Original Article

A Molecular and Serological Study on Visceral Leishmaniasis in Asymptomatic Stray Dogs in Mashhad, Iran

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ABSTRACT

Zoonotic visceral leishmaniasis is caused by *Leishmania infantum* (*L. infantum*), and its major reservoir hosts are domestic dogs, most of which are asymptomatic. This study aimed to detect *L. infantum* spp. in asymptomatic stray dogs by molecular and serological methods in Mashhad, Iran, during 2011-12. In this study, 94 asymptomatic stray dogs were randomly selected and their blood samples were collected for indirect fluorescent antibody testing. Furthermore, tissue samples from all the *L. infantum* seropositive stray dogs were examined using semi-nested polymerase chain reaction (PCR). According to the results, 11.7 %(11/94) of the dogs were *L. infantum* seropositive. The PCR positivity rate of *L. infantum* was 63.6% (7/11) in at least one of the collected specimens of the seropositive dogs. The *L. infantum* kinetoplast DNA (kDNA) was detected in the liver of 36% (4/11), the spleen of 27% (3/11), and the skin of 54.5 %(6/11) of the stray dogs. In this study, based on the molecular and serological examinations, visceral leishmaniasis infection among the stray dogs in Mashhad was confirmed.

Keywords: Leishmania infantum, Stray dogs, Mashhad

Analyse sérologique et moléculaire de laleishmaniose viscérale chez les chiens errants asymptomatiquesde la ville de Mechhed

Résumé: Les chiens errants ont été identifiés comme la source principale dela leishmaniose viscérale provoquée par l'espèce *Leishmania infantum*. Un nombre important de chiens infectés ne présente aucun signe clinique et sont considérés comme des vecteurs potentiels de cette maladie. Cette étude a pour objectif d'évaluer par le biais d'examens sérologiques et moléculaires, le taux d'infection par la leishmaniose viscérale chez les chiens errants de la ville de Mechhed . Entre 2011 et 2013, 94 chiens ont été sélectionnés au hasard et des échantillons de sang ont été prélevés pour mener destests d'immunofluorescence indirecte. De plus, une analyse moléculaire par seminested-PCR a été effectuée sur des échantillons de tissus provenant des chiens errants séropositifs.Ces analyses ont révélés que 11 chiens (11,7%) présentaient un sérum positif contre le*leishmania infantum*.La méthode seminested-PCRP a permi de confirmer l'infection par le *Leishmania infantum* a été détectée dans 36% des tissus hépatiques, 27% des tissus de rate et 54,5% des tissus de peau prélevés à partirdes chiens errants séropositifs. Nos analyses sérologiques et moléculaires ont donc confirmés la présence de l'infection au *Leishmania Infantum* chez les chiens errants de la ville de Mechhed ne présentant aucun symptômeapparent. **Mots-clés:** Leishmania Infantum, chien errant, ville de Mechhed

INTRODUCTION

Visceral leishmaniasis (VL) is known as a zoonotic and anthroponotic disease, and dogs and humans are the reservoirs of the infection (Baneth, 2005). L. infantum and L. chagasi are the main causes of zoonotic VL, while anthroponitic infection is caused by L. donovani in India and East Africa. In the Middle East, zoonotic VL is caused by L. infantum, and domestic dogs are the major reservoir hosts (Baneth, 2005). The prevalence of canine leishmaniasis varies in endemic areas (between 10% and 70% in the Mediterranean region) and is assumed to be hinged upon the climate (Mancianti et al., 1986; Kirmse et al., 1987). Therefore, the main reservoirs of VL caused by L. infantum in Iran are dogs and wild carnivores (Nadim et al., 1978; Hamidi et al., 1982; Bokai et al., 1998; Mohebali et al., 2001). Canine leishmaniasis is a chronic disease clinically characterized bv lymphadenopathy, cutaneous manifestations, weight loss, and anemia. In addition, many infected dogs are asymptomatic and are reservoir hosts of the disease in endemic areas (Dantas-Torres, 2007; Moshfe et al., 2009; Hosseininejad et al., 2012). The molecular methods have high sensitivity and specificity to discriminate between various pathogens in humans and animals. The polymerase chain reaction (PCR) assay significantly improved the diagnosis of VL in men and animals (Lachaud et al., 2002). This study sought to investigate zoonotic VL in asymptomatic stray dogs by indirect immunofluorescent antibody test (IFAT) and semi-nested PCR.

MATERIALS AND METHODS

Study area. This study was performed during February 2011-12 in Mashhad, Iran. This city is located at 36.20° latitude and 59.35° east longitude. The mean annual temperature and precipitation are $26.7 \, ^{\circ}$ C and 255.7 mm, respectively. In Mashhad, summers are typically hot and dry and sometimes the temperature exceeds $35 \, ^{\circ}$ C (95 $^{\circ}$ F), and winters are typically cool to

cold and somewhat damper, with overnight lows routinely dropping below freezing.

Sampling. In this study, asymtomatic stray dogs were randomly selected during a zoonosis control program performed by Mashhad Municipality from February 2011-12. The blood samples were collected from jugular vein and kept in plain tubes. The blood tubes were transported to laboratory under cool condition. The blood clots were centrifuged for 5-10 min at $800 \times g$, the sera were stored at -20 °C prior to the serological examination. After a euthanasia procedure, tissue samples were collected from the spleen, liver, and skin of the stray dogs and stored at -20 °C for DNA extraction.

The Indirect Immunofluorescent Antibody Test (IFAT). In the present study, IFAT and L. infantum (MegaCor Co., Austria MegaScreen antigen FLUOLEISH) were used as the tools for serodiagnosis. For each reaction, 10 µl of a serum dilution (1:50) in phosphate-buffered saline (PBS) were added over the slide holes. The slides were incubated at 37°C and humidity of 70% for 30 min and rinsed with PBS for 5 min. The conjugate was diluted to 1:200 in PBS with 0.025% Evans Blue, and then 15 µl of this solution was placed over the slide holes. The incubation and washing steps were repeated once again as outlined above. The slides were mounted in buffered glycerin covered with a coverslip and read under an Olympus BX-FLA fluorescence microscope equipped with a 100W Mercury Apo lamp with $400 \times$ magnification. The IFAT result was considered as positive if the dilution of serum gave an evident yellow-green fluorescent signal upon microscopic examination.

DNA extraction. DNA extraction from the obtained samples was carried out by DNA-plus Extraction Kit (CinnaGen Co., Iran) according to the manufacturer's instructions. The extracted DNA was stored at -20 °C for the detection of *Leishmania* kDNA detection.

PCR assay. As described by Aransay et al. in 2000, semi-nested PCR was employed for the amplification of the variable regions of the minicircle kDNA (slightly modified). Negative (tube without DNA) and positive

(DNA from *Leishmania*) controls were performed for each experiment. After amplification

, the PCR products were visualized on a 2% agarose gel electrophoresis stained with ethidium bromide under ultraviolet light.

Ethical consideration. Study protocols and methodologies were revised and approved by the Ethics Committee of Ferdowsi University of Mashhad, Mashhad, Iran.

 Table 1. The results of seminested-polymerase

 chain reaction in the 11 seropositive stray dogs

No —	Semi nested PCR		
	Skin	Spleen	Liver
1	-	+	-
2	-	-	-
3	+	-	+
4	+	-	+
5	-	-	-
6	+	+	+
7	-	-	-
8	+	+	-
9	+	-	-
10	+	+	-
11	-	-	-
Total	6 (54.5%)	4 (36%)	3 (27%)

RESULTS

Out of the 94 serum samples, 11.7 (11/94) of them were positive for *Leishmania*-specific antibodies with the titer of 1:50. The PCR positivity rate was 63.6 (7/11) in the tissue samples of the stray dogs. The kDNA of *L. infantum* was detected in 27% (3/11) of the liver, 36% (4/11) of the spleen, and 54% (6/11) of skin samples in asymptomatic stray dogs (Table 1). According to the results, four seropositive dogs were PCR negative.

DISCUSSION

Given the results of the present study, 11.7% of the serum samples collected from the asymptomatic stray dogs were positive for antibodies against *L. infantum*. In Iran, the sero-prevalence of canine leishmaniasis was reported to be 18.2% in Ardabil, 12.2% in Chaharmahal-and-Bakhtiari and Qom, and 4.4% in Khuzestan and Bushehr provinces by using direct

agglutination test (DAT) (Mohebali et al., 2005), 7.6% in Alborz Province by DAT (Haddadzade et al., 2013), 7.6% and 8% in Khorasan Razavi Province by DAT and IFAT, respectively (Adinezadeh et al., 2013; Sabzevari et al., 2013; Heidarpour et al., 2014), 8.9% in Tehran Province, 10% in Khuzestan Province, and 16% in Chaharmahal- and- Bakhtiari Province by DAT (Hosseininejad et al., 2012), and 15.4% in Kerman and Sistan-and-Baluchestan provinces by enzyme-linked immunosorbent assay (ELISA) (Mahshid et al., 2014). These serological studies confirmed that canine leishmaniasis is a common infection in different parts of Iran. The inconsistency between the results might be attributable to differences in sensitivity and specificity of the serological methods or the impact of climate change. In the present study, semi-nested PCR was used to detect the kDNA of Leishmania spp. in tissue samples of seropositive stray dogs (Aransay et al., 2000). Regarding the results of this study, semi-nested PCR is a useful method to detect Leishmania in reservoirs and vectors (Lachaud et al., 2002; Azizi et al., 2006; Rassi et al., 2007). Moreover, the kDNA of L. infantum was detected in 50% of the skin, 35.5% of the spleen, and 28.5% of the liver specimens of the dogs. According to the literature, the kDNA of Leishmania spp. was detected in the skin tissue more than the other tissues (Manna et al., 2004; de Andrade et al., 2006; Xavier et al., 2006; Solano-Gallego et al., 2009; de Queiroz et al., 2011; Reis et al., 2013). Although PCR method has high sensitivity, 36% of the seropositive dogs were PCR negative due to insufficient concentration of Leishmania kDNA in the tissue samples or the presence of PCR inhibitor. Our findings were in line with the results obtained by (de Andrade et al., 2006; Maia et al., 2009; Moshfe et al., 2009; Camargo et al., 2010; Hamarsheh et al., 2012).

According to the results of the present study, a multitude of asymptomatic stray dogs were infected by *L. infantum* and the skin samples were better than the other tissues for the detection of *Leishmania* kDNA by PCR. Therefore, the results of PCR or serological data

alone are not sufficient for the diagnosis of leishmaniasis and both of them should be used to detect this parasite.

Ethics

I hereby declare all ethical standards have been respected in preparation of the submitted article.

Conflict of Interest

The authors declare that they have no conflict of interest.

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References

- Adinezadeh, A., Eshrat Beigom, K., Mohebali, M., Shojaee, S., Rokni, M.B., Zarei, Z., *et al.*, 2013. Endoparasites of stray dogs in Mashhad, Khorasan Razavi province, Northeast Iran with special reference to zoonotic parasites.Iran J Parasitol 8, 459.
- Aransay, A.M., Scoulica, E., Tselentis, Y., 2000. Detection and identification of Leishmania DNA within naturally infected sand flies by seminested PCR on minicircle kinetoplastic DNA. Appl Environ Microbiol 66, 1933-1938.
- Azizi, K., Rassi, Y., Javadian, E., Motazedian, M., Rafizadeh, S., Yaghoobi Ershadi, M., *et al.*, 2006. Phlebotomus (Paraphlebotomus) alexandri: a probable vector of Leishmania infantum in Iran. Ann Trop Med Parasitol 100, 63-68.
- Baneth, G., Day, M., Roura, X., Shaw, S, 2005. Leishmaniosis. In : Shaw S; Day M (Eds.). Arthropodborne infectious diseases of the dog and cat pp:89-99.
- Bokai, S., Mobedi, I., Edrissian GhH, N.A., 1998. Seroepidemiological study of canine visceral leishmaniasis in Meshkin-Shahr, northwest of Iran. Arch Inst Razi 41, 48-49.
- Camargo, J., Langoni, H., Troncarelli, M., Machado, J., Lucheis, S.B., Padovani, C., 2010. Performance of IFAT, ELISA, direct parasitological examination and PCR on lymph node aspirates for canine visceral leishmaniasis

diagnosis. J Venom Anim Toxins Incl Trop Dis 16, 414-420.

- Dantas-Torres, F., 2007. The role of dogs as reservoirs of Leishmania parasites, with emphasis on Leishmania (Leishmania) infantum and Leishmania (Viannia) braziliensis. Vet Parasitol 149, 139-146.
- de Andrade, H.M., Reis, A.B., dos Santos, S.L., Volpini, Â.C., Marques, M.J., Romanha, A.J., 2006. Use of PCR– RFLP to identify Leishmania species in naturally-infected dogs. Vet Parasitol 140, 231-238.
- de Queiroz, N., da Silveira, R., de Noronha, A., Oliveira, T., Machado, R., Starke-Buzetti, W., 2011. Detection of Leishmania (L.) chagasi in canine skin. Vet Parasitol 178, 1-8.
- Haddadzade, H., Fattahi, R., Mohebali, M., Akhoundi, B., Ebrahimzade, E., 2013. Seroepidemiologcal investigation of visceral leishmaniasis in stray and owned dogs in Alborz province, Central Iran using direct agglutination test. Iran J Parasitol 8, 152.
- Hamarsheh, O., Nasereddin, A., Damaj, S., Sawalha, S., Al-Jawabreh, H., Azmi, K., *et al.*, 2012. Serological and molecular survey of Leishmania parasites in apparently healthy dogs in the West Bank, Palestine. Parasit Vectors 5, 183-183.
- Hamidi, A., Nadim, A., Edrissian, G.H., Tahvildar-Bidruni, G., Javadian, E., 1982. Visceral leishmaniasis of jackals and dogs in northern Iran. Trans R Soc Trop Med Hyg 76, 756-757.
- Heidarpour, M., Pourtaghi, M., Khoshnegah, J., 2014. Prevalence and risk factors for canine leishmaniasis in Mashhad, north east of Iran. Iran J Vet Sci Tech 4, 37-46.
- Hosseininejad, M., Mohebali, M., Hosseini, F., Karimi, S., Sharifzad, S., Akhoundi, B., 2012. Seroprevalence of canine visceral leishmaniasis in asymptomatic dogs in Iran. Iran J Vet Res 13, 54-57.
- Kirmse, P., Mahin, L., Lahrech, T., 1987. Canine leishmaniasis in Morocco with special reference to infantile kala-azar. Trans R Soc Trop Med Hyg 81, 212-213.
- Lachaud, L., Marchergui-Hammami, S., Chabbert, E., Dereure, J., Dedet, J.P., Bastien, P., 2002. Comparison of six PCR methods using peripheral blood for detection of canine visceral leishmaniasis. J Clin Microbiol 40, 210-215.
- Mahshid, M., Baharak, A., Iraj, S., Sina, K., Javad, K., Mehdi, B., 2014. Seroprevalence of canine visceral leishmaniasis in southeast of Iran. J Parasit Dis 38, 218-222.

- Maia, C., Ramada, J., Cristóvão, J.M., Gonçalves, L., Campino, L., 2009. Diagnosis of canine leishmaniasis: conventional and molecular techniques using different tissues. The Veterinary Journal 179, 142-144.
- Mancianti, F., Gradoni, L., Gramiccia, M., Pieri, S., Marconcini, A., 1986. Canine leishmaniasis in the Isle of Elba, Italy. Trop. Med. Parasitol 37, 110-112.
- Manna, L., Vitale, F., Reale, S., Caracappa, S., Pavone, L.M., Della Morte, R., *et al.*, 2004. Comparison of different tissue sampling for PCR-based diagnosis and follow-up of canine visceral leishmaniosis. Vet Parasitol 125, 251-262.
- Mohebali, M., Hajjaran, H., Hamzavi, Y., Mobedi, I., Arshi, S., Zarei, Z., *et al.*, 2005. Epidemiological aspects of canine visceral leishmaniosis in the Islamic Republic of Iran. Vet Parasitol 129, 243-251.
- Mohebali, M., Hamzavi, Y., Fallah, E., Zarei, Z., 2001. Study of canine visceral leishmaniasis in some parts of Iran and its health importance. Tehran Univ Vet Fac J 56, 55-59.
- Moshfe, A., Mohebali, M., Edrissian, G., Zarei, Z., Akhoundi, B., Kazemi, B., *et al.*, 2009. Canine visceral leishmaniasis: asymptomatic infected dogs as a source of L. infantum infection. Acta Trop 112, 101-105.
- Nadim, A., Navid-Hamidid, A., Javadian, E., Bidruni, G.T., Amini, H., 1978. Present status of kala-azar in Iran. Am J Trop Med Hyg 27, 25-28.

- Rassi, Y., Azizi, K., Motazedian, M., Javadian, E., Rafizadeh, S., Fakhar, M., *et al.*, 2007. The seminested PCR based detection of Leishmania infantum infection in asymptomatic dogs in a new endemic focus of visceral leishmaniasis in Iran. Iran J Arthropod Borne Dis 1, 38-42.
- Reis, L.E.S., Coura-Vital, W., Roatt, B.M., Bouillet, L.É.M., Ker, H.G., de Brito, R.C.F., *et al.*, 2013. Molecular diagnosis of canine visceral leishmaniasis: a comparative study of three methods using skin and spleen from dogs with natural Leishmania infantum infection. Vet Parasitol 197, 498-503.
- Sabzevari, S., Razmi, G.R., Naghibi, A., Khoshnegah, J., 2013. A serological study of Leishmania infantum in dogs of Khorasan Razavi province, Iran. J Parasit Dis 37, 189-191.
- Solano-Gallego, L., Koutinas, A., Miró, G., Cardoso, L., Pennisi, M., Ferrer, L., *et al.*, 2009. Directions for the diagnosis, clinical staging, treatment and prevention of canine leishmaniosis. Vet Parasitol 165, 1-18.
- Xavier, S.C., de Andrade, H.M., Monte, S.J.H., Chiarelli, I.M., Lima, W.G., Michalick, M.S.M., *et al.*, 2006. Comparison of paraffin-embedded skin biopsies from different anatomical regions as sampling methods for detection of Leishmania infection in dogs using histological, immunohistochemical and PCR methods. BMC Vet Res 2, 17.