

Planktonic-based assessment of the landside-dammed lake (Erzurum-Turkey)

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Abstract

The aim of this study was to identify the variation of plankton communities in the Tortum Lake. Changes in phytoplankton and zooplankton communities in relation to the abiotic environment were analyzed using multivariate analysis. Water samples were taken monthly from three sampling points of the Tortum Lake between June 2012 and May 2013. Water temperature (5.28-23.05°C), dissolved oxygen (1.54-13.68 mgL⁻¹), and pH (7.22-9.01) were measured in situ. Chlorophyll-a and total orthophosphate concentrations ranged from 0.18 to 5.70 mgL⁻¹ and from 0.01 to 0.00 mgL⁻¹, respectively. In the Tortum Lake, *Ceratium hirundinella* (18%), *Botryococcus braunii* (51%), *Chlamydomonas microsphaerella* (25%), *Microcystis aeruginosa* (7%), *Melosira varians* (1%), *Monoraphidium contortum* (1%), Copepoda (66%), *Daphnia* (33%) and *Keratella* (1%) were found. Some species such as *M. aeruginosa* were increased by organic and inorganic pollution in Tortum Lake.

Keyword: Phytoplankton, Zooplankton, Biodiversity index, Tortum Lake, Multivariate analysis

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Introduction

The composition and biomass of phytoplankton and zooplankton species in lakes depend on a complex combination of factors, such as temperature, light and availability of nutrients (Dantas *et al.*, 2012). Seasonal and spatial variations of plankton composition are affected by coastal structure, top-and bottom currents and predators (Tanyolac, 2009).

Phytoplankton use as orthophosphate ions for growth so that it is responded to decreased phosphorus. This is partly due to community resilience (Padisak *et al.*, 2003).

Zooplankton grazing usually provokes a decrease in phytoplankton biomass; however, some inedible algae may increase their abundances in a lake during active grazing phases because of the effect of the selective feeding, and therefore, they can take advantage of the availability of nutrients when the competition pressure with counterpart algae diminishes (Queimalin *et al.*, 1998). Zooplankton composition, in turn, also determines the responses of the grazing pressure on phytoplankton. Particularly, microphagous and macrophagous zooplankton may exert a different top-down impact on the phytoplankton community (Sommer *et al.*, 2003).

Diversity indices such as Shanon-Weaner index appeared to detect significant differences in the structure of the communities (Offem *et al.*, 2011).

The three main categories of zooplankton found in Minneapolis lakes are rotifers, copepods and cladocerans. Rotifers tend to be the smallest among the types. Despite their small size, they are important in the aquatic food web because of their abundance, distribution and a wide range of feeding habits. Copepods and cladocerans are larger zooplankton and members of the class of Crustacea. Copepods are the most diverse group of crustaceans. Rotifer plays an important role in aquatic ecosystems mainly because of their enormous reproductive potential. There is a negative relation between the ratio of rotifers and macrozooplankton as versus the ratio of small algae (Bronmark and Hansson, 2005)

In the Kuzgun Reservoir, Bacillariophyta was the dominant group, followed by Chlorophyta and Dinophyta. The dominant species were *Synedra delicatissima*, *Asterionella formosa*, *Fragilaria crotonensis*, *Cyclotella kiitzingiana*, *Cyclotella ocellata*, *Oocystis borgei*, *Staurastrum longiradiatum*, *Ankistrodesmus falcatus*, *Ceratium hirundinella*, and *Peridinium inctum*. Maximum phytoplankton density was observed in late spring (Gurbuz *et al.*, 2004).

According to Demir *et al.* (2013), the examination of functional groups of phytoplankton communities in Lake Mogan seemed to be a useful method for ecological status and may provide evidence for further examinations between the Q quality index and the

ecological condition of other Turkish lakes.

Tortum Lake, the biggest landslide lake with surface area 6.63 km², located in the East Anatolia Region of Turkey and 92 km from the Erzurum city (Altuner, 1982; Orhan and Karahan, 2010). The aims of the present study were to determinate the changes of phytoplankton composition by using multivariate analysis with zooplankton composition and some physico-chemical data.

Materials and methods

Study site

The Tortum Lake is located in the northeast part of Turkey. The lake with 11 km length and 0.77 and 1 km width, is at 1000 m above sea level. The lake has an area of 6.77 m², the volume of 223×10^6 m³, an average depth of 110 m. The amount of sediment reaching the basin of the Tortum Lake is

estimated to be 2.5 million m³. The lake vanishes quickly as a result of sedimentation and some calculations revealed that the lake will be completely disappeared in 250-300 years (Altuner, 1982; Kivrak, 2006).

Sampling and laboratory procedures

Plankton and water samples were monthly collected from 3 different stations (1st site 40° 37' 10" N and 40° 37' 37" E; 2ed site 40° 37' 6" N and 41° 37' 35" E; 3rd site 40° 39' 7" N and 41° 39' 29" E) between June 2012 and May 2013 (Fig. 1). Water temperature (°C) (Thermo Orion 3 Star), dissolved oxygen (mgL⁻¹) (DO) (Thermo Orion 3 Star) and pH (Thermo Orion 3 Star) were measured in situ. Chlorophyll-a concentration (mgL⁻¹) was determined by the acetone extraction method using a spectrophotometer (Beckman Coulter DU 730) (Strickland and Parsons, 1972).

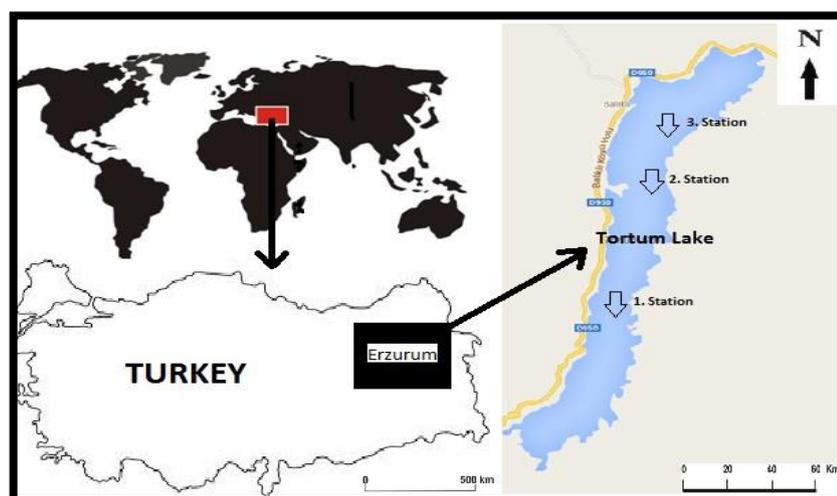


Figure 1: Map of the sampling site locations.

Total hardness ($\text{mgL}^{-1}\text{CaCO}_3$), Ca-hardness ($\text{mgL}^{-1}\text{CaCO}_3$), Mg-hardness ($\text{mgL}^{-1}\text{CaCO}_3$) and total orthophosphate ($\text{PO}_4\text{-P}$, mgL^{-1}) were calculated by standard methods (APHA, 1995).

Phytoplankton samples were fixed using Lugol's iodine. Zooplankton samples were fixed in 4% formaldehyde. The common taxonomic literature was used for phytoplankton taxa (Krieger, 1932; Round, 1953; Cramer, 1991; Kelly, 1997; John *et al.*, 2002). Zooplankton taxa were identified according to Smirnov (1974), Segers (1993) and Dussart (1969). Phytoplankton counts were carried out by the inverted microscope (Utermohl, 1958). Cell dimension of algae was measured with a Zeiss microscope. Total phytoplankton bio-volume was estimated by the corresponding geometrical form (Hillebrand *et al.*, 1999; Sun and Liu, 2003) using the $1\text{ mm}^3\text{ m}^{-3}$ of algal volume to $1\text{ mg wet weight m}^{-3}$ biomass calculation. Cell dimension of zooplankton was

calculated with the stereomicroscope. Total zooplankton bio-volume was calculated the corresponding geometrical form (Akbay, 1982).

Biodiversity indices

Shannon-Weiner (H') index was considered for the present study. This index is applied to biological systems which are derived from a mathematical formula by Shannon (1948) (Turkmen and Kazancı, 2010):

$$H' = -\sum_{i=1}^S p_i \log_e p_i$$

where: p_i : n_i/n

s : a total number of species and

p_i : number of individuals belonging to i species (n_i) / total number of individuals (n) (Hill, 1973; Krebs, 1998; Kwak and Peterson, 2007; James and Aderaje, 2010).

Statistical analysis

The relationship between environmental variables and phytoplankton and zooplankton assemblages was analyzed using canonical correspondence analysis (CCA). CCA is useful for

identifying environmental variables which are important in determination of community composition and the role of spatial variation in the communities (Black *et al.*, 2004). In the multivariate analysis, the matrix abiotic data, phytoplankton communities and zooplankton communities were accounted for each station using the XLSTAT program (Braak and Smilauer, 2002). For hierarchical cluster analysis, the similarity between species and sites were calculated and a one-way ANOVA test was used to find the statistical differences in the physical and chemical variables using the SPSS software (version 20).

Results

In this study, it was observed the positive interaction between water quality parameters and plankton biomass in the Tortum Lake.

Chlorophyll-a ranged from 5.70 to 0.18 mgL⁻¹, pH from 9.01 to 7.22, temperature from 23.05 to 5.28 °C, dissolved oxygen from 13.68 to 1.54 mgL⁻¹, total orthophosphate from 0.01 to 0.00 mgL⁻¹, total hardness from 16.90 to 9.4 mgL⁻¹ CaCO₃, Mg-hardness from 27.17 to 6.42 mgL⁻¹ CaCO₃, and Ca-hardness from 9.64 to 5.72 mgL⁻¹ CaCO₃, respectively (Table 1).

In our study, *C. hirundinella* (18%), *B. braunii* (51%), *C. microsphaerella* (25%), *M. aeruginosa* (7%), *M. varians* (1%), *M. contortum* (1%), Copepoda (66%), *Daphnia* (33%) and *Keratella*

(1%) were found. In addition, phytoplankton biomass was calculated between 0.12 mgL⁻¹ and 34.19 mgL⁻¹ and zooplankton biomass range 0.08 mgL⁻¹ to 36.72 mgL⁻¹ (Fig. 3).

Chlorophyll-a value reached a peak during the months not only in March but also in June. Even though *B. braunii* and *C. microsphaerella* were reduced on phytoplankton biomass; *M. aeruginosa* was increased in phytoplankton biomass between January and April (Fig. 3).

The result of CCA based on eight variables are given in the table and illustrated in the figure. The proportion of species variance is explained by each axis. For ecological data the percentage of explained variance is usually low. The plankton communication had an eigen-value of 0.26 explaining 99.4% and environmental parameters had 0.001 explaining 0.46% variability (Tables 2 and 3).

Discussion

The present study showed that phytoplankton and zooplankton biomass was affected by temporal and spatial changes of water quality parameters (Fig. 2). According to of the obtained data on Chlorophyll-a, pH, temperature, dissolved oxygen (DO), total orthophosphate (PO₄-P), total hardness (TH), Mg-hardness and Ca-hardness, Tortum Lake is classified as the oligotrophic lake (Table 1) (Wetzel, 2001).

Kıvrak (2006) found *C. hirundinella*, *C. krammeri*, *C. glomerata* and *C. microsphaerella* in the Tortum Lake between 2002 and 2003. In addition, Bacillariophyta and Cyanobacteria (*M. aeruginosa*) were identified in 1982 (Akbaş, 1982). In our study, *B. braunii* was identified the highest level in the lake, whereas *M. varians* and *M. contortum* were found the lower level (Fig. 3). Phytoplankton composition seems to respond quickly not only to the seasonal changes of environmental

parameters but to anthropogenic disturbances. Phytoplankton communities are located in a competitive area and changes in water quality lead to the formation of high compositional diversity (Scheffer *et al.*, 2003).

The variance of sample scores on each axis reflects the importance of the axis as measured by the mean value whereas the variances of the species scores along the axes are equal (Braak and Verdonschot, 1995).

Table 1: The simple statistic for physico-chemical parameters of Lake Tortum.

| Parameter | Site | Mean | SD | Min. | Max. |
|---|------|-------|------|-------|-------|
| Chl-a (mgL ⁻¹)* | 1 | 1.66 | 1.53 | .38 | 5.30 |
| | 2 | 1.81 | 1.89 | .18 | 5.70 |
| | 3 | 1.08 | .69 | .37 | 2.48 |
| Temp (°C) | 1 | 13.12 | 5.32 | 5.95 | 23.05 |
| | 2 | 13.21 | 5.55 | 5.28 | 19.73 |
| | 3 | 13.44 | 5.07 | 5.55 | 19.53 |
| PO ₄ -P (mgL ⁻¹) | 1 | .004 | .003 | .00 | .01 |
| | 2 | .003 | .004 | .00 | .01 |
| | 3 | .003 | .004 | .00 | .01 |
| DO (mgL ⁻¹) | 1 | 10.06 | 3.81 | 1.54 | 13.48 |
| | 2 | 10.60 | 3.22 | 3.38 | 13.90 |
| | 3 | 10.67 | 3.51 | 4.17 | 13.68 |
| pH | 1 | 8.47 | .52 | 7.22 | 9.01 |
| | 2 | 8.54 | .24 | 8.21 | 8.94 |
| | 3 | 8.55 | .20 | 8.23 | 8.86 |
| Total Hardness (mgL ⁻¹ CaCO ₃) | 1 | 11.95 | 2.54 | 9.40 | 16.90 |
| | 2 | 12.25 | 1.87 | 10.10 | 15.88 |
| | 3 | 12.42 | 2.11 | 10.68 | 16.90 |
| Mg-Hardness (mgL ⁻¹ CaCO ₃) | 1 | 13.08 | 5.35 | 6.42 | 24.64 |
| | 2 | 12.62 | 3.63 | 8.65 | 22.26 |
| | 3 | 13.65 | 4.82 | 8.94 | 27.17 |
| Ca-Hardness (mgL ⁻¹ CaCO ₃) | 1 | 7.04 | .84 | 5.80 | 8.52 |
| | 2 | 7.20 | .88 | 5.94 | 9.16 |
| | 3 | 6.82 | 1.05 | 5.72 | 9.64 |

*Max. maximum. Min. minimum. SD standard deviation. $p < 0.05$.

Table 2: Summary statistics for canonical correspondence analysis (CCA).

| | F1 | F2 | F3 | F4 |
|-------------------------|--------|--------|--------|--------|
| Eigenvalue | 0.267 | 0.001 | 0.000 | 0.000 |
| Constrained inertia (%) | 99.400 | 0.457 | 0.131 | 0.009 |
| Cumulative % | 99.400 | 99.857 | 99.988 | 99.996 |
| Total inertia | 33.697 | 0.155 | 0.044 | 0.003 |
| Cumulative % (%) | 33.697 | 33.852 | 33.897 | 33.900 |

The first axis was associated with Ca hardness, DO and pH, while the second axis was related to PO₄-P, total hardness, Mg hardness and water temperature. *M. varians* and Copepoda were positioned close to the center of ordination diagram. *M. aeruginosa* and Keratella were positioned on the positive side of the first axis, while *C.hirundinella* and *M.contortum* were positioned on the negative side of the second axis. The Chlorophyll-a in water

surface remained the lowest during the fall season in Tortum Lake (Fig. 3). The Chlorophyll-a in water surface remained the highest during the summer season in all stations except the Outfall Bay, where the highest value was recorded in winter (Abdul Azis *et al.*, 2003) (Figs. 5 and 6).

Chlorophyll-a is the primary photosynthetic pigment contained in algae.

Table 3: Canonical Correlation Analysis (CCA) of biotic and abiotic variations. The species names are abbreviated to the part in italic as follows: *Ceratium hirundinella* (CER HIR), *Botryococcus braunii* (BOT BRA), *Chlamydomonas microsphaerella* (CHL MIC), *Microcystis aeruginosa* (MIC AER), *Melosira varians* (MEL VAR), *Monoraphidium contortum* (MON CON), Copepoda (COPE), Daphnia (DAPH) and Keratella (KER).

| Variables | BOT BRA | MIC AER | CER HIR | CHL MIC | MEL VAR | MON CON | DAPH | COPE | KER |
|-------------|---------|---------|---------|---------|---------|---------|--------|--------|--------|
| BOT BRA | 1 | 0.789 | 0.439 | 0.685 | -0.064 | 0.207 | -0.013 | 0.157 | -0.079 |
| MIC AER | 0.789 | 1 | 0.392 | 0.718 | -0.039 | -0.034 | -0.088 | -0.068 | -0.029 |
| CER HIR | 0.439 | 0.392 | 1 | 0.662 | 0.040 | -0.060 | -0.101 | -0.078 | 0.071 |
| CHL MIC | 0.685 | 0.718 | 0.662 | 1 | 0.014 | -0.041 | -0.099 | -0.073 | -0.082 |
| MEL VAR | -0.064 | -0.039 | 0.040 | 0.014 | 1 | -0.040 | 0.238 | -0.044 | -0.091 |
| MON CON | 0.207 | -0.034 | -0.060 | -0.041 | -0.040 | 1 | -0.101 | -0.004 | 0.195 |
| DAPH | -0.013 | -0.088 | -0.101 | -0.099 | 0.238 | -0.101 | 1 | 0.379 | -0.215 |
| COPE | 0.157 | -0.068 | -0.078 | -0.073 | -0.044 | -0.004 | 0.379 | 1 | 0.053 |
| KER | -0.079 | -0.029 | 0.071 | -0.082 | -0.091 | 0.195 | -0.215 | 0.053 | 1 |
| Chl-a | -0.128 | -0.027 | -0.107 | -0.083 | 0.404 | 0.033 | -0.039 | 0.085 | 0.156 |
| TOP | 0.078 | 0.016 | 0.011 | 0.003 | -0.132 | -0.117 | 0.291 | 0.211 | -0.286 |
| Temperature | 0.096 | -0.061 | 0.175 | 0.030 | -0.143 | 0.173 | 0.090 | 0.640 | 0.202 |
| DO | 0.071 | 0.233 | 0.100 | 0.222 | 0.153 | -0.331 | 0.148 | -0.297 | -0.576 |
| pH | -0.167 | -0.320 | 0.242 | 0.173 | 0.153 | 0.009 | -0.130 | -0.037 | -0.139 |
| TS | -0.188 | -0.310 | -0.183 | -0.144 | -0.160 | 0.353 | -0.126 | 0.081 | 0.479 |
| Mg | -0.329 | -0.280 | -0.172 | -0.175 | -0.003 | 0.133 | -0.161 | -0.017 | 0.411 |
| Ca | 0.088 | -0.001 | -0.083 | 0.032 | 0.156 | 0.508 | 0.114 | 0.167 | 0.489 |

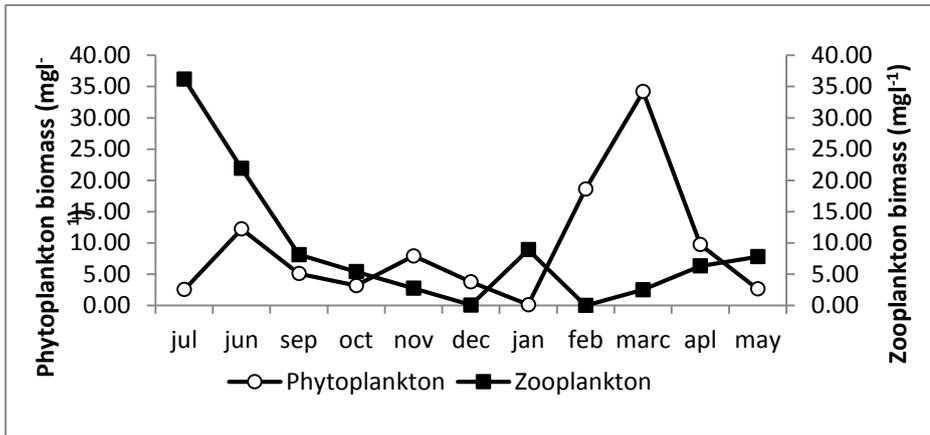


Figure 2: Seasonal variation of phytoplankton and zooplankton biomass.

Because Chlorophyll-a concentration can be easily measured in a water sample, it is a practical common way to estimate the phytoplankton biomass in the water bodies. In our study, the mean Chlorophyll-a was calculated as high value whilst phytoplankton biomass

was calculated as less value in May due to grazing pressure by zooplanktons (Fig. 3). According to biodiversity indices, species diversity increased in winter season.

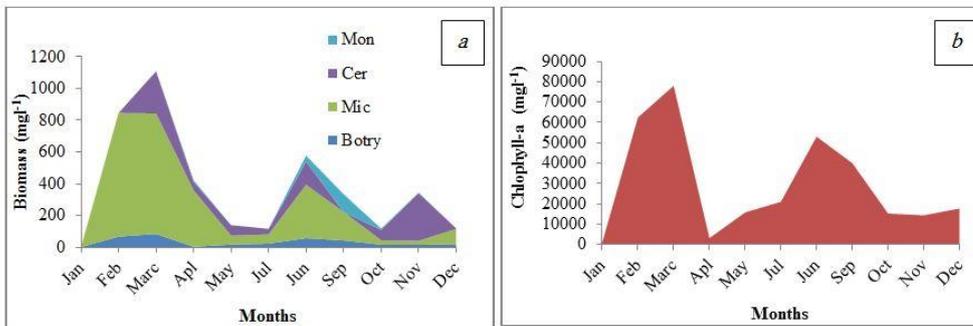


Figure 3: Seasonal variation of phytoplankton biomass (a) (Mon: *Monoraphidium contortum*, Cer: *Ceratium hirundinella*, Mic: *Microcystis aeruginosa* Botry: *Botryococcus braunii*) Seasonal variation of Chla: Chlorophyll-a (b).

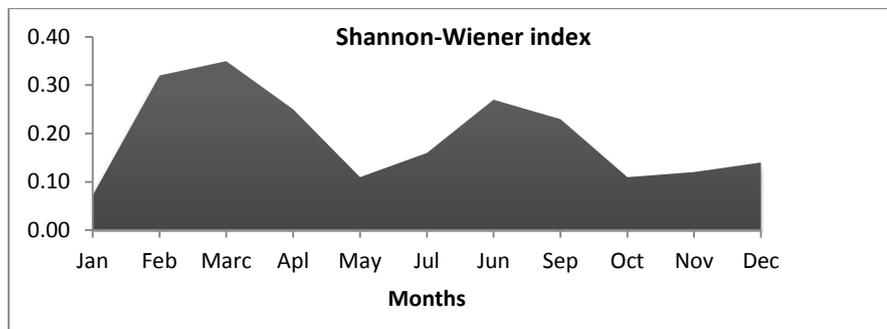


Figure 4: The variation of Shannon-Weiener diversity connects to months.

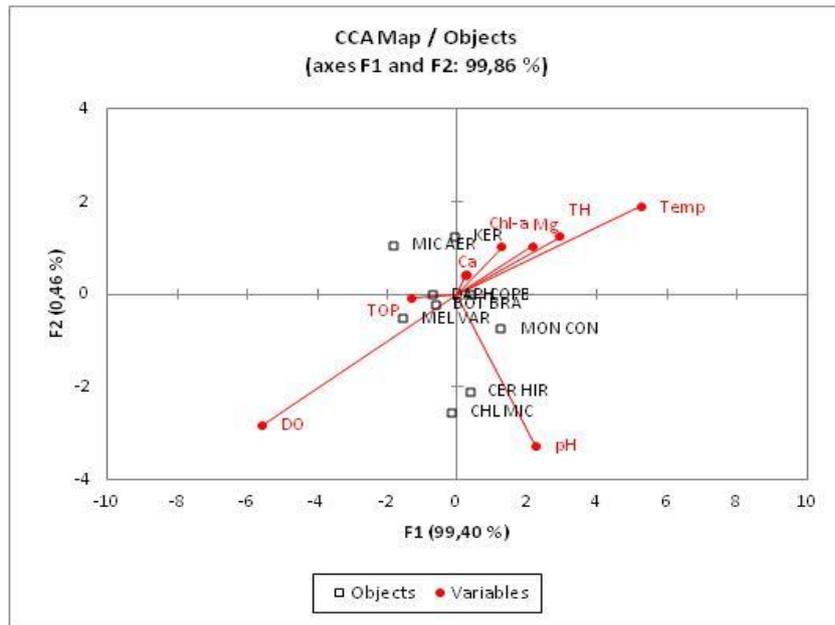


Figure 5: Species-conditional triplot based on a canonical correspondence analysis of the example phytoplankton and zooplankton data displaying 33.69% of the inertia (= weighted variance) in the abundances and 99.4 % of variance in the weighted averages and class totals of species with respect to the environmental variables. The eigenvalues of axis 1 (horizontally) and axis 2 (vertically) are 0.267 and 0.001, respectively; the eigenvalue of the axis 3 (not displayed) is 0.000. Species (triangles) are weighted averages of site scores (circles). Quantitative environmental variables are indicated by arrows. The species names are abbreviated to the part in *italic* as follows: *Ceratium hirundinella* (CER HIR), *Botryococcus braunii* (BOT BRA), *Chlamydomonas microsphearella* (CHL MIC), *Microcystis aeruginosa* (MIC AER), *Melosira varians* (MEL VAR), *Monoraphidium contortum* (MON CON), Copepoda (COPE), Daphnia (DAPH) and Keretella (KER).

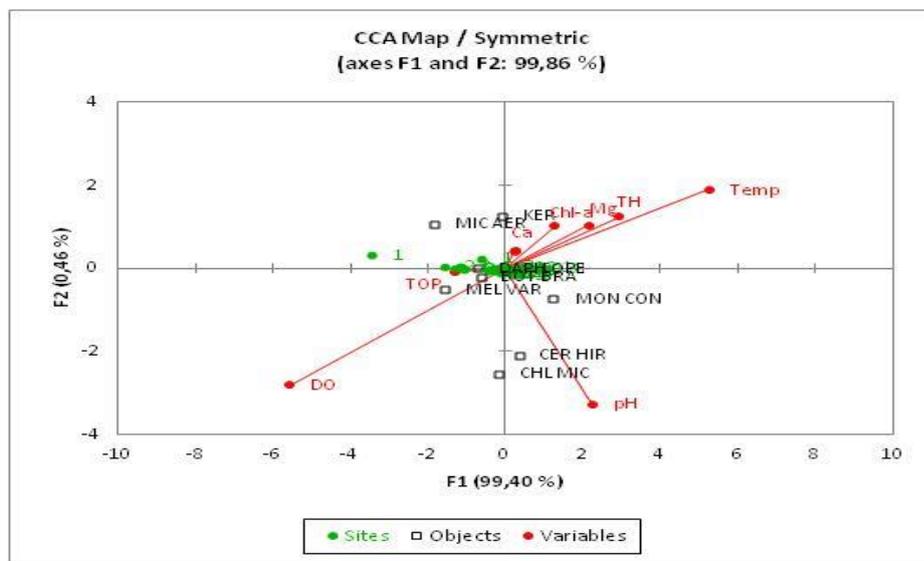


Figure 6: CCA of phytoplankton, zooplankton, environmental parameters and sites in Tortum Lake. The species names are abbreviated to the part in *italic* as follows: *Ceratium hirundinella* (CER HIR), *Botryococcus braunii* (BOT BRA), *Chlamydomonas microsphearella* (CHL MIC), *Microcystis aeruginosa* (MIC AER), *Melosira varians* (MEL VAR), *Monoraphidium contortum* (MON CON), Copepoda (COPE), Daphnia (DAPH) and Keretella (KER).

Mostly, the main contributor to phytoplankton biomass was the dinophyte *C. hirundinella*, which is regarded as an indicator of meso-eutrophic waters (Wasielawska, 2006). In this study, some indicators of meso-eutrophic waters were determined such as *C. hirundinella* and *M. aeruginosa*. However, total orthophosphate concentration, total hardness and Ca hardness were found to be lower than the values of meso-eutrophic lakes (Table 3, Fig. 3).

Interaction between phytoplankton biomass and zooplankton biomass were found as negative correlation and statistically significant ($r=-0.099$, $p<0.05$) (Table 3). There was negative correlation between *C. hirundinella* and *Daphnia*, but positive correlation between temperature, pH, dissolved oxygen, and total orthophosphorus. The increased algal biomass together with higher water temperatures allow much earlier egg development as well as higher growth rates of protozoans, rotifer and crustacean zooplankton in lakes of temperate zone (Kalff, 2001).

The similarity between months and sites according to both phytoplankton and zooplankton were estimated through a hierarchical classification analysis. This method was also useful to verify the groups obtained from the CCA (Beamuda *et al.*, 2010). All stations demonstrated similar characteristic in November, December, February and March. Sites 1 and 3 showed similarity in September, January and April, as well as site 2 and

3 in October and May because of location sites and a threat of domestic waste (Fig. 6).

We concluded that the Tortum Lake was affected by anthropogenic sources. Phytoplankton and zooplankton communities were able to flow the main seasonal changes of physical and chemical conditions in this lake. Our results demonstrated that long-term monitoring programs are needed due to protect the geological structure and eutrophication in this lake. Additionally, further research could be conducted on water bio-physico-chemical parameters and the effect of sediment characteristics on water quality.

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References

- Abdul Azis, P.K., Al-Tisan, I.A., Daili, M.A., Green, T.N., Dalvi, A.G.I. and Javeed, M.A., 2003.** Chlorophyll and plankton of the Gulf coastal waters of Saudi Arabia bordering a desalination plant. Paper presented at IDA Conference. March 2002 at Manama. Bahrain and also published in *Desalination*, 154, 291-302.
- Akbay, N., 1982.** Fitoplankton ve zooplankton hacim ve ağırlıklarının (Biomass) hesaplanmasında geometrik şekillerin kullanılması.

- Bayındırlık ve İskan Bakanlığı devlet su İşleri genel müdürlüğü İşletme bakım dairesi başkanlığı. Ankara. Turkey. 28P.
- Altuner, Z., 1982.** A research of phytoplankton and benthic algae in the Tortum Lake. PhD Thesis. Ataturk University. Institute for Natural Medicine, 80P.
- APHA, 1995** Standard methods for the examination of water and wastewater. Nineteenth ed. American Public Health Association. Washington. DC. 1216P.
- Beamuda, S.G., Diaza, M.M., Bacalab, N.B. and Pedrozoa, F.L., 2010.** Analysis of patterns of vertical and temporal distribution of phytoplankton using multifactorial analysis: Acidic Lake Caviahue. Patagonia. Argentina. *Limnology*, 40, 140-147.
- Black, P., Harrison, C., Lee, C., Marshall, B. and Wiliam, D., 2004.** Working inside the black box: Assessment for learning in the classroom. Phi delta kappan, 86(1), 8-21.
- Braak, C. J. and Smilauer, P., 2002.** CANOCO reference manual and CanoDraw for Windows user's guide: software for canonical community ordination (version 4.5). www.canoco.com.
- Braak C.J.E. and Verdonschot, P.E.M., 1995.** Canonical correspondence analysis and related multivariate methods in aquatic ecology. *Aquatic Sciences*, 57, 1015-1621.995 1015-1621.
- Bronmark C.B. and Hansson, L.A., 2005.** The biology of lakes and ponds. Second edition. Oxford University Press. 284P.
- Cramer J., 1991.** Bibliotheca phycologia. Desmides de madagascar (Chlorophyta. Zygothryxaceae) par P. Bourrelly and A. Coute. Berlin Stuttgart. 346P.
- Dantas E.W., Bittencourt-Oliveira, M.C. and Moura, A.N., 2012.** Dynamics of phytoplankton associations in three reservoirs in northeastern Brazil assessed using Reynolds' theory. *Limnology*, 42, 72-80.
- Demir, A.N., Fakioglu, Ö. and Dural, B., 2013.** Phytoplankton functional groups provide a quality assessment method by the Q assemblage index in Lake Mogan (Turkey). *Turkish Journal of Botany*, 37, 1-11.
- Dussart, B.H., 1969.** Les copepodes des eaux continentales d'Europe occidentale. Tome 11: Cyclopoïdes et biologie. N. Boubee and Cie. Paris. 292P.
- Gurbuz, H., Kivrak, E. and Soyupak, S., 2004.** Phytoplankton community structure in a high mountain reservoir. Kuzgun Reservoir. Turkey. *J Fresh Eco*, 19, 651-655.
- Hill, M.O., 1973.** Diversity ve evenness: A unifying notation and its consequences. *Ecology*, 54(2), 427-432.

- Hillebrand, H., Dürselen, C.D., Kirschtel Pollingher, D.U. and Zohary, T., 1999.** Biovolume calculation for pelagic and benthic microalgae. *Journal of Phycology*, 35, 403-424.
- James, B.K. and Adejare, L.I., 2010.** Nutrients and phytoplankton production dynamics of a tropical harbor in relation to water quality indices. *Journal of American Science*, 6(9), 261-275.
- John, P.M., Whitton, B.A. and Brook, A.J., 2002.** The freshwater algal flora of the British Isles. Cambridge Univ. Press. Cambridge. 498P.
- Kalff, J., 2001.** Limnology. Prentice Hall upper Saddle River, New Jersey. 07458 United States of America. 523P.
- Kelly MG., 1997.** Use of Trophic Diatom Index to Monitor Eutrophication in Rivers. Water Research. pp. 236-242.
- Kivrak, E., 2006.** Seasonal and long term changes of the phytoplankton in the Lake Tortum in relation to environmental factors. Erzurum. Turk Bio. Bratislava. 61, 339-345.
- Krebs, C.J., 1998.** Ecological methodology, 2nd edition. Benjamin/Cummings, Menlo Park, California. 624P. ISBN-13: 9780321021731.
- Krieger, W.V., 1932.** Die desmidiaceen der deutschen limnologischen sunda-expedition. *Archiv für Hydrobiologie Supplement*, 11, 129-230.
- Kwak, T.J. and Peterson, J.T., 2007.** Community indices, parameters, and comparisons. Pages 677-763 in C.S. Guy and M.L. Brown, Editors. Analysis and interpretation of freshwater fisheries data. American Fisheries Society, Bethesda, Maryland. ISBN-13: 978-1-888569-77-3.
- Offem, B.O., Ayotunde, E.O., Ikpi, G.U., Ada, F.B. and ve Ochang, S.N., 2011.** Plankton-based assessment of the trophic state of three tropical lakes. *Journal of Environmental Protection*, 2, 304-315.
- Orhan, T. and Karahan, F., 2010.** Uzundere İlçesi ve yakın çevresinin ekoturizm potansiyelinin değerlendirilmesi artvin çoruh Üniversitesi Orman Fakültesi Dergisi. Turkey. 11, 27-42.
- Padisak, J., Borics G., Feher, G., Grigorszky, I., Schmidt, A.O. and Zambone-Doma, Z., 2003.** Dominant species and frequency of equilibrium phases in late summer phytoplankton assemblages in Hungarian small shallow lakes. *Hydro*, 502, 157-168.
- Queimalinos, C.P., Modenutti, B.E. and Balseiro, E.G., 1998.** Phytoplankton responses to experimental enhancement of grazing pressure and nutrient recycling in a small Andean lake. *Freshwater Biology*, 40, 41-49.
- Round, F.D., 1953.** An investigation of two benthic algal communities in

- Malham Tarn. York. *Journal of Ecology*, 41, 174-197.
- Scheffer, M., Reinaldi, S., Huisman, J. and Weissing, F.J., 2003.** Why plankton communities have no equilibrium: solutions to the paradox. *Hydrobiologia*, 491, 9- 18.
- Segers, H., 1993.** Guides to the identification of the microinvertebrates of the continental waters of the world. Rotifera. Vol. 2. The Lecanidae (Monogononta). SPB Academic Publishing. 226P.
- Smimov, N., 1974.** Fauna of USSR. Crustacea. 2. Chydoridae. I.P.S.T. Jerusalem. 644P.
- Sommer, U., Sommer, F., Santer, B., Zöllner, E., Jürgens, K., Jamieson, K. and Gocke, K., 2003.** Daphnia versus copepod impact on summer phytoplankton: functional compensation at both trophic levels. *Oecologia*, 135, 639-647.
- Sun, J. and Liu, D., 2003.** Geometric models for calculating cell biovolume and surface area for phytoplankton. *Journal of Phytoplankton Research*, 25, 1331-1346.
- Tanyolac J., 2009.** Limnology (freshwater science). Hatiboğlu Press. Ankara. Turkey. 290P.
- Turkmen, G. and Kazanci, N., 2010.** Applications of various biodiversity indices to benthic macroinvertebrate assemblages in streams of a national park in Turkey. Review of Hydrobiology, 3(2), 111-125.
- Utermohl, H., 1958.** Zur vervollkommnung deer quantitativen phytoplankton-methodik. *Mitteilungen der Internationale Vereinigung für Theoretische und Angewandte Limnologie*. 5, 567-596.
- Wasielewska, E.S., 2006.** Trophic status of lake water evaluated using phytoplankton community structure-change after two decades. *Polish Journal of Environmental Studies*, 15, 139-144.
- Wetzel, R.G., 2001.** Limnology. Saunders Company. London. 743P.