

illicifolium Sargassum alginate impurities was 71.20% which this result is consistent with the values reported by Orive et al. (2002) for Sargassum illicifolium sodium alginate (63%) and Torres et al. (2007) for Sargassum vulgarea brown algae for which the intensity was reduced by 52.7%. Klock et al. (1994) noted that the remaining contaminants detected in the fluorescence spectra of alginates from Durvillaea potatorum (brown algae) could not be identified. The in vitro and in vivo biocompatibility tests showed that these impurities did not initiate a foreign body reaction (Klock et al., 1997)

Conclusions

In this study, *S. illicifolium* brown seaweed was collected from the Oman Sea, Chabahar Bay, prepared for

determination chemical composition and extraction of sodium alginate in the laboratory condition. We found that this had species 9.8 ± 0.8 . 41.6±2.27. 26.4±1.80, 12.1±0.89, 8.4±0.38 % DW of CP, ash, NDF, ADF, and CF, respectively. From one kilogram cleaned seaweed after proportional dehydration only 10% DW obtained and 28.2 g sodium alginate can be extracted. Thus, from 100 kg wet seaweed, it can be obtained 10 kg seaweed DW and 2.82 kg sodium alginate. Total economical production cost is estimated 7.66 \$, compared to imported Chinese brand 11 \$ based on data from imported

Acknowledgements

The authors acknowledge the Iranian Fisheries Sciences Research Institute for financial supporting, Chabahar Offshore Fisheries Research Center, for organizing the project, collecting seaweed and laboratory analyzing.

References

Abkenar, A.M., Gharanjik, B.M. and Azini, A., 2014. Alginate from seaweed, Final project report Iranian Fisheries Research Organization Project. 98 P.

Ajdari, H., Ajdar, Z., Abkenar, A.M. and Gharanjik, B.M., 2003. Stock seaweed back to the costal on Oman Sea. Final project report Iranian Fisheries Research Organization Project. 120 P.

Alves, C., Pinteus, S., Simões, T., Horta, A., Silva, J., Tecelão, C. and Pedrosa, R., 2016 Bifurcaria bifurcata: A key macro-alga as a source of bioactive compounds and functional ingredients. International Journal of Food Science Technology, 51, 1638–1646.

Doi: 10.1111/ijfs.13135.

AOAC, 2009. Official Methods of Analysis of the Association of Official Analytical Chemists, 15th ed. Washington, pp. 456-579.

Chan, P.T. and Matanjun, P., 2017.

Chemical composition and physicochemical properties of tropical red seaweed, *Gracilaria changii*. Food Chemistry, 221, 302–310.

Doi: 10.1016/j.foodchem.2016.10.066.

Cofrades, S., López-Lopez, I., Bravo, L., Ruiz-Capillas, C., Bastida, S., M.T. and Jiménez-Larrea, Colmenero, F., 2010. Nutritional and antioxidant properties different brown and red Spanish edible seaweeds. Food *Science and Technology* International, 16, 361–370. Doi: 10.1177/1082013210367049.

Denis, C., Morançais, M., Li, M., Deniaud, E., Gaudin, P., Wielgosz-Collin, G., Barnathan, G., Jaouen, P. and Fleurence, J., 2010. Study of the chemical composition of edible red macroalgae *Grateloupia turuturu* from Brittany (France). *Food Chemistry*, 119, 913–917.

Doi: 10.1016/j.foodchem.07.047.

Domínguez, R., Borrajo, P. and Lorenzo, J.M., 2015. The effect of

cooking methods on nutritional value of foal meat. *Journal of Food Composition and Analysis*, 43, 61–67. Doi: 10.1016/j.jfca.04.007.

Draget, K.I. and Taulor, C., 2009. Chemical, physical and biological properties of alginates and their biomedical implications Doi.org/10.1016/j.foodhyd.10.007. *Food Hydrocolloids*, 25, 2, pp. 251-256.

FAO/WHO/UNU Expert Consultation, 2007. Protein and Amino Acids Requirements in Human Nutrition. FAO/WHO/UNU; Rome, Italy: Amino acid requirements of adults; pp. 135–159.

FAO. 2020. The State of World Fisheries and Aquaculture 2020. Sustainability in action. Rome.https://doi.org/10.4060/ca9229en

Fleurence, J., 1999. Seaweed proteins:
Biochemical, nutritional aspects and potential uses. *Trends in Food Science* and *Technology*, 10, 25–28.
Doi: 10.1016/S0924-2244(99)00015-1.

Gharanjic, B.M. and Rohani, K., 2010. Atlas of marine seaweed of the Persian Gulf and Oman sea beaches. Iranian Fisheries Research Organization. 202 P.

Gómez-Ordóñez, E., Jiménez-Escrig, A. and Rupérez, P., 2010. Dietary fiber and physicochemical properties of several edible seaweeds from the northwestern Spanish coast. *Food Research International*, 43, 2289–2294.

- Doi: 10.1016/j.foodres.2010.08.005.
- Hafezieh, M., Ajdari, D., Ajdehakosh Por, A. and Hosseini, S.H., 2014.

 Using Oman Sea Sargassum illicifolium meal for feeding white leg shrimp Litopenaeus vannamei.

 Iranian Journal of Fisheries Sciences, 13(1), 73-80.
- Herbreteau, F., Coiffard, L.J.M., Derrien, A. and De Roeck-Holtzhauer, Y., 1997. The fatty acid composition of five species of macroalgae. *Botanica Marina*, 40, 25–27.
- Doi: 10.1515/botm.1997.40.1-6.25.
- Hernández-Carmona, G., McHugh, D.J., Arvizu-Higuera, D.L. and Rodríguez Montesinos, Y.E., 2002. Pilot plant scale extraction of alginates from *Macrocystis pyrifera*. 4. Conversion of alginic acid to sodium alginate, drying and milling. *Journal of Applied Phycology*, 14, 445–451.
 - Doi.org/10.1023/A:1022372807813
- ISO, 1998. Determination of Ash Content, ISO 936:1998 Standard. International Standards Meat and Meat Products. International Organization for Standardization; Genéve, Switzerland: 1998.
- **Ito, K. and Hori, K., 1989**. Seaweed: Chemical composition and potential food uses. *Food Reviews International*, 5, 101–144. Doi: 10.1080/87559128909540845
- Kasimala, M.B., Mebrahtu, L., PiusMagoha, P. and Asgedom, Gh.,2015. A review on biochemical composition and nutritional aspects

- of seaweeds. Caribbean journal of Science and Technology, 3, 789-797.
- Karkhaneh Yousefi, M., **Seved** Hashtroudi. M., Mashinchian Moradi, A. and Ghasempour, A., 2020. Seasonal variation fucoxanthin content in four species of brown seaweeds from Qeshm Island, Persian Gulf and evaluation of their antibacterial and antioxidant activities. Iranian Journal Fisheries Sciences, 19(5), 2394-2408
- Kloareg, B. and Quatrano, R.S., 1988. Structure of the Cell Walls of Marine Algae and Ecophysiological Functions of the Matrix Polysaccharides. *Oceanography and Marine Biology: An Annual Review*, 26, 259-315.
- Klöck, G., Pfeffermann, A., Ryser, C., Gröhn, P., Kuttler, B., Hahn, H.J. and Zimmermann, U., 1997. Biocompatibility of mannuronic acid-richalginates. *Biomaterials*, 18(10), 707-713.
- Klöck, G., Frank, H., Houben, R., Zekorn, T., Horcher, A., Siebers, U., Wöhrle, M., Federlin, K. and Zimmermann, U., 1994. Production of purified alginates suitable for use in immunoisolated transplantation. *Applied Microbiology* and *Biotechnology*, 40(5), 638-643.
- Larsen, B., Salem, D.M., Sallam, M.A., Mishrikey, M.M. and Beltagy, A.I., 2003. Characterization of the alginates from algae harvested at the Egyptian Red Sea

- coast. Carbohydrate Research, 338(22), 2325-2336.
- Mæhre. H.K.. Malde. M.K.. Eilertsen, K.E. and Elvevoll, E.O., 2014. Characterization of protein, lipid mineral contents and common Norwegian seaweeds and evaluation of their potential as food and feed. Journal of the Science of Food and Agriculture, 94, 3281–3290.

Doi: 10.1002/jsfa.6681.

- Manivannan, K., Thirumaran, G., Devi, G.K., Hemalatha, A. and Anantharaman, P. 2008. **Biochemical** composition of seaweeds from Mandapam coastal regions along Southeast Coast of India. American Journal ofBotany, 1, 32–37.
- McHugh, D.J., 1987. Production. properties and uses of alginates. FAO Fishing Technology, 288, 58-115.
- Orive, G., Ponce, S., Hernandez, R.M., Gascon, A.R., Igartua, M. and Pedraz, J.L., 2002. Biocompatibility of microcapsules for cell immobilization elaborated with different type of alginates. Biomaterials, 23(18), 3825-3831.Doi:10.1016/S0142-9612(02)00118-7
- Peinado, I., Girón, J., Koutsidis, G. and Ames, J.M., 2014. Chemical composition, antioxidant activity and sensory evaluation of five different species of brown edible seaweeds. Food Research

- 36-44. International, 66, Doi: 10.1016/j.foodres.2014.08.035.
- Peng. Y., Xie, E., Zheng, K., Fredimoses, M., Yang, X., Zhou, X., Wang, Y., Yang, B., Lin, X., Liu, J., et al. 2013. Nutritional and chemical composition and antiviral of cultivated activity seaweed Sargassum naozhouense Tseng et Lu. Marine Drugs, 11, 20-32.Doi: 10.3390/md11010020.
- Rodrigues, D., Freitas, A.C., Pereira, Rocha-Santos, Vasconcelos, M.W., Roriz, Rodríguez-Alcalá, L.M., Gomes, **A.M.P.** and **Duarte**, **A.C.**, 2015. Chemical composition of red, brown and green macroalgae from Buarcos bay in Central West Coast of Portugal. Food Chemistry, 183,197– 207.

Doi: 10.1016/j.foodchem.2015.03.057.

- Saini, R., Badole, S.L. and Zanwar, **A.A., 2013**. In: Bioactive Dietary and Plant Extracts Dermatology. Watson R.R., Zibadi S., editors. Humana Press; Totowa, NJ, USA: 2013. pp. 73–82
- Sánchez-Machado, **D.I.**, López-Cervantes, J., López-Hernández, J., Paseiro-Losada, P. and Simal-Lozano, J., 2004. Determination of the uronic acid composition of seaweed dietary fibre by HPLC. Biomedical Chromatography, 18, 90–97. Doi:
 - 10.1002/bmc.297.
- Torres, M.R., Sousa, A.P., Silva Filho, E.A., Melo, D.F., Feitosa,

J.P., de Paula, R.C. and Lima, M.G., 2007. Extraction and physicochemical characterization of Sargassum vulgare alginate from Brazil. *Carbohydrate*Research, 342(14), 2067-2074.

DOI:10.1016/j.carres.2007.05.022

Truus, K., Vaher, M. and Taure, I., 2001. Algal biomass from *Fucus vesiculosus* (Phaeophyta):

investigation of the mineral and alginate components'. *Proceedings* of the Estonian Academy of Sciences, 50(2), 95–103.

Viswanathan, S. and Nallamuthu, T., 2014. Extraction of sodium alginate from selected seaweeds and their physiochemical and biochemical properties. *Extraction*, 3(4).