# A COMPARATIVE CYTOLOGICAL STUDY IN THE D GENOME-BEARING SPECIES OF TRITICUM-AEGILOPS COMPLEX 

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The wild D genome-bearing Aegilops L. species. are considered to be sources of useful alleles which can be used in bread wheat improvement. We examined the karyotype asymmetry of the D genome-bearing species and analyzed the data to look for their evolutionary correlations. A total number of eleven accessions belonging to seven D genome species and Triticum aestivum L. were used for this study. The observations of the analyzed factors showed that Ae. vavilovii (Zhuk.) Chen. posses the most symmetric karyotypes and Ae. cylindrica Host. the most asymmetric ones. Both species Ae. crassa Boiss. and Ae. vavilovii carrying a common genomic formula (XD) showed a similar karyotype asymmetry. The results of this study compared with the literature showed that the degree of karyotype asymmetry of the D genome has a correlation with the time of divergence from its ancestral progenitors. Our results suggest that the karyotype asymmetry analysis can be a useful tool to have an overall view over the genomic relationships and modifications and also a good measure to estimate the relative date of the origin of allopolyploidy of the D genome cluster.

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## Introduction

Cytological and molecular studies in a group of related plant taxa allow biosystematists and plant breeders to understand the evolution and exploit the diversity of gene pools. As Kihara (1947) has noted, "the history of
the earth is recorded in the layers of its crust. The history of all organisms is inscribed in their chromosomes" understanding the characteristics of the plant cell nucleus is critical to learning about the genome, its behavior, modulation and generation of
biodiversity. This information is of value for both fundamental reasons and applications in plant breeding, Table 1. List of accessions of the D genome-bearing species of Aegilops-Triticum species used in this study and their locations.

| Species | 2 n | Ploidy <br> level | Genome <br> type | Location |
| :--- | :---: | :---: | :---: | :--- |
| Ae. tauschiii | 14 |  |  |  |
| subsp. tauschii | 14 | 2 x | D | 1, Iran, Azerbaijan, Khoy |
| subsp. strangulata | 28 | 4 x | XD | 15700, Iran, Fasa |
| Ae. crassa | 28 | 4 x | XD | 201, Iran, Yasooj |
| var. crassa | 28 | 4 x | XD | 15702, Iran, Shiraz |
| var. crassa |  |  |  |  |
| var. macranthera |  |  |  |  |
| Ae. cylindrica | 28 | 4 x | DC | 470, Iran, Yasooj |
| var. prokhanovii | 28 | 4 x | DC | 513, Iran, Jolfa |
| var. cylindrica | 28 | 4 x | DN | 2270004, John Innes Centre |
| Ae. ventricosa | 42 | 6 x | XDS | 226000, John Innes Centre |
| Ae. vavilovii | 42 | 6 x | XDU | 15649, Iran, Lordegan |
| Ae. juvenalis | 42 | 6 x | ABD | 90, Iran, Lorestan, Noorabad |
| Triticum aestivum |  |  |  |  |

ecology, phylogeography and genetics (Bennett, 1982 \& 1984).

As Waines and Barnhart (1992) have pointed out the success of beard wheat (Triticum aestivum L., $2 \mathrm{n}=6 \mathrm{x}$ $=42$; AABBDD ) and bread-making quality are largely affected by the D genome involved in its genome. The wild pool of the D genome distributed among Aegilops spp. are sources of disease resistance and other useful alleles that can be incorporated into the cultivated wheat (Waines \& Barnhart, 1992). Thus, the cytological and molecular studies in this group are of brilliant importance.

The D genome-bearing species of the genus Aegilops have been classified at the sectional level into Cylindropyrum and Vertebrata sections, both possessing a D genome component derived from their diploid progenitor Ae. tauschii Coss. (DD) (McFadden and Sears, 1946; Wan et al., 2000). The above genomes are existed at three ploidy levels: diploid (Ae. tauschii), tetraploid (Ae. cylindrica Host., Ae. ventricosa Tausch. and the tetraploid cytotype of Ae. crassa Boiss.), and hexaploid (Ae. vavilovii (Zhuk.) Chen., Ae. juvenalis (Thell.) Eig. and the hexaploid cytotype of Ae. crassa) all founded on a basic chromosome number of seven (Slageren, 1994).

Although the D genome-bearing species have been the matter of intensive and long studies (Badaeva et al., 2002; Rayburn \& Gill, 1987; Kimber \& Zhao, 1983; Zhao \& Kimber, 1984; Cunado et al., 1996; Saeidi et al., 2006, 2008; Sheidai et al., 2002), however a numerical and comparative karyotype analysis in this group seems to be still lacking.

This study is aimed to characterize the cytological and karyotypic details of Aegilops species-bearing the D genome and $T$. aestivum in order to construct a series of numerical parameters and a comparative cytological model for this group of species.

## Materials and methods <br> Plant materials

The karyotype analysis was performed on ten accessions belonging to six Aegilops species of the D genome cluster and one from T. aestivum. Nine accessions belonging to Ae. tauschii, Ae. crassa, Ae. cylindrica, Ae. juvenalis and T. aestivum were collected from Iran and two accessions of Ae. ventricosa and Ae. vavilovii were received from John Innes Center (Norwich, England) (Table 1).

## Chromosome spread preparation

Somatic chromosomes of meristematic root tip cells were studied from germinating seeds using Agayev (1996) method. Briefly, pretreatment was performed in saturated solution of monobromonaphthalene, washed in distilled water for 10 min , fixed in chromic acidformaldehyde mixture (1/1) at about $4{ }^{\circ} \mathrm{C}$ for 24 h , washed under tap water for 3 h . Then the materials were transferred into $70 \%$ ethanol solution and kept refrigerated till staining. For staining, the materials were transferred into distilled water for about 5-6 min and treated with 1 N NaOH at $60^{\circ} \mathrm{C}$ for 10 min , washed in distilled water thoroughly for 30 min then stained in aceto-iron-hematoxylin at $30^{\circ} \mathrm{C}$ for 24 h , washed in distilled water for at least 30 min , macerated for $10-15$ $\min$ in cellulase-pektinase enzyme solution at $37{ }^{\circ} \mathrm{C}$.

Table 2. Parameters and their descriptions used for the measurements of this study.

|  | Parameters | Descriptions |
| :---: | :--- | :--- |
| 1 | Total haploid chromosome length <br> (TCL) | Total Chromosome length of the haploid complement |
| 2 | Mean Chromosome Length <br> (MCL) | Mean Chromosome length of the haploid complement |
| 3 | Total Form percent (TF\%; <br> Huziwara 1962) | Ratio between the shortest arms of the chromosomes and their total <br> length; the TF\% value is considered to be close to $50 \%$ in most <br> symmetric karyotypes and less than $50 \%$ based on the degree of <br> asymmetry |
| 4 | R (Siljak-Yakovlev 1986) | Ratio between the longest and the shortest arms of the chromosomes |
| 5 | Asymmetry index (AsI\%; <br> Arano \& Saito 1980) | $100 \times \sum$ L/ $/$ TCL; where l is long arms in chromosome set and TCL <br> is total chromosome length in chromosome set |
| 6 | Karyotype formulae (Levan et al. <br> 1964) | According to their arm ratios (long/short) designated by the position <br> of the centromere: 1 (metacentric; M), 1-1.7 (metacentric; m), 1.7-3 <br> (submetacentric; sm), 3-7 (subtelocentric; st), 7-39 (telocentric; t)]. |
| 7 | intrachromosomal asymmetry <br> index (A1; Romero Zarko 1986) | = $1-\left[\sum(b / B) / n\right] ;$ where $b$ and $B$ are the mean length of short and <br> long arms of each pair of homologues respectively, $n$ is the number <br> of homologues, The value of A1 is considered to be close to 0 if all <br> chromosomes are metacentric and near to one if all chromosomes <br> are telocentric. |
| 8 | interchromosomal asymmetry <br> index (A2; Romero Zarko 1986) | $=s / x ;$ where $s$ and $x$ are standard deviation and the mean length of <br> the chromosomes |

Table 3. Karyotype features of the D genome-bearing species of Aegilops-Triticum ( $\mathrm{n}=$ chromosome number, TL = Total haploid chromatin length, $\mathrm{MCL}=$ Mean Chromosome Length, $\mathrm{SE}=$ Standard Error, $\mathrm{TF} \%=$ Total form percent, $\mathrm{R}=$ ratio between the longest and the shortest arms of the chromosomes, AsI $\%$ = Asymetry index, Karyotype formulae, $\mathrm{A} 1=$ intrachromosomal and $\mathrm{A} 2=$ interchromosomal asymmetry index).

| Species | 2 n | TCL | MCL $\pm$ SE | TF\% | R | AsI\% | Karyotype formulae | A1 | A2 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ae. tauschii | 14 | 56.546 | $8.078 \pm 1.330$ | 40.17 | 1.51 | 58.29 | $5 \mathrm{~m}+2 \mathrm{sm}$ | 0.302 | 0.164 |
| Ae. crassa | 28 | 110.392 | $7.885 \pm 1.107$ | 40.06 | 1.57 | 57.74 | $11 \mathrm{~m}+3 \mathrm{sm}$ | 0.294 | 0.140 |
| Ae. cylindrica | 28 | 94.393 | $6.742 \pm 0.817$ | 31.58 | 1.48 | 67.62 | $1 \mathrm{M}+6 \mathrm{~m}+3 \mathrm{sm}+2 \mathrm{st}$ <br> +2 t | 0.500 | 0.121 |
| Ae. ventricosa | 28 | 99.528 | $7.109 \pm 1.063$ | 35.92 | 1.81 | 63.36 | $1 \mathrm{M}+6 \mathrm{~m}+5 \mathrm{sm}+2 \mathrm{st}$ | 0.397 | 0.149 |
| Ae. vavilovii | 42 | 147.7 | $7.033 \pm 1.462$ | 41.81 | 2.07 | 57.24 | $2 \mathrm{M}+13 \mathrm{~m}+6 \mathrm{sm}$ | 0.260 | 0.207 |
| Ae. juvenalis | 42 | 165.883 | $7.899 \pm 1.172$ | 37.96 | 1.66 | 60.92 | $1 \mathrm{M}+11 \mathrm{~m}+8 \mathrm{sm}+1 \mathrm{st}$ | 0.363 | 0.148 |
| T. aestivum | 42 | 207.584 | $9.884 \pm 1.457$ | 38.11 | 1.69 | 60.21 | $2 \mathrm{M}+11 \mathrm{~m}+7 \mathrm{sm}+1 \mathrm{st}$ | 0.354 | 0.147 |

The roots were gently squashed in $45 \%$ acetic acid, on a slide glass and were observed and photographed under an Olympus AX-40 light microscope.

In order to characterize the karyotypic asymmetry 510 chromosome spreads from different individuals of each accession were examined. All chromosomal sizes were measured with computer-aided program Image Tool 3.0. The parameters measured in each metaphase chromosome spread are listed in Table 2.

## Results and Discussion

The morphological observations of our chromosomal studies are shown as karyotypes in Fig. 1. The resulted karyotype formulae and their analyzed parameters are shown in Tables 3 \& 4 .

The results showed that while sub-telocentric (st) together with telocentric ( t ) chromosome types appeared at low frequency ( $7.08 \%$ ), the metacentric (m) and sub-metacentric (sm) types dominated the observed karyotypes with $92.92 \%$ frequency. The only two telocentric chromosomes observed, were exclusively indicators of Ae. cylindrica (see Table 3). Our results showed that satellites are restricted only to one chromosome in: Ae. tauschii, Ae. ventricosa, Ae. cylindrica and Ae. vavilovii and two chromosomes in Ae. crassa, Ae. juvenalis and T. aestivum. No B chromosome was observed among the materials studied.

Total chromosome length (TCL) was a variable parameter in this study; which can be caused by




Fig. 1. Somatic chromosomes of the D genome-bearing species. 1. Aegilops tauschii, 2. Ae. crassa, 3. Ae. cylindrica, 4. Ae. ventricosa, 5. Ae. vavilovii, 6. Ae. juvenalis, 7. T. aestivum. Scale bar $=20 \mu \mathrm{~m}$.


Fig. 2. Scatter diagram shows the relationships between the D genome-bearing species of Aegilops-Triticum based on the intrachromosomal (A1) and interchromosomal (A2) asymmetry indices. Values of A1 and A2 are summarized in Table 3. $(1=A e$. tauschii, $2=A e$. crassa, $3=A e$. cylindrica, $4=A e$. vavilovii, $5=A e$. ventricosa, $6=A e$. juvenalis, $7=T$. aestivum).
different ploidy levels and presumably different genomic combinations (Table 3). As it was expected, the lowest $(56.546 \mu \mathrm{~m})$ and the highest ( $207.584 \mu \mathrm{~m}$ ) TCL values were found in a diploid (Ae. tauschii) and a hexaploid species (T. aestivum with AABBDD formula) respectively; it must be mentioned that there is a big difference between the LT of T. aestivum ( $207.584 \mu \mathrm{~m}$ ) and the other two hexaploids i.e., (Ae. juvenalis, TCL $=165.883$ and Ae. vavilovii, TCL $=$ 147.7). Based on these results it can be concluded that generally the chromosomal length in the genus Aegilops is shorter than that of T. aestivum. The highest chromosome length variation was found in Ae. vavilovii (XXDDSS) [SE (standard error) of $\mathrm{MCL}=1.462 \mu \mathrm{~m}$ ], and the lowest chromosome length variation was scored in Ae. cylindrica ( SE of $\mathrm{MCL}=0.817 \mu \mathrm{~m}$ ) (Table 3). The ratio between the longest and the shortest arms (R) ranges from 1.48 in Ae. cylindrica to 2.07 in Ae. vavilovii (Table 3). Asymmetry Index (AsI\%) ranged from the minimum 57.24 in $A e$. vavilovii to the maximum 67.62 in Ae. cylindrica (Table 3).

Intra-specific length variation of short and long chromosome arms among the species- bearing the D genome showed that the coefficient of variability of short arms length was higher than that of the long arms length (Table 5); it can be interpreted that the TCL changes of the chromosomes is mainly influenced by the short arms rather than the long arms.

The degree of karyotype asymmetry as indicated by TF\% values ranged between 31.58\% (Ae. cylindrica) and $41.81 \%$ (Ae. vavilovii) (Table 3).

Three groups of the D genome-bearing species in the scatter diagram constructed based on A1 and A2 asymmetry indices (Fig. 2) were formed: (1) Ae. cylindrica with the most asymmetrical karyotype, (2) Ae. vavilovii with the most symmetrical karyotype and (3) a group of Ae. tauschii, Ae. crassa, Ae. ventricosa, Ae. juvenalis and T. aestivum with an intermediate place between the two latter groups. Inter- and intrachromosomal asymmetry indices (A1 and A2 in Table 3) showed that $A e$. juvenalis to be the most similar species to T. aestivum.

Based on the results of this study (the factors studied and the resulted asymmetry indices) Ae. cylindrica showed to be the most asymmetric karyotype (with the formulae of $1 \mathrm{M}+6 \mathrm{~m}+3 \mathrm{sm}+2 \mathrm{st}+2 \mathrm{t}$ ) among the species studied. Regarding the asymmetry indices observed in Ae. cylindrica it can be suggested that the karyotype asymmetry in this species is mainly affected by the place of the centromers rather than length of the chromosomes. Ae. vavilovii with the least chromosomal arm ratio variability, showed the most symmetric karyotype (with the formulae of $2 \mathrm{M}+13 \mathrm{~m}+6 \mathrm{sm}$ )
pə!pms
Table 5. Coefficient of variability for intra-specific chromosome length variation of short and long arms for the D genome-bearing species of Aegilops-Triticum
among the species studied. Regarding all the analyzed factors, a high similarity was found between $A e$. tauschii (DD) and Ae. crassa (XXDD) (see Table 3). The high karyotype asymmetry similarity, as indicated by TF $\%$ and AsI $\%$ between Ae. crassa (XXDD; TF\% = 40.06, AsI $\%=57.74$ ) and Ae. vavilovii (XXDDSS; $\mathrm{TF} \%=41.18, \mathrm{AsI} \%=57.24)$, confirms the presence of two similar genomes ( D and X ) in common between these two species. The only other species possessing D and X genomes, i.e., Ae. juvenalis (XXDDUU), showed no similarity regarding the degree of karyotype asymmetry with the above mentioned species.

As Romero Zarco (1986) has mentioned, karyotype asymmetry can be a good expression of the general morphology of karyotype in plants. According to the definition by Sharma (1990) symmetrical karyotypes are more primitive than asymmetrical ones; longer chromosomes than shorter ones; median centromers with chromosome arms of equal length are more primitive than chromosomes with arms of unequal length; low basic numbers give rise to higher ones. Considering the above notions and the results of this study, it can be suggested that the asymmetric karyotypes observed within the D genome cluster species such as Ae. cylindrica are to be young. This suggestion is in accordance with the conclusion drawn by Badaeva, et al. (2002) who used C-banding and pAs1-FISH patterns, that Ae. cylindrica is a recently originated species. Based on the results of this study, Ae. vavilovii with the most symmetric karyotype, can be considered as the oldest relative to the hexaploid Aegilops species in the D genome cluster and also $T$. aestivum. This result is in accordance with Wang, et al. (1997) who based on the study of chloroplast and mitochondrial DNA variation suggested that $A e$. juvenalis originated shortly before the other hexaploid species of the D genome cluster in Aegilops. However Badavea, et al. (2002) believed that Ae. juvenalis is the oldest hexaploid among the respective species. In the same way, Ae. crassa showed a high symmetric karyotype (nearest species to the Ae. vavilovii in karyotype asymmetry; Table 3); suggesting a high chromosomal rearrangement event following the formation of this tetraploid species. Badaeva, et al. (2002) showed high genomic modifications in this species. Compare with its parental genomes, Ae. ventricosa with an intermediate karyotype asymmetry revealed some minor genomic modifications, that is in accordance with the observations of Dubcovsky, et al. (1994), Bardsley, et al. (1999) and Badaeva, et al. (2002) who noted minor differences between the $A e$. ventricosa genomes and its progenitor species (Ae. tauschii and Ae. uniaristata Vis.).

Finally, the results of this study showed that the karyotype asymmetry analysis can be a good tool to reveal the genomic modifications and relative duration of allopolyploid formations in the D genome cluster. However, making any conclusion should be provided to considering the other genomic combinations involved.

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